







Possible incomplete penetrance of Xq28 int22h-1/int22h-2 duplication

Alexis Billes^{1,2} | Mathilde Pujalte³ | Guillaume Jedraszak^{2,4} | Daniel Amsallem⁵ |
 Elise Boudry-Labis⁶ | Odile Boute⁷ | Sonia Bouquillon⁶ |
 Elise Brischoux-Boucher⁸  | Patrick Callier^{9,10,11} | Charles Coutton^{12,13}  |
 Anne-Laude Avice Denizet⁸ | Klaus Dieterich^{14,15} | Paul Kuentz^{16,17} |
 James Lespinasse¹⁸ | Benoît Mazel¹⁹ | Gilles Morin¹ | Florence Amram¹ |
 Perrine Pennamen²⁰ | Marlène Rio^{21,22} | Juliette Piard^{8,23}  |
 Audrey Putoux^{24,25} | Mélanie Rama⁶ | Virginie Roze-Guillaume¹⁶ |
 Caroline Schluth-Bolard^{3,26,27}  | Marianne Till³ | Chloé Trouvé⁸ |
 Gaëlle Vieville²⁸ | Caroline Rooryck^{29,30}  | Damien Sanlaville^{3,26} |
 Nicolas Chatron^{3,26} 

Correspondence

Alexis Billes, CHU Amiens Picardie, Service de Génétique Clinique, Amiens, France.

Email: alex.bill@tutanota.com

Abstract

Xq28 int22h-1/int22h-2 duplication is the result of non-allelic homologous recombination between int22h-1/int22h-2 repeats separated by 0.5 Mb. It is responsible for a syndromic form of intellectual disability (ID), with recurrent infections and atopic diseases. Minor defects, nonspecific facial dysmorphic features, and overweight have also been described. Half of female carriers have been reported with ID, whereas all reported evaluated born males present mild to moderate ID, suggesting complete penetrance. We collected data on 15 families from eight university hospitals. Among them, 40 patients, 21 females (one fetus), and 19 males (two fetuses), were carriers of typical or atypical Xq28 int22h-1/int22h-2 duplication. Twenty-one individuals were considered asymptomatic (16 females and 5 males), without significantly higher rate of recurrent infections, atopia, overweight, or facial dysmorphism. Approximately 67% live-born males and 23% live-born female carriers of the typical duplication did not have obvious signs of intellectual disability, suggesting previously undescribed incomplete penetrance or low expression in certain carriers. The possibility of a second-hit or modifying factors to this possible susceptibility locus is yet to be studied but a possible observational bias should be considered in assessing such challenging X-chromosome copy number gains. Additional segregation studies should help to quantify this newly described incomplete penetrance.

For affiliations refer to page 244

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Authors. *Clinical Genetics* published by John Wiley & Sons Ltd.

KEYWORDS

CLIC2, incomplete penetrance, *RAB39B*, X-linked intellectual disability, Xq28 duplication, Xq28 int22h-1/int22h-2 duplication

1 | INTRODUCTION

X-linked intellectual disability (XLID) is a genetically heterogeneous disorder that affects 1/1000 males^{1,2} associated with more than 140 genes.³ Various X-chromosome abnormalities have been described,⁴ many inherited from a healthy heterozygous carrier mother. Pathogenic microduplications and functional disomies have long been described within the Xq28 gene-dense region.^{5,6} The presence of several low-copy repeats (LCRs) in the region predisposes it to recurrent chromosomal rearrangements by non-allelic homologous recombination (NAHR).⁷⁻¹⁰ One of the most commonly reported variants is the Xq28 duplication that includes the dosage sensitive *MECP2* gene, responsible for Lubs X-linked intellectual developmental disorder syndrome (OMIM # 300260).^{11,12}

More distally from the *MECP2* gene, El-Hattab et al. first reported a recurrent ~0.5 Mb duplication between LCRs int22h-1 and int22h-2 as being pathogenic in 2011 and then in 2015.^{13,14}

To date, 35 cases (19 males and 16 females) with typical int22h-1/int22h-2-mediated duplication have been reported in the literature.^{15,16} It is considered to be causal for syndromic XLID, with a number of inconstant distinguishing behavioral, facial, anthropometric, and immune features in males. A minority of female carriers are symptomatic, presenting with milder ID than males.¹⁷⁻²⁰

The duplication includes three disease-causing genes: *F8*, *RAB39B*, and *CLIC2*. The gene *F8* encodes coagulation factor VIII, deficient in hemophilia type A (HA) (OMIM 306700).²¹ *RAB39B* and *CLIC2* are two major candidate genes for neurodevelopmental disorders observed in cases of duplication at this locus. Indeed, deleterious *CLIC2* sequence variants have been proposed to be causative for X-linked syndromic ID in a single family,²² whereas loss-of-function variants of *RAB39B* lead to Waisman syndrome (OMIM 311510),^{23,24} a form of early onset parkinsonism with ID. However, Vanmarsenille et al.²⁰ reported an inconstant increase in *CLIC2* mRNA expression in intellectually disabled male carriers of recurrent int22h-1/int22h-2-mediated duplications and concluded that *CLIC2* “diplosensitivity” is unlikely. The same study showed increased expression of *RAB39B* in male individuals with ID and recurrent int22h-1/int22h-2-mediated duplications. It is presumed that *RAB39B* is dose sensitive. The authors concluded that the involvement of *RAB39B* in neurodevelopmental disorders in Xq28 int22h-1/int22h-2 duplication is still unclear but plausible.

Of note, healthy male carriers of atypical Xq28 duplications overlapping the int22h1-int22h2 interval have been described.²⁵⁻²⁷ Following an initial puzzling observation of an asymptomatic male carrier with the typical int22h-1/int22h-2-mediated duplication and given the males reported to carry the atypical Xq28 duplication without ID, we decided to study the penetrance of this syndrome in males.

In this retrospective study, we describe a cohort of 15 unrelated families with typical (10 families) and atypical (5 families) Xq28 int22h-1 and int22h-2 duplication.

