



Neuroprotective effects of lactate and ketone bodies in acute brain injury

Guillaume Plourde, Hélène Roumes, Laurent Suissa, Lorenz Hirt, Émilie Doche, Luc Pellerin, Anne-Karine Bouzier-Sore, Hervé Quintard

► To cite this version:

Guillaume Plourde, Hélène Roumes, Laurent Suissa, Lorenz Hirt, Émilie Doche, et al.. Neuroprotective effects of lactate and ketone bodies in acute brain injury. *Journal of Cerebral Blood Flow and Metabolism*, 2024, 44 (7), pp.1078-1088. 10.1177/0271678X241245486 . hal-04766006

HAL Id: hal-04766006

<https://hal.science/hal-04766006v1>

Submitted on 5 Nov 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Neuroprotective effects of lactate and ketone bodies in acute brain injury

Guillaume Plourde, Hélène Roumes, Laurent Suissa, Lorenz Hirt, Émilie Doche, Luc Pellerin, Anne-Karine Bouzier-Sore, Hervé Quintard

► To cite this version:

Guillaume Plourde, Hélène Roumes, Laurent Suissa, Lorenz Hirt, Émilie Doche, et al.. Neuroprotective effects of lactate and ketone bodies in acute brain injury. *Journal of Cerebral Blood Flow and Metabolism*, 2024, 44 (7), pp.1078-1088. 10.1177/0271678X241245486 . hal-04631544

HAL Id: hal-04631544

<https://hal.science/hal-04631544v1>

Submitted on 2 Jul 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Neuroprotective effects of lactate and ketone bodies in acute brain injury**

2 Test

3 **Guillaume Plourde, MD, PhD, FRCPC *** (0000-0002-3641-6281)

4 Division of Intensive Care Medicine

5 Department of Medicine

6 Centre hospitalier de l'Université de Montréal

7

8 **Hélène Roumes, PhD** (0000-0002-0387-7823)

9 Centre de Résonance Magnétique des Systèmes Biologiques (CRMSB)

10 Univ. Bordeaux, CNRS, CRMSB, UMR 5536, F-33000 Bordeaux, France

11

12 **Prof. Luc Pellerin, PhD** (0000-0002-1016-1970)

13 IRMETIST Inserm U1313

14 Université et CHU de Poitiers

15

16 **Prof. Laurent Suissa, MD** (0000-0003-2345-8711)

17 Neurovascular Unit

18 CHU de Marseille

19

20 **Prof. Lorenz Hirt, MD** (0000-0002-2921-5000)

21 Division of Neurology

22 Department of Clinical Neuroscience

23 Centre hospitalier universitaire vaudois

24

25 **Émilie Doche, MD**

26 Neurovascular Unit

27 CHU de Marseille

28

29 **Anne-Karine Bouzier-Sore, PhD** (0000-0002-3470-0940)

30 Centre de Résonance Magnétique des Systèmes Biologiques (CRMSB)

31 Univ. Bordeaux, CNRS, CRMSB, UMR 5536, F-33000 Bordeaux, France

32

33 **Prof. Hervé Quintard, MD, PhD** (0000-0002-3870-6370)

34 Division of Intensive Care Medicine

35 Department of Anesthesiology, Clinical Pharmacology, Intensive Care and Emergency Medicine

36 Geneva University Hospital

37

38 * Corresponding author : **Guillaume Plourde, MD, PhD, FRCPC**

39 Division of Intensive Care Medicine

40 Department of Medicine

41 Centre hospitalier de l'Université de Montréal

42 1051 rue Sanguinet, Montréal, Canada

43 Email: guillaume.plourde.1@umontreal.ca

44 Phone: +1.514.890.8000

45

46 Running head: Neuroprotective effects of lactate and ketones

47 Word count: 3709 words

48 **Unstructured abstract**

49 The goal of neurocritical care is to prevent and reverse the pathologic cascades of secondary
50 brain injury by optimizing cerebral blood flow, oxygen supply and substrate delivery. While
51 glucose is an essential energetic substrate for the brain, we frequently observe a strong decrease
52 in glucose delivery and/or a glucose metabolic dysregulation following acute brain injury. In
53 parallel, during the last decades, lactate and ketone bodies have been identified as potential
54 alternative fuels to provide energy to the brain, both under physiological conditions and in case of
55 glucose shortage. They are now viewed as integral parts of brain metabolism. In addition to their
56 energetic role, experimental evidence also supports their neuroprotective properties after acute
57 brain injury, regulating in particular intracranial pressure control, decreasing ischemic volume,
58 and even improving cognitive functions as well as survival. In this review, we present available
59 data supporting a future therapeutic use of lactate and ketone bodies in acute brain injury. We
60 also review preclinical and clinical evidence exploring the mechanisms underlying their
61 neuroprotective effects, and identify research priorities in the field of neuroenergetics.

