

# Prevalence of faecal carriage of colistin-resistant Gram-negative rods in a university hospital in western France, 2016

Marion Saly,<sup>1,2</sup> Aurelie Jayol,<sup>1,2,3,4,5,\*</sup> Laurent Poirel,<sup>3,4,5</sup> Francis Megraud,<sup>1</sup> Patrice Nordmann<sup>3,4,5,6</sup> and Veronique Dubois<sup>1,2</sup>

## Abstract

Plasmid-mediated and chromosomally-encoded colistin resistance is increasingly being reported worldwide. We aimed to determine the prevalence of faecal carriage of colistin-resistant Gram-negative rod isolates in a university hospital in western France. From February to May 2016, rectal swabs from 653 patients hospitalized in various clinical settings were recovered and subsequently screened for colistin resistance using the SuperPolymyxin medium. Antimicrobial susceptibilities were determined according to EUCAST guidelines. Genetic detection of plasmid-mediated colistin resistance was performed by PCR. The faecal carriage with intrinsic colistin-resistant isolates was high (23 %), while the faecal carriage with Gram-negative rods showing acquired resistance was low (1.4 %). No isolate carried the plasmid-mediated *mcr-1/mcr-2* genes. It was noteworthy that none of the patients carrying isolates with acquired colistin resistance had previously received a colistin-based treatment, while these isolates were not multidrug resistant.

Plasmid-mediated *mcr-1* [1] and *mcr-2* genes [2] conferring colistin resistance in *Enterobacteriaceae* have been discovered recently in animals worldwide. Simultaneously, human infections and faecal carriage with *Enterobacteriaceae* isolates carrying the *mcr-1* gene were reported extensively during 2015 [3]. In France, a high prevalence of the plasmid-borne *mcr-1* gene has been reported among extended-spectrum  $\beta$ -lactamase (ESBL)-producing *Escherichia coli* isolates collected from the faeces of diarrheic veal calves [4]. This finding may suggest that the dissemination of *mcr-1* from animals to humans may have already occurred quite extensively.

We therefore conducted a prospective study to evaluate the prevalence of faecal carriage of acquired and intrinsic colistin-resistant Gram-negative rods among patients admitted to or hospitalized at the Pellegrin University Hospital of Bordeaux (1350 beds), which is the largest public hospital in south-west France. From February to May 2016, rectal swabs from 653 patients hospitalized in various clinical settings (emergency, intensive care unit, surgery, medical units) were screened. Colistin-resistant isolates were screened using the SuperPolymyxin medium, which contains colistin sulfate at a concentration of  $3.5 \text{ mg l}^{-1}$ , allowing the isolation of any colistin-resistant Gram-negative rods [5]. Colonies growing on this medium were identified

using a MALDI-TOF mass spectrometer (Bruker, Champs-sur-Marne, France). Isolates belonging to species that are intrinsically resistant to colistin were not further investigated. Minimum inhibitory concentrations (MICs) of colistin were determined for the other isolates using the reference broth microdilution (BMD) method. The results were interpreted according to the European Committee on Antimicrobial Susceptibility Testing (EUCAST) guidelines, 2016 [6]. All colistin-resistant isolates were subsequently screened for the *mcr-1* and *mcr-2* genes by PCR, as described previously [1, 2].

Out of the 653 rectal swabs, 150 gave isolates that are known to be intrinsically resistant species, namely *Proteus* sp. ( $n=75$ ), *Morganella* sp. ( $n=22$ ), *Providencia* sp. ( $n=12$ ), *Serratia* sp. ( $n=6$ ) and *Hafnia* sp. ( $n=35$ ) [the latter genus was recently described as being naturally resistant to colistin (A. Jayol, personal communication)].

Nine isolates with an acquired colistin trait were recovered, including three *Escherichia coli*, two *Klebsiella pneumoniae*, one *Raoultella ornithinolytica*, two *Enterobacter cloacae* and one *Stenotrophomonas maltophilia* (Table 1). The MIC values of colistin ranged from 4 to  $128 \text{ mg l}^{-1}$  for those isolates that exhibited various profiles of resistance to the other

Table 1. Colistin-resistant *Enterobacteriaceae* isolates

Species	MIC of CS (mg l <sup>-1</sup> )	Additional antibiotic resistances	ESBL	Unit
<i>E. coli</i>	4	AMX, TIC, SXT	No	Emergency
<i>E. coli</i>	4	AMX, TIC, FQ, GM	No	Surgery ICU
<i>E. coli</i>	8	AMX, C1G	No	Neurosurgery ICU
<i>K. pneumoniae</i>	64	FQ, SXT	No	Medical ICU
<i>K. pneumoniae</i>	32	AMX, TIC, C3G, C4G, FQ, FSF, FT, GM, SXT	Yes	Surgery
<i>R. ornithinolytica</i>	16	No	No	Gastroenterology
<i>E. cloacae</i>	64	AMX, TIC, C3G, C4G, FT, GM, SXT	Yes	Medical ICU
<i>E. cloacae</i>	8	FT	No	Emergency
<i>S. maltophilia</i>	128	–	No	Medical ICU