2 | FAMILIES

Through the French Array Comparative Genomics Hybridization (CGH) network AChro-Puce, we collected data from 15 unrelated families in 8 French university hospitals (written informed consents were received from the patients). The pedigrees are presented in Figure 1. Ten families with typical Xq28 int22h-1/int22h-2-mediated duplication carriers (Families 1, 2, 3, 5, 6, 7, 8, 10, 11, 12) and five families with atypical duplication carriers (Families 4, 9, 13, 14, 15), presented in Figure 2. Here, we describe the four families in which an unaffected male carrier was identified and Family 15, containing a symptomatic male individual without ID. Other families are described in Figures 1, 2, and Table 1.

2.1 | Family 1

The proband (II-5) showed early childhood language regression, had no motor delay, and no dysmorphic features. His clinical examination was normal, except for large tonsils associated with probable recurrent infections. His metabolic blood tests were also normal. He was overweight, of large stature and had an enlarged head circumference. A familial study showed that the duplication was maternally inherited and that it was present in three of six sons. Brother II-1 was an asymptomatic carrier. He went to school and earned a vocational diploma. Brother II-3, also a carrier, required specialized education but experienced neonatal anoxia, a potential confounding factor. The asymptomatic mother (I-2) was a carrier of the typical Xq28 duplication and was unemployed. She was not tested for X-inactivation bias. The non-carrier brothers II-2, II-4, and II-6 had no particular problems or phenotype to report.

2.2 | Family 3

The proband IV-1 was a male fetus (mother III-2). Prenatal array-CGH analysis was performed at 27 weeks of gestation after identification of bilateral talipes equinovarus. This phenotype has not been reported for patients with typical Xq28 duplications and was considered to be an incidental finding. Based on reported complete penetrance in males of Xq28 duplication, the parents opted for termination of the pregnancy. For the second male fetus, IV-2 (asymptomatic mother III-4),

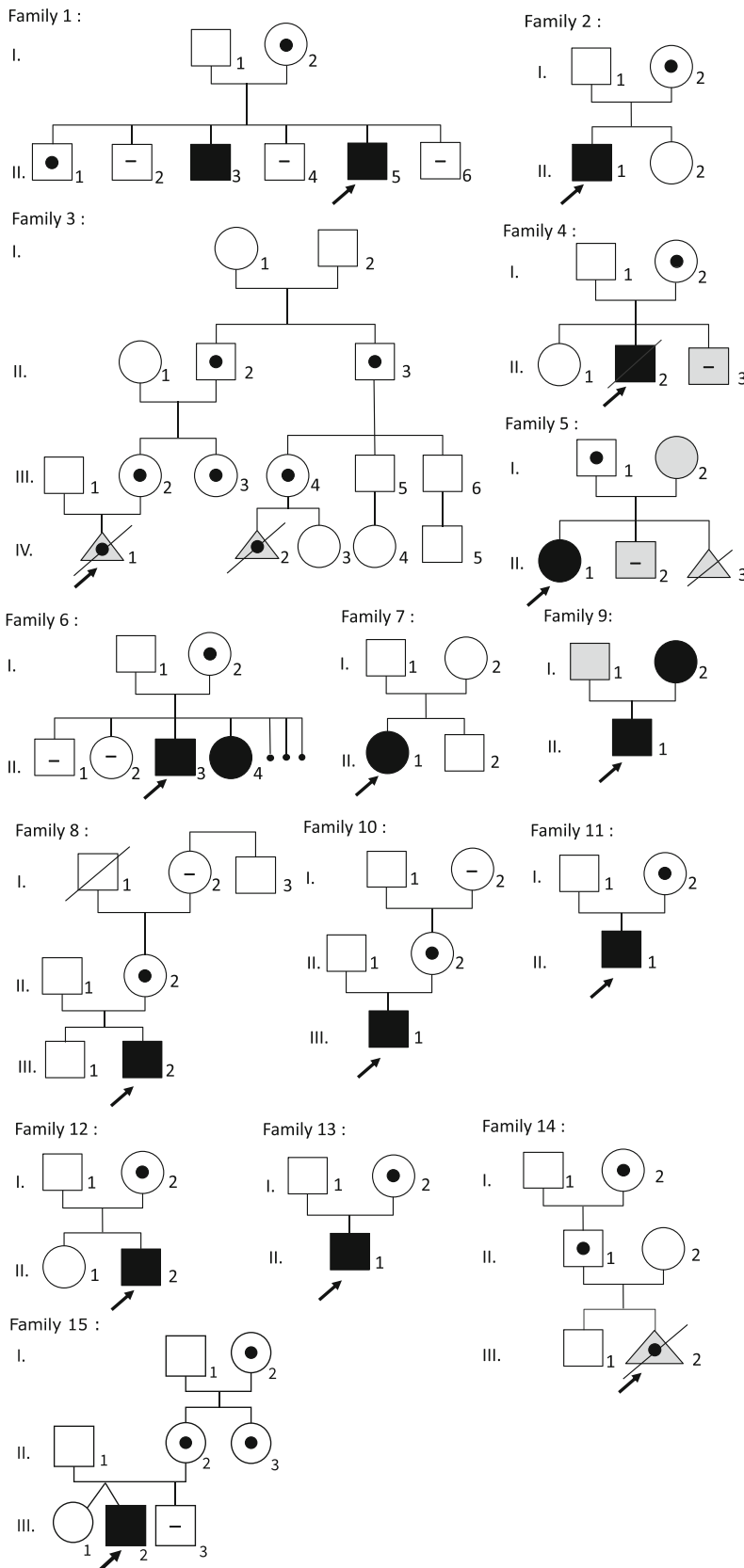


FIGURE 1 Pedigrees. The squares (males) and circles (females) are filled with black (symptomatic carrier), white (asymptomatic or status unknown), and gray (noticeable phenotype not related to Xq28 syndrome); black dot (asymptomatic carrier); minus sign (tested non-carrier). Arrows indicate the probands. The status concerning intellectual disability (ID) for Family 6 I-2 is unknown. Family 3 II-2 was not tested but was a compulsory carrier. Family 3 IV-1 had bilateral talipes equinovarus and Family IV-2 intrauterine fetal growth restriction and duplex kidneys. Family 4 II-3 had the same phenotype as his brother. Family 5 II-2 has seizures treated by sodium valproate, Family 5 II-1 and II-2 have features of fetal valproate syndrome, and Family 5 II-3 had spina bifida. Family 9 I-1 has West syndrome and mild ID. Siblings of Family 12 I.1 are not represented but there is a history of depressive disorder (sister), schizophrenia (another sister), and dyslexia (brother's niece). Family 14 III-2 had myelomeningocele. Family 15 III-2 has anterior balanic hypospadias and hemophilia due to the recurrent pathogenic intron 22 inversion of F8 (II-2, II-3, and I-2 are also carriers of this inversion); III-3 was a tested non-carrier for F8 int22 inversion and considered to be non-carrier of the familial xq28 duplication (these two elements considered to be linked).