62

63

64 Keywords: alternative brain fuels, brain metabolism, ketone bodies, lactate, neuroprotection,
65 stroke, TBI, hypoxia.

66

67 **Introduction**

68 Acute brain injury refers to brain damage that occurs suddenly, such as traumatic brain injury
69 (TBI), stroke, subarachnoid hemorrhage (SAH) or global ischemia, for example. The insult itself
70 leads to a primary injury, which is followed by secondary brain damage, a pathologic cascade that
71 arises hours to days after the initial injury. The main causes of this secondary brain injury are
72 cerebral blood flow disturbances, which lead to hypoxia and a strong decrease of glucose delivery
73 to the brain. To improve patient outcome, the goal of neurocritical care, after detection of the acute
74 brain injury, is to avoid or reverse the underlying cascades that occur during this secondary brain
75 injury period by optimizing cerebral blood flow, oxygen supply and substrate delivery to the brain.²
76 Glucose is an essential substrate to satisfy brain energy requirements. However, its availability is
77 frequently reduced following acute brain injury.^{3,4} In the last decades, lactate and ketone bodies
78 have been identified as alternative fuels to provide energy to the brain and preserve glucose for
79 other critical metabolic needs. Results are also accumulating concerning their potential
80 neuroprotective effects and clinical applications.

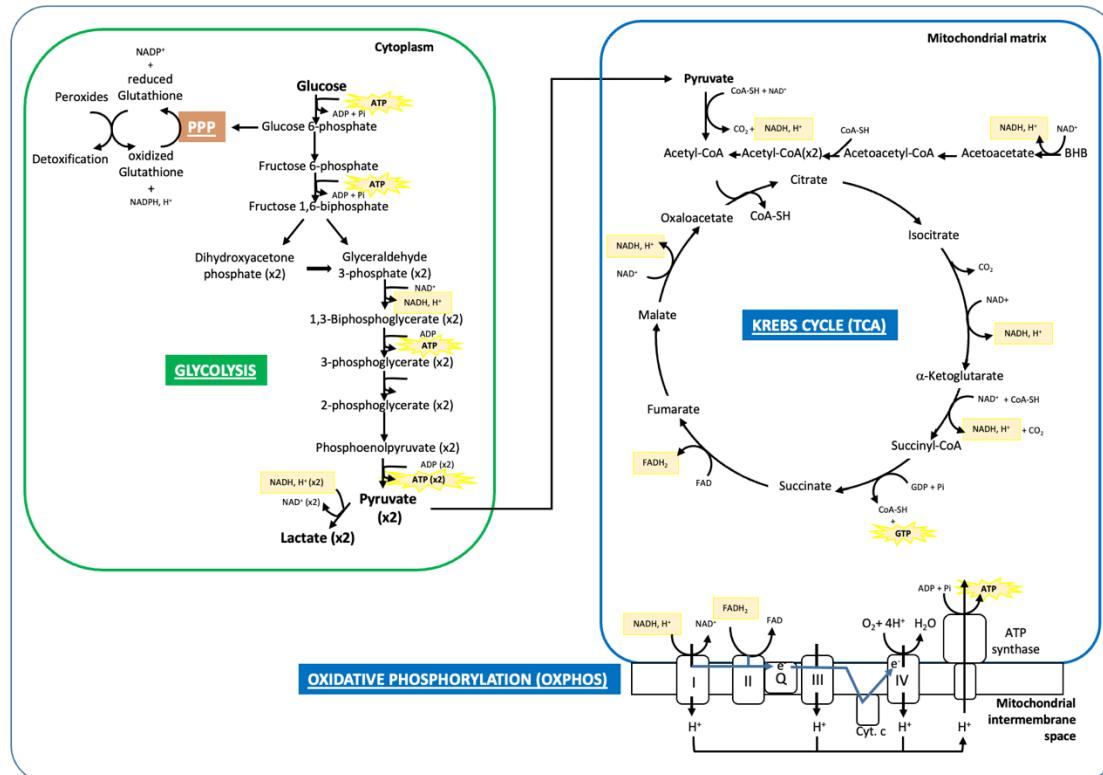
81 In this review, we present available data supporting a therapeutic use of lactate and ketone bodies
82 in acute brain injury. We also review preclinical and clinical evidence exploring the mechanisms
83 underlying their neuroprotective effects, and identify research priorities in the field of
84 neuroenergetics.