AMX, amoxicillin; C1G, first-generation cephalosporin; C3G, third-generation cephalosporin; C4G, fourth-generation cephalosporin; CS, colistin; ESBL, extended-spectrum beta-lactamase; FQ, fluoroquinolone; FSF, fosfomycin; FT, nitrofurantoin; GM, gentamicin; ICU, intensive care unit; SXT, cotrimoxazole; TIC, ticarcillin.

antibiotics (Table 1). Only two isolates (a single *K. pneumoniae* and a single *E. cloacae*) were multidrug resistant and produced an extended-spectrum  $\beta$ -lactamase. All those isolates were negative for the *mcr-1* and *mcr-2* genes. The two colistin-resistant *E. cloacae* isolates presented 'skip wells' with the BMD method, suggesting a heteroresistant phenotype. These isolates could belong to a cluster-dependent colistin-heteroresistant complex, as described recently [7].

Out of the nine patients carrying an acquired colistin-resistance trait, five were hospitalized in an intensive care unit, and two were from the community. None of the patients had previously received a colistin-based treatment, further highlighting the occurrence of colistin-resistant isolates without obvious corresponding antibiotic selection, as previously notified by Olaitan *et al* [8].

This prospective study indicates a high prevalence (23 %) of faecal carriage with intrinsic colistin-resistant Gram-negative rods, but a low prevalence (1.4 %) with isolates showing acquired resistance. No isolate carried the plasmid-mediated *mcr-1* and *mcr-2* genes, suggesting that there is currently low diffusion of these resistance determinants among human clinical samples. Similar observations were recently made during hospital- and community-based surveys in Switzerland [9]. However, single *E. coli* and *K. pneumoniae* isolates with a plasmid-borne *mcr-1* gene were isolated in 2016 from clinical infections in our hospital (A. Jayol, personal communication). Both isolates were ESBL producers and the *E. coli* one was recovered from a patient without a recent history of travel. Regular screening of plasmid-mediated colistin resistance, which is now quite easy to implement owing to the availability of a selective medium (SuperPolymyxin), may now be important to monitor and therefore possibly detect at an early stage the emergence of

colistin-resistant isolates, with the ultimate goal of preventing their further spread in France.

#### Funding information

This work was supported by internal funding from the University of Bordeaux, France, and the University of Fribourg, Switzerland.

#### Conflicts of interest

The authors declare that there are no conflicts of interest.

#### References

- Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R *et al*. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis* 2016;16:161–168.
- Xavier BB, Lammens C, Ruhel R, Kumar-Singh S, Butaye P *et al*. Identification of a novel plasmid-mediated colistin-resistance gene, *mcr-2*, in *Escherichia coli*, Belgium, June 2016. *Euro Surveill* 2016; 21.
- Skov RL, Monnet DL. Plasmid-mediated colistin resistance (*mcr-1* gene): three months later, the story unfolds. *Euro Surveill* 2016;21.
- Haenni M, Poirel L, Kieffer N, Châtre P, Saras E *et al*. Co-occurrence of extended spectrum  $\beta$  lactamase and MCR-1 encoding genes on plasmids. *Lancet Infect Dis* 2016;16:281–282.
- Nordmann P, Jayol A, Poirel L. A universal culture medium for screening polymyxin-resistant gram-negative isolates. *J Clin Microbiol* 2016;54:1395–1399.
- EUCAST. *Breakpoints tables for interpretation of MICs and zone diameters, Version 1.0*. EUCAST. 2016.
- Guérin F, Isnard C, Sinel C, Morand P, Dhalluin A *et al*. Cluster-dependent colistin hetero-resistance in *Enterobacter cloacae* complex. *J Antimicrob Chemother* 2016;71:3058–3061.
- Olaitan AO, Morand S, Rolain JM. Emergence of colistin-resistant bacteria in humans without colistin usage: a new worry and cause for vigilance. *Int J Antimicrob Agents* 2016;47:1–3.
- Liassine N, Assouvie L, Descombes MC, Tendon VD, Kieffer N *et al*. Very low prevalence of MCR-1/MCR-2 plasmid-mediated colistin resistance in urinary tract Enterobacteriaceae in Switzerland. *Int J Infect Dis* 2016;51:4–5.