ultrasound showed intrauterine growth retardation and duplex kidneys. Array-CGH identified the Xq28 duplication. The parents opted for termination of the pregnancy. Further familial investigations

showed that individual II-3 carried the duplication. He was a craftsman and considered to be asymptomatic. Individual II-2 could not be genetically tested for the duplication but is considered to be an

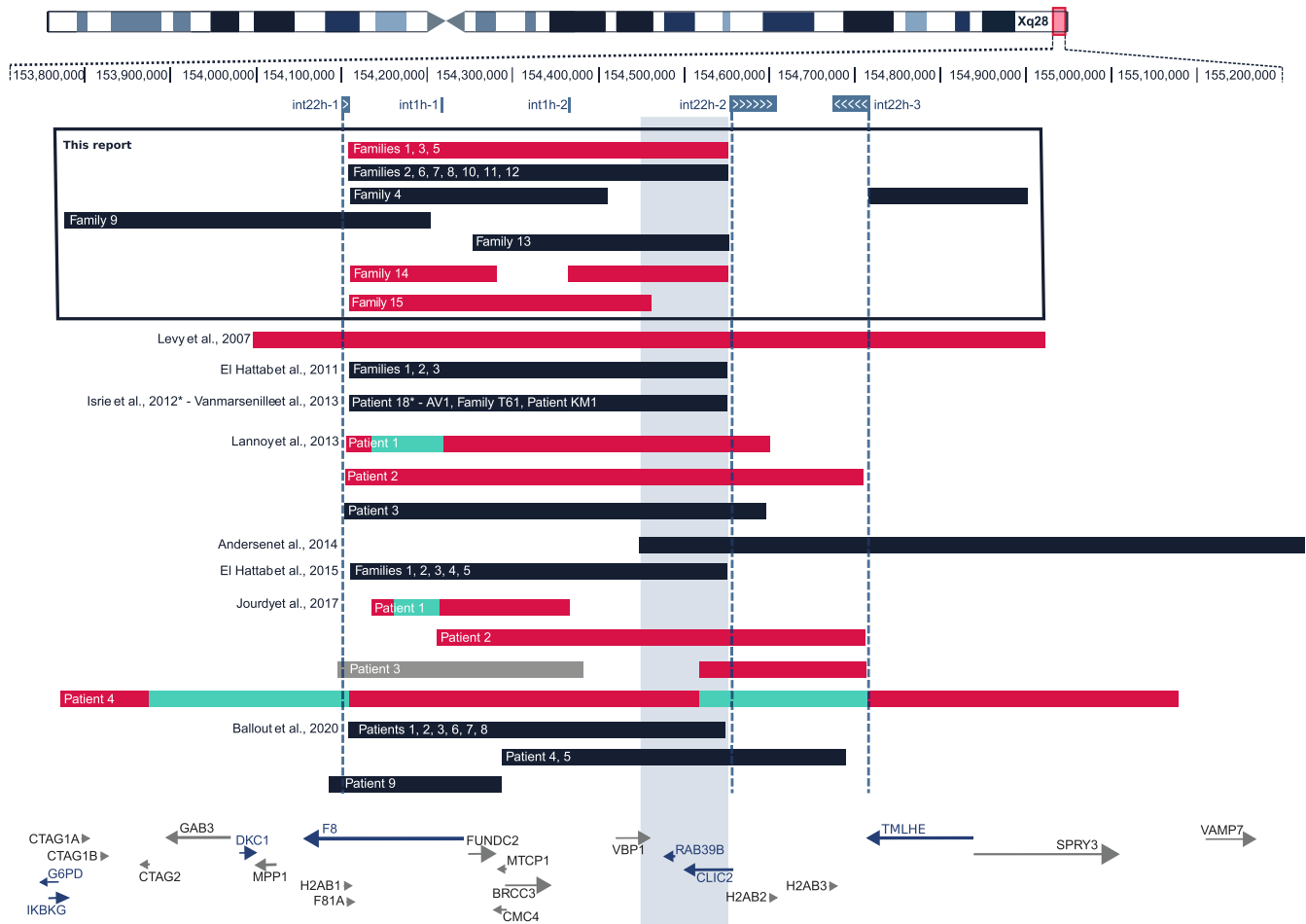


FIGURE 2 Xq28 int22h-1/int22h-2-mediated duplications in our cohort and in the literature sorted by date. CNV are represented as dark blue boxes (male duplication carriers with intellectual disability [ID]), red boxes (male duplication carriers without ID, or families with at least one male carrier without ID), green boxes (triplication), or a light gray box (deletion). The light gray-blue box is the minimum overlap area of duplication with OMIM “morbid” genes in patients with ID (except for the *F8* region, considered to be non-causal for ID). The blue arrows represent OMIM “morbid” genes and the light gray arrows OMIM “non-morbid” genes. The direction of the arrows indicates the direction of transcription. Patient 1 from the study of Lannoy et al. was reported to have moderate HA; Patient 2, severe HA with intron 22 inversion of *F8*; and Patient 3, ID and no HA. Patient 1 from the study of Jourdy et al. was reported to have HA and intron 1 inversion of *F8* and Patients 2, 3, and 4, HA and intron 22 inversion. The four carriers of Family 15 also have the *F8* intron 22 inversion (1 male patient with HA and 3 female patients without HA). The direction of the LCR is represented in accordance with the literature.³⁵ *Patient 18 from the study of Isrie et al. is the same as Patient AV1 from that of Vanmarsenille et al.

obligatory carrier (Figure 1). We have no information about his professional activity or study path, but he was not reported to have an ID. He had two asymptomatic carrier daughters, III-3 and III-2. No female carrier was tested for X-inactivation bias in this family. No history of intellectual impairment was reported in the rest of the family.