85

86 **Glucose for brain energy supply**

87 While the brain represents only 2% of the total body mass, its glucose consumption is around
88 20% of the systemic consumption, which represents more than 10 times the predicted value
89 relative to its mass. Glucose is an essential substrate to maintain normal brain metabolism and
90 function. It is required for glycolysis, which in turn provides 2 ATP, and 2 pyruvates that can enter
91 the tricarboxylic acid (TCA) cycle to ensure energy production, in majority through the generation
92 of reducing equivalents (NADH and FADH₂), which are then used in the electron transport chain

93 to generate around 30 more ATP molecules (the exact yield of ATP from one round of the TCA
 94 cycle can vary but it has been experimentally estimated that NADH can contribute about 2.5 to 3
 95 ATP, and each FADH₂ can contribute about 1.5 to 2 ATP) (**Figure 1**). In addition to provide
 96 energy to the brain, glucose is a precursor in the synthesis of neurotransmitters and participates
 97 in the pentose phosphate pathway (PPP), also known as the hexose monophosphate shunt,
 98 which is a metabolic pathway that runs parallel to glycolysis and is essential for NADPH and
 99 pentose sugar production, as well as for redox homeostasis. Indeed, the PPP plays a crucial role
 100 in maintaining reduced glutathione (GSH) levels in cells by supplying the necessary reducing
 101 equivalents in the form of NADPH. Reduced glutathione acts as a critical antioxidant and
 102 contributes to the cellular defense mechanisms against oxidative stress due to reactive oxygen
 103 species (ROS) production. The PPP is therefore a major pathway for neuronal protection after an
 104 acute brain insult during which high amount of ROS are produced.



105
 106 **Figure 1: Glucose metabolism pathway.** In the cytoplasm, glucose is metabolized to pyruvate through
 107 glycolysis. This pyruvate can either provide lactate or be redirected to the mitochondria to provide ATP
 108 through (i) the Krebs cycle (TCA cycle) coupled to (ii) oxidative phosphorylation (OXPHOS).

109
110 During the acute phase following brain injury, cerebral glucose availability may become
111 insufficient to meet the metabolic demands due to cerebral blood flow failure, blood-brain barrier
112 dysfunction or diffusion impairment. Indeed, glucose storage is very low in the brain. If glycogen
113 can be found in astrocytes, its consumption during ischemia can sustain brain functions for only
114 2 min (measurements made in rats (Swanson et al. 1989)). In human, a decrease in glycemia
115 linked with a decrease in cerebral glucose concentration was demonstrated to be deleterious in
116 the context of acute brain injury, leading to neuroglycopenia [Vespa P, McArthur DL, Stein N, et
117 al. Tight glycemic control increases metabolic distress in traumatic brain injury: a randomized
118 controlled withinsubjects trial. Crit Care Med 2012; 40:1923–1929 / Magnoni S, Tedesco C,
119 Carbonara M, et al. Relationship between systemic glucose and cerebral glucose is preserved in
120 patients with severe traumatic brain injury, but glucose delivery to the brain may become limited
121 when oxidative metabolism is impaired: implications for glycemic control. Crit Care Med 2012;
122 40:1785–1791].
123 Glucose infusion following brain injury has been tested decades ago on a rat model of brain
124 ischemia, aiming to reverse this substrate-deprived state and restore brain energy metabolism.
125 Surprisingly, it was associated with increased mortality and extension of injury.¹ Since then, these
126 results have been consistently replicated in various experimental models of ischemic brain
127 injury.^{1–3} In patients, spontaneous high plasma glucose levels have also been associated with
128 increased infarct size, worse clinical outcomes and increased mortality in ischemic stroke and
129 SAH, especially when it happens before or early after ischemia.^{1–3}
130 With the same idea in mind to control glucose level, insulin therapy was also tested, based on the
131 early promising results of intensive insulin therapy in critically ill surgical patients¹. Many trials
132 have investigated whether insulin-based glycemic control could improve the outcomes of brain-
133 injured patients. Among these, the large GIST-UK trial failed to show a benefit of intensive insulin
134 therapy in patients with acute stroke, although it was likely underpowered due to slow enrollment

135 which justified its early termination.¹ Overall, this trial and other studies pooled in two recent
136 systematic reviews suggest that tight glycemic control causes more symptomatic and
137 asymptomatic hypoglycemic episodes, both associated with poor outcomes.^{1,2}