2.3 | Family 5

In this family, the asymptomatic father (I-1) and his daughter (II-1) were carriers of the typical Xq28 duplication. His son (II-2) did not carry the duplication.

The mother was treated with sodium valproate for seizures (treatment continued during pregnancy). Her two children showed

behavioral problems that may fit the spectrum of valproate fetopathy. The daughter (II-1) had a specific learning disability without ID and anxiety-depressive disorder. For this study, she was considered to be symptomatic, as involvement of Xq28 duplication could not be ruled out. Her brother (patient II-2) had oral language disorders and a working memory defect.

2.4 | Family 14

In Family 14, female fetus III-2 was prenatally affected by myelomeningocele with a cerebral impact for which amniocentesis was performed. Array-CGH showed an atypical Xq28 duplication (cf. Table 1 and Figure 2) inherited from the asymptomatic father (II-1), who had

TABLE 1 Characteristics of CNVs (minimal genomic coordinates; Hg19) with the patients' phenotype.

Family	Family 1	Family 2	Family 3
CNV genomic coordinates (hg19)	chrX:154,118,619-154,560,375	chrX:154,120,738-154,560,375	chrX:154,118,619-154,560,375
Typical duplication	+	+	+
Atypical duplication	-	-	-
Length	441 kb	439,6 kb	441 kb
Individual	II-5	II-1	I-2 IV-1
Gender	M	M	F M
Age at screening	6	37	18
Skewed X inactivation	NP	NP	NP
Phenotype			
Symptomatic	+	-	+
Unaffected	-	+	-
Intellectual disability	+	-	+
Motor delay	-	-	+
Language delay	+	+	-
Hypotonia	-	-	-
Behavioral abnormalities	+	-	+
Sleep disturbance	-	+	-
Psychiatric signs	-	-	-
Recurrent infections	-	-	-
Atopia	-	-	-
Overweight	+	-	-
Dysmorphic features	-	Strabismus Low-settled ears Imperforate anus	Bilateral talipes equinovarus
Other	Encopresis	Perinatal anoxia	Medical termination of pregnancy
Study level/schooling	NA	Stopped at middle school	Polycystic ovaries
	NA	Vocational baccalaureate	Medical termination of pregnancy
	NA	special needs educational division	Medical termination of pregnancy
	NA		IUGR
	NA		Duplex kidneys
	NA		Medical termination of pregnancy
	NA		NA

TABLE 1 (Continued)

Family	Family 1		Family 2		Family 3		Family 4		Family 5		Family 6		Family 7	
	Profession	NA	Unemployed	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Craftsman
Family														
CNV genomic coordinates (hg19)														
Typical duplication	-													+
Atypical duplication	+													-
Length	296.3 kb													440 kb
Individual	II-2													II-1
Gender	M													F
Age at screening	2													28
Skewed X inactivation														90/10
Phenotype														
Symptomatic	+													+
Unaffected	-													-
Intellectual disability	+													+
Motor delay	+													UN
Language delay	+													UN
Hypotonia	+													UN
Behavioral abnormalities	UN													Self-harming
Sleep disturbance	UN													Sleep apnea
Psychiatric signs	-													+
Recurrent infections	UN (VZV septicemia)													UN
Atopia	Eczema													-
Overweight	-													-
Dysmorphic features	Microcephaly (-2.5 SD) Large fontanelles													Facial dysmorphism Long breakable teeth High-arched palate
Other	Convulsive encephalopathy Cerebral MRI anomalies Died at 2 years													Lymphoedema
Study level/schooling	NA													Vocational qualification

(Continues)

TABLE 1 (Continued)

Family	Family 4	Family 5	Family 6	Family 7
Profession	NA	NA	Housewife	NA
Family	Family 8	Family 9	Family 10	Family 11
CNV genomic coordinates (hg19)	chrX:154,120,798-154,560,375	chrX:153,786,432-154,213,569	chrX:154,118,619-154,560,375	chrX:154,118,619-154,560,375
Typical duplication	+	-	+	+
Atypical duplication	-	+	-	-
Length	440 kb	427 kb	441,757 kb	442 kb
Individual	III-2	II-2	III-1	II-1
Gender	M	F	M	F
Age at screening	14	38	11	16
Skewed X inactivation		84/16	98/02	NP
Phenotype				
Symptomatic	+	-	+	+
Unaffected	-	+	-	-
Intellectual disability	+	-	+	+
Motor delay	+	+	-	-
Language delay	+	+	-	+
Hypotonia	+	+	-	-
Behavioral abnormalities	Aggressiveness Intolerance to Frustration Tantrum	ADHD Stereotypy	ADD Anxiety Impulsiveness Self-harm	Enuresis
Sleep disturbance				
Psychiatric signs				
Recurrent infections				
Atopia				
Overweight				
Dysmorphic features	Facial asymmetry Triangular facies Long eyelashes Large ears Thin superior lip High arched palate Retrogathism Unilateral ptosis Scoliosis	Micro-plagiocephaly Hypertrophy of the nasal bones Middle face hypoplasia Blepharophimosis Low set ears Pectus excavatum Single transverse palmar fold Long thing fingers	Thick eyebrows Index fingers clinodactyly	

TABLE 1 (Continued)

Family	Family 12	Family 13	Family 14	Family 15	Total = 15
Atopia	-		NA	-	
Overweight	+	+	NA	-	
Dysmorphic features	-	-	NA	-	
Other	Astigmatism Normal cerebral IRM	Gynecomastia Pes planus	Myelomeningocele	Anterior balanic hypospadias Severe early onset hemophilia (F8 int22 inversion)	F8 int22 inversion carrier, no hemophilia F8 int22 inversion carrier, no hemophilia
Study level/schooling	Needs	specialized educational care	Baccalaureate	specialized educational care	School certificate
technician's certificate	No difficulties				Advanced
Profession	NA	Cashier	Cleaner	NA	NA
				Needs	Technician

Note: Probands are shown in bold. Empty cells correspond to data not provided. Fetus family 3 IV-1 and IV-2 and fetus family 14 III-2, with bilateral talipes equinovarus, intrauterine fetal growth restriction and duplex kidneys, and myelomeningocele, respectively, were considered to be neither symptomatic nor asymptomatic because the antenatal signs were too nonspecific and have not been reported for xq28 duplication in the literature.

Abbreviations: ADD, attention deficit disorder; ADHD, attention-deficit/hyperactivity disorder; F, female; GW, gestational week; IUGR, intrauterine fetal growth restriction; M, male; NA, not applicable; NP, not-performed; OCD, obsessive-compulsive disorder; UN, unknown.

an advanced technician's certificate and did not report academic difficulties. These duplications were considered to be incidental findings in the fetus but because of the severity of the myelomeningocele, the parents opted to terminate the pregnancy at 23 weeks gestation (fetal autopsy was rejected). The paternal grandmother was a heterozygous carrier.

We considered this duplication to be atypical because there were normal array-CGH probes within the copy number variation (CNV) (Figure 2). Both duplications have been proven to be in *cis*. We were unable to further characterize this chromosomal rearrangement.

2.5 | Family 15

The proband of this family was a male (III-1) diagnosed with hypospadias at birth and severe hemophilia at 1 year of age due to multiple episodes of ecchymosis. He carried an atypical Xq28 duplication that did not span *RAB39B* nor *CLIC2*, associated with the recurrent pathogenic intron 22 inversion of *F8* responsible for hemophilia A (HA). He showed normal psychomotor development. His mother (II-2), an aunt (II-3), and his grandmother (I-2) were asymptomatic carriers. For this study, we considered patient III-1 to be symptomatic because of hypospadias (already described in the Xq28 duplication syndromic spectrum) but he did not show a neurodevelopmental disorder.

2.6 | Overall cohort

Our cohort included 40 patients, 21 females (one fetus) and 19 males (two fetuses). Twenty-one individuals were considered to be asymptomatic (16 females and 5 males). Proband had inherited the Xq28 duplication from an unaffected carrier parent in 13 cases (11 mothers, 2 fathers) and inheritance was not known in one case (Family 7). Asymptomatic carrier mothers possibly inherited the duplication from their fathers in Families 8 and 10, in which segregation was impossible for the father and mothers were proven non-carriers (Figure 1). The CNV of all probands was confirmed by targeted analysis, such as qPCR, MAQ, MLPA, or FISH (not available for Family 5). For two of the symptomatic carriers, environmental factors could explain part of their phenotype (Family 1, individual II.3: perinatal anoxia; Family 5, individual II-1: fetal valproate syndrome; see Table 1 and Figure 1). No identical formal investigation of ID was performed on our patients.

The cohort included three carrier fetuses. Their neurodevelopmental status is unknown (terminated pregnancies, impossible to carry out a childhood evaluation).

The Xq28 chromosomal microduplications from our cohort are presented in Figure 2 and compared to those reported in the literature. Typical recurrent Xq28 int22h-1/Int22h-2-mediated duplication was present in Families 1, 2, 3, 5, 6, 7, 8, 10, 11, and 12. Atypical Xq28 duplications were present in Families 4, 9, 13, 14, and 15 (Figure 2).

In Family 4, the proband and his asymptomatic mother were also carriers of another Xq28 duplication, classified as a variant of unknown significance (arr[GRCh37] Xq28(154722370_154908471)x2 mat).

Although recurrent infections, atopia, overweight or obesity, and facial dysmorphism have been described in the literature, none of these elements were clearly apparent in our cohort. Patient II-1 of Family 11 was the only one reported with recurrent infections (otitis). Patient II-2 from Family 4 died from varicella zoster virus septicaemia at 2 years of age, which can be considered as a sign of immune dysfunction, but without certitude. He was also the only patient in our cohort with atopia. Finally, patient II-5 from Family 1 had a tonsillectomy, which could be a sign of recurrent infections, but his medical record did not explicitly mention it. Six of our patients were overweight or obese and seven were not (status unknown for 24, nonapplicable for the three fetuses). We found no recurrent or distinctive facial dysmorphism in our cohort.

3 | DISCUSSION

Our study is the first to report unaffected male carriers of the typical int22h-1/int22h-2 duplication (Families 1, 3, 5). This finding suggests incomplete penetrance of this CNV in male hemizygous carriers and should change genetic counseling with respect to neurodevelopment when this variant is identified. Among the 27 patients with a typical int22h-1/int22h-2 Xq28 duplication in our cohort, 8 of 12 (~67%) live-born male carriers are intellectually disabled (including Patient II-3 from Family 1, who experienced perinatal cerebral anoxia). Patient I-1 from Family 10 and I-1 from Family 8 are likely male carriers, but they were not tested and therefore their symptomatic/asymptomatic status is unknown. Three of 13 (~23%) live-born female carriers of a typical Xq28 duplication are intellectually disabled (including a probable case of valproate fetal syndrome for Patient II-1, Family 5). Taking into account the typical and atypical duplication carriers, 11/17 (~65%) live-born male carriers are intellectually disabled (~59% excluding questionable Patient II-3 from Family 1) and 4/20 (20%) of live-born female carriers are intellectually disabled (15% excluding questionable Patient II-1, Family 5). Interestingly, the variant is present 13 times in gnomAD-SV v4.0 including three cases of male hemizygous carriers.

These results are inconsistent with data from the most recent review of the literature in which 100% of assessable males and 50% of assessable females with Xq28 duplications were reported to be intellectually disabled.¹⁷ In Family 4, the Xq28 duplication is shorter than the typical duplication, and spare the two ID candidate genes *RAB39B* and *CLIC2* (Figure 2). The proband (II-2) and his mother (I-2) are also carriers of another small Xq28 duplication, considered to be a variant of unknown significance (arr[GRCh37] Xq28(154722370_154908471)x2 mat) that affects the *TMLHE* gene. A complex chromosomal rearrangement with a duplication of *TMLHE* has already been reported for an individual with severe HA without ID²⁸ in which the proband's brother had the same phenotype but was not a carrier of the *TMLHE* duplication. This may suggest that this CNV is not causative, but no further studies were performed. In Family 9 of our cohort, the duplication only partially overlapped with the classical Xq28 int22h-1/int22h-2-mediated duplication (Figure 2), excluding *RAB39B* and *CLIC2*. The familial history suggests a

neurological condition for the father (seizures) and ID for the mother. Genetic investigations could be pursued. In Family 14, we found two monoallelic duplications. These duplications appear to spare the *BRCC3* gene but candidate genes *RAB39B* and *CLIC2* are included (see Figure 2). Ballout et al. reported an atypical Xq28 duplication associated with ID in males (patient 9)^{15,16} that spares *RAB39B* and *CLIC2*.

There are several possible explanations for our observations. A two-hit model, as discussed before for 16p12.1 and 16p13.11 deletions,^{29,30} could explain the observed incomplete penetrance in males and Xq28 int22h-1/int22h-2-mediated duplication could be a risk factor for ID. In our study, there was no exome or genome sequencing data to investigate this possibility, except for Patient II-2 of Family 4 (no additional variant reported). Polygenic etiology or disruption of regulatory elements could also be involved. Another as yet unexplored possibility is the existence of environmental modifying factors (exogenic agents in pregnancy, educational environment, etc.).

In females, incomplete penetrance could result from skewed X-inactivation but no correlation between affected females and X-inactivation has been yet described.³¹⁻³⁴ In our study, six females were tested for skewed X-inactivation. Two asymptomatic carrier mothers (Family 6, Patient I-2; Family 8, Patient II-2) and two symptomatic carrier probands (Family 7, Patient II-1; Family 9, Patient I-2) show skewed X-inactivation. However, one symptomatic carrier female does not show skewed X-inactivation (Family 6, Patient II-4). Thus, there is no clear correlation between skewed X-inactivation and the phenotype.

The last possibility is that of an observational/reporting bias. Xq28 int22h-1/int22h-2-mediated duplication could be a rare CNV incidentally identified in affected patients referred for testing in a setting in which array-CGH are used as a first-tier test. In our four families with asymptomatic male carriers of Xq28 duplications, family testing was required after discovery of the CNV in fetuses, with array-CGH indicated for common ultrasound signs. Asymptomatic male carriers of atypical duplications encompassing the int22h-1/int22h-2 interval and the candidate genes for ID have been reported (Levy et al.²⁵; Lannoy et al.²⁶; Jourdy et al.²⁷; gnomAD DUP_X_54663 and DUP_X_54668) but are poorly discussed in the reports that describe the affected patients. To date and to our knowledge, no benign or likely benign typical Xq28 int22h-1/int22h-2 duplication are described in publicly available databases (Decipher, Clinvar). The variant has been observed 13 times in gnomAD-SV v4 population database with equivalent allele frequencies for females ($n = 10$, 0.015%) and males ($n = 3$, 0.01%).

Our retrospective study is based on data collection by multiple teams in multiple French university hospitals. Some of the collected data from the medical records may be imprecise. The method used to evaluate ID was not specified in our data, but array-CGH testing of probands was requested by specialists accustomed to determining ID. To overcome the absence of formal neuropsychological evaluation of unaffected carriers, we used educational attainment to approximate the autonomy of carriers and therefore the absence of ID.

Our study sheds new light on the possible incomplete penetrance of Xq28 int22h-1/int22h-2-mediated duplication. This has an

important impact on genetic counseling, in particular, in situations of prenatal diagnosis. In our cohort, three fetuses had inherited the duplication from their mother. In all cases, the CNV was considered to be an incidental finding with respect to the reason for the referral. Several couples opted for termination of the pregnancy, assuming complete penetrance.

Further characterization of the penetrance of Xq28 duplication will require extensive segregation studies to increase the number of male carrier cases and thus clarify genetic counseling. If incomplete penetrance is confirmed, this would justify the pursuit of genetic investigations beyond array-CGH to rule out the presence of another pathogenic variant before genetic counseling.

Our cohort study allowed us to assert incomplete penetrance for ID in males carrying the 0.5 Mb typical Xq28 int22h-1/int22h-2-mediated duplication, as we report four unaffected male carriers. Further studies with extensive segregation analysis are required to better determine and quantify penetrance and expressivity so that genetic counseling can be adapted accordingly.

AUTHOR CONTRIBUTIONS

Nicolas Chatron co-written the paper with Alexis Billes. Alexis Billes made the figures. Mathilde Pujalte helped to collect data on families, to review the manuscript and she created the framework for the Table 1. Gilles Morin made feedback on the manuscript. Audrey Putoux, Nicolas Chatron, Mathilde Pujalte, Damien Sanlaville, Caroline Schluth Bolard and Marianne Till from the regional university hospital Hospices Civils de Lyon and James Lespinasse from Centre Hospitalier de Chambéry provided information on families 1 and 2. Alexis Billes, Gilles Morin, Guillaume Jedraszak and Florence Amram from the regional university hospital of Amiens Picardie provided information on family 3. Klaus Dieterich, Gaëlle Vieville and Charles Coutton from the regional university hospital of Grenoble-Alpes provided information on family 4. Patrick Callier and Benoît Mazel from the regional university hospital of Dijon Bourgogne provided information on family 5. Perrine Pennamen and Caroline Rooryck from the regional university hospital of Bordeaux provided information on families 6, 7, 8 and 9. Elise Brischoux-Boucher, Daniel Amsallem, Paul Kuentz, Juliette Piard, Virginie Roze-Guillaumey, Chloé Trouve and Anne-Laude Avice Denizet from regional university hospital of Besançon provided information on families 10, 11, 12 and 13. Odile Boute, Sonia Bouquillon, Elise Boudry-Labis and Mélanie Rama from the regional university hospital of Lille provided information on family 14. Marlène Rio from the Necker-Enfants Malades Hospital provided information on family 15.

AFFILIATIONS

¹CHU Amiens Picardie, Service de Génétique Clinique, Amiens, France

²CHU Amiens Picardie, Laboratoire de Génétique Constitutionnelle, Amiens, France

³Service de Génétique, Groupement Hospitalier Est, Hospices Civils de Lyon, Bron, France

⁴CHU Amiens Picardie, Département de génétique, UR4666 HEMATIM, CURS, Université Picardie Jules Verne, Amiens, France

⁵Service de Neuropédiatrie, CHU de Besançon, Besançon, France

⁶Institut de Génétique Médicale, Hôpital Jeanne de Flandre, Centre Hospitalier Universitaire de Lille, Lille, France

⁷Génétique Clinique, Centre Hospitalier Universitaire de Lille, Hôpital Jeanne de Flandre, Lille, France

⁸Centre de Génétique Humaine - CHU de Besançon, Université de Bourgogne-Franche-Comté, Besançon, France

⁹Inserm UMR 1231 GAD Génétique des Anomalies du Développement, Université de Bourgogne, Dijon, France

¹⁰Unité Fonctionnelle Innovation diagnostique dans les maladies rares, laboratoire de génétique chromosomique et moléculaire, Plateau Technique de Biologie, CHU Dijon Bourgogne, Dijon, France

¹¹Fédération Hospitalo-Universitaire Médecine Translationnelle et Anomalies du Développement (FHU TRANSLAD), CHU Dijon Bourgogne et Université de Bourgogne-Franche Comté, Dijon, France

¹²Service de Génétique, Génomique, et Procréation, Centre Hospitalier Universitaire Grenoble Alpes, La Tronche, France

¹³INSERM 1209, CNRS UMR 5309, Institut pour l'Avancée des Biosciences (IAB), Université Grenoble Alpes, Grenoble, France

¹⁴Université Grenoble Alpes, Inserm, U1216, CHU Grenoble Alpes, Medical Genetics, Grenoble Institute of Neurosciences, Grenoble, France

¹⁵CHU Grenoble, UM Génétique Chromosomique, Grenoble, France

¹⁶Oncobiologie Génétique Bioinformatique, PCBio, CHU de Besançon, Besançon, France

¹⁷UFR Des Sciences de Santé, INSERM-Université de Bourgogne UMR1231 GAD "Génétique des Anomalies du Développement", FHUTRANSLAD, Dijon, France

¹⁸Centre Hospitalier de Chambéry, Service de Cytogénétique, Chambéry, France

¹⁹Centre de Génétique et Centre de Référence Anomalies du Développement et Syndromes Malformatifs, FHUTRANSLAD - CHU Dijon Bourgogne, Dijon, France

²⁰CHU Bordeaux, Laboratoire de Génétique Biologique, Bordeaux, France

²¹Université Paris Cité, Institut Imagine, Inserm U1163, Paris, France

²²Service de Médecine Génomique des maladies rares, AP-HP, Centre Hôpital Necker-Enfants Malades, Paris, France

²³UMR 1231 GAD, Inserm, Université de Bourgogne Franche Comté, Dijon, France

²⁴Hospices Civils de Lyon, Service de Génétique, Groupement Hospitalier Est, Bron, France

²⁵Equipe GENDEV, Centre de Recherche en Neurosciences de Lyon, INSERM U1028 CNRS UMR5292, Université Claude Bernard Lyon 1, Lyon, France

²⁶Institute NeuroMyoGène, Laboratoire Physiopathologie et Génétique du Neurone et du Muscle, CNRS UMR 5261-INSERM U1315, Université de Lyon - Université Claude Bernard Lyon 1, Lyon, France

²⁷Laboratoire de Diagnostic Génétique, Institut de Génétique Médicale d'Alsace, Hôpitaux Universitaires de Strasbourg, Strasbourg, France

²⁸Département de Génétique et Procréation, Hôpital Couple Enfant, CHU Grenoble-Alpes, Grenoble, France

²⁹CHU de Bordeaux, Service de Génétique Médicale, Bordeaux, France

³⁰Univ. Bordeaux, Maladies Rares: Génétique et Métabolisme (MRGM), U 1211 INSERM, Bordeaux, France

ACKNOWLEDGMENTS

We would like to thank all members of AchroPuce for the present, past, and future collaborations. We would like to posthumously thank Pr Lionel Van Maldergem^(†), from regional university hospital of Besançon, for the information he provided about his patients.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

PEER REVIEW

The peer review history for this article is available at <https://www.webofscience.com/api/gateway/wos/peer-review/10.1111/cge.14525>.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ETHICS STATEMENT

Written informed consent was received from the patients. The authors adhere to the Declaration of Helsinki Principles. This study was approved by the local institutional Ethics Committee (CNIL Register Number 22_5867).

ORCID

Elise Brischoux-Boucher  <https://orcid.org/0000-0002-4816-1431>

Charles Coutton  <https://orcid.org/0000-0002-8873-8098>

Juliette Piard  <https://orcid.org/0000-0001-5244-5325>

Caroline Schluth-Bolard  <https://orcid.org/0000-0002-2279-4273>

Caroline Rooryck  <https://orcid.org/0000-0002-9789-3591>

Nicolas Chatron  <https://orcid.org/0000-0003-0538-0981>

REFERENCES

1. Ropers HH, Hamel BCJ. X-linked mental retardation. *Nat Rev Genet.* 2005;6(1):46-57.
2. Chiurazzi P, Schwartz CE, Gecz J, Neri G. XLMR genes: update 2007. *Eur J Hum Genet.* 2008;16(4):422-434.
3. Neri G, Schwartz CE, Lubs HA, Stevenson RE. X-linked intellectual disability update 2017. *Am J Med Genet A.* 2018;176(6):1375-1388.
4. Whibley AC, Plagnol V, Tarpey PS, et al. Fine-scale survey of X chromosome copy number variants and indels underlying intellectual disability. *Am J Hum Genet.* 2010;87(2):173-188.

5. Sanlaville D, Schluth-Bolard C, Turleau C. Distal Xq duplication and functional Xq disomy. *Orphanet J Rare Dis.* 2009;20(4):4.
6. Sanlaville D, Prieur M, de Blois MC, et al. Functional disomy of the Xq28 chromosome region. *Eur J Hum Genet.* 2005;13(5):579-585.
7. Bauters M, Van Esch H, Friez MJ, et al. Nonrecurrent MECP2 duplications mediated by genomic architecture-driven DNA breaks and break-induced replication repair. *Genome Res.* 2008;18(6):847-858.
8. del Gaudio D, Fang P, Scaglia F, et al. Increased MECP2 gene copy number as the result of genomic duplication in neurodevelopmentally delayed males. *Genet Med.* 2006;8(12):784-792.
9. Liu P, Carvalho CMB, Hastings PJ, Lupski JR. Mechanisms for recurrent and complex human genomic rearrangements. *Curr Opin Genet Dev.* 2012;22(3):211-220.
10. Watson CT, Marques-Bonet T, Sharp AJ, Mefford HC. The genetics of microdeletion and microduplication syndromes: an update. *Annu Rev Genomics Hum Genet.* 2014;15:215-244.
11. Lubs H, Abidi F, Bier JA, et al. XLMR syndrome characterized by multiple respiratory infections, hypertelorism, severe CNS deterioration and early death localizes to distal Xq28. *Am J Med Genet.* 1999;85(3):243-248.
12. Van Esch H, Bauters M, Ignatius J, et al. Duplication of the MECP2 region is a frequent cause of severe mental retardation and progressive neurological symptoms in males. *Am J Hum Genet.* 2005;77(3):442-453.
13. El-Hattab AW, Fang P, Jin W, et al. Int22h-1/int22h-2-mediated Xq28 rearrangements: intellectual disability associated with duplications and in utero male lethality with deletions. *J Med Genet.* 2011;48(12):840-850.
14. El-Hattab AW, Schaaf CP, Fang P, et al. Clinical characterization of int22h1/int22h2-mediated Xq28 duplication/deletion: new cases and literature review. *BMC Med Genet.* 2015;16:12.
15. Ballout RA, Dickerson C, Wick MJ, et al. Int22h1/Int22h2-mediated Xq28 duplication syndrome: de novo duplications, prenatal diagnoses, and additional phenotypic features. *Hum Mutat.* 2020;41(7):1238-1249.
16. Ballout RA, El-Hattab AW. The int22h1/int22h2-mediated Xq28 duplication syndrome: an intersection between neurodevelopment, immunology, and cancer. *Genes (Basel).* 2021;12(6):860.
17. Ballout RA, El-Hattab AW, Schaaf CP, Cheung SW. Xq28 duplication syndrome, Int22h1/Int22h2 mediated. In: Adam MP, Mirzaa GM, Pagon RA, et al., eds. *GeneReviews*[®]. University of Washington; 1993 Accessed June 21, 2023. <http://www.ncbi.nlm.nih.gov/books/NBK349624/>
18. Isrie M, Froyen G, Devriendt K, et al. Sporadic male patients with intellectual disability: contribution of X-chromosome copy number variants. *Eur J Med Genet.* 2012;55(11):577-585.
19. Andersen EF, Baldwin EE, Ellingwood S, Smith R, Lamb AN. Xq28 duplication overlapping the int22h-1/int22h-2 region and including RAB39B and CLIC2 in a family with intellectual and developmental disability. *Am J Med Genet A.* 2014;164A(7):1795-1801.
20. Vanmarsenille L, Giannandrea M, Fieremans N, et al. Increased dosage of RAB39B affects neuronal development and could explain the cognitive impairment in male patients with distal Xq28 copy number gains. *Hum Mutat.* 2014;35(3):377-383.
21. Antonarakis SE, Kazazian HH, Tuddenham EG. Molecular etiology of factor VIII deficiency in hemophilia A. *Hum Mutat.* 1995;5(1):1-22.
22. Takano K, Liu D, Tarpey P, et al. An X-linked channelopathy with cardiomegaly due to a CLIC2 mutation enhancing ryanodine receptor channel activity. *Hum Mol Genet.* 2012;21(20):4497-4507.
23. Gregg RG, Metzberg AB, Hogan K, Sekhon G, Laxova R. Waisman syndrome, a human X-linked recessive basal ganglia disorder with mental retardation: localization to Xq27.3-qter. *Genomics.* 1991;9(4):701-706.
24. Wilson GR, Sim JC, McLean C, et al. Mutations in RAB39B cause X-linked intellectual disability and early-onset Parkinson disease with α -synuclein pathology. *Am J Hum Genet.* 2014;95(6):729-735.
25. Levy S, Sutton G, Ng PC, et al. The diploid genome sequence of an individual human. *PLoS Biol.* 2007;5(10):e254.
26. Lannoy N, Grisart B, Eeckhoudt S, et al. Intron 22 homologous regions are implicated in exons 1-22 duplications of the F8 gene. *Eur J Hum Genet.* 2013;21(9):970-976.
27. Jourdy Y, Chatron N, Fretigny M, et al. Molecular cytogenetic characterization of five F8 complex rearrangements: utility for haemophilia A genetic counselling. *Haemophilia.* 2017;23(4):e316-e323.
28. Chen C, Xie X, Wu X, et al. Complex recombination with deletion in the F8 and duplication in the TMLHE mediated by int22h copies during early embryogenesis. *Thromb Haemost.* 2017;117(8):1478-1485.
29. Paciorowski AR, Keppler-Noreuil K, Robinson L, et al. Deletion 16p13.11 uncovers NDE1 mutations on the non-deleted homolog and extends the spectrum of severe microcephaly to include fetal brain disruption. *Am J Med Genet A.* 2013;161(7):1523-1530.
30. Girirajan S, Rosenfeld JA, Cooper GM, et al. A recurrent 16p12.1 microdeletion supports a two-hit model for severe developmental delay. *Nat Genet.* 2010;42(3):203-209.
31. Plenge RM, Stevenson RA, Lubs HA, Schwartz CE, Willard HF. Skewed X-chromosome inactivation is a common feature of X-linked mental retardation disorders. *Am J Hum Genet.* 2002;71(1):168-173.
32. Van den Veyver IB. Skewed X inactivation in X-linked disorders. *Semin Reprod Med.* 2001;19(2):183-191.
33. Minks J, Robinson WP, Brown CJ. A skewed view of X chromosome inactivation. *J Clin Invest.* 2008;118(1):20-23.
34. Shvetsova E, Sofronova A, Monajemi R, et al. Skewed X-inactivation is common in the general female population. *Eur J Hum Genet.* 2019;27(3):455-465.
35. Ross MT, Grafham DV, Coffey AJ, et al. The DNA sequence of the human X chromosome. *Nature.* 2005;434(7031):325-337.

How to cite this article: Billes A, Pujalte M, Jedraszak G, et al. Possible incomplete penetrance of Xq28 int22h-1/int22h-2 duplication. *Clinical Genetics.* 2024;106(3):234-246. doi:10.1111/cge.14525