138 **Ketone bodies and lactate as alternative fuels**

139 It is well established that the brain is metabolically flexible and can metabolize other substrates,
140 such as ketone bodies (KB) and lactate.⁵ KB are produced in the liver from fatty acids. They
141 represent a major source of energy for the brain in neonates, who rely on breastfeeding for their
142 diet which is rich in lipids. In adults, when glucose availability is limited in situations such as
143 fasting, a very low-carbohydrate diet (ketogenic diet), or prolonged exercise, KB are produced as
144 alternative fuels and used significantly by the brain. The three main KB synthetized by the liver
145 are acetoacetate (AcAc), beta-hydroxybutyrate (BHB; AcAc can be converted to BHB, which is
146 the predominant ketone body in circulation) and acetone, a minor KB that is produced as a
147 byproduct. KB enter the brain via monocarboxylate transporters (MCTs) and provide energy to
148 the brain in the form of acetyl-CoA entering the tricarboxylic acid cycle to be oxidized, hence
149 reducing reliance on glycolysis for energy production. This alternative energy production turns out
150 to be highly efficient: BHB, one of the main KB, is converted to acetyl-CoA through only three
151 enzymatic reactions, compared to the ten reactions with several rate-limiting enzymes required
152 to convert glucose to acetyl-CoA. It has been shown in non-fasted adult humans infused with [2,4-
153 ¹³C₂]-BHB and using ¹H-¹³C polarization transfer spectroscopy that KB can cross the blood-brain
154 barrier and are metabolized by the brain [PMID : 12142574]. These observations have been
155 confirmed in pre-clinical studies (Roy et al. 2015). In euglycemic conditions, ketone bodies are
156 predominantly metabolized by neurons and account for 6% of total coenzyme A oxidation in
157 normal conditions.¹ This phenomenon was found to be amplified in a model of trauma.¹ On a
158 juvenile rat model of TBI infused with [2,4-¹³C₂]-BHB, it has been shown using ¹³C magnetic
159 resonance spectroscopy (MRS) that KB were extensively metabolized, mainly in neurons [PMID
160 : 36005582].

161 Concerning lactate, experiments in the 1990s led by Pellerin and Magistretti suggest that lactate
162 is commonly used for basal brain metabolism under physiological conditions. They described a
163 metabolic cooperation between astrocytes and neurons, termed the astrocyte-neuron lactate
164 shuttle (ANLS) (Pellerin and Magistretti 1994). In this model, glutamate released as part of
165 synaptic activity is then taken up by astrocytes, one of their major function to avoid exitotoxicity.
166 Concomitantly with glutamate uptake, a Na^+ influx will occur and its intracellular concentration will
167 rise. As a consequence, Na^+/K^+ ATPase activity will increase, leading to enhanced glucose
168 utilization and lactate production by astrocytes. This astrocytic lactate is then transferred to
169 neurons, through the MCTs (isoforms MCT1 and 4 for astrocytes, MCT2 for neurons; same
170 transporters as the ones used for KB). Once in neurons, lactate is converted back to pyruvate
171 and used as an energetic substrate. Since its description, the ANLS has gained popularity and
172 has been validated in experimental models *in vitro*, *ex vivo* and *in vivo*, although remaining
173 controversial (Bak and Walls 2018, Barros and Weber 2018). Using infusion of [3- ^{13}C]lactate and
174 ^{13}C MRS it has been shown in rats that lactate efficiently cross the blood-brain-barrier and once
175 in the brain, it is preferentially a neuronal substrate (Bouzier et al. 2000). These observations
176 were confirmed in another study that showed that extracerebral lactate contributes at least two-
177 fold more to replenish the neuronal than the glial pyruvate pools [PMID : 25522255]. Authors
178 concluded that a primary utilization of exogenous lactate takes place in neurons rather than in
179 astrocytes (Bouzier et al. 2000). Moreover, lactate entry into neurons through its transporter
180 MCT2 was shown to be necessary to sustain brain function (Suzuki et al. 2011, Netzahualcoyotzi
181 and Pellerin 2020, Roumes et al. 2021). Under supraphysiological conditions, lactate can
182 represent up to 60% of brain energy supply. More recently, preferential lactate uptake by the brain
183 over glucose has been described in a healthy rat model¹ and in exercising man¹⁻³, likely reflecting
184 an adaptation of the cerebral metabolic ratio to transient hyperlactatemia and relative glucose
185 shortage. A similar observation has been made in patients with acute brain injury, for who an

186 increase in lactate production occurred during spontaneous glucose depletion despite the
187 absence of brain hypoxia.¹⁻³ Finally, blocking astrocyte lactate transport in a rat model of cerebral
188 ischemia altered functional recovery, suggesting an obligatory role of lactate in basal
189 neuroenergetics.¹ This obligatory role was further confirmed in another rat model of neonatal
190 hypoxia-ischemia, in which reductions in brain lesion volume and ROS production were observed
191 when lactate was injected, but not when it was co-administered with oxamate, a lactate
192 dehydrogenase inhibitor therefore avoiding its conversion into pyruvate and therefore its
193 metabolism (Roumes et al. 2021).

194 Thus both KB and lactate can serve as an alternative oxidative substrate for the brain and are
195 mainly metabolized by neurons. They both enter through MCTs and deliver energy after their
196 oxidation in the TCA cycle, entering either at the acetylCoA or at the pyruvate level, respectively
197 (**Figure 2**). In both cases, their metabolism also lead to the conversion of cytosolic NAD⁺ into
198 NADH and thus modify the redox potential of the cell (during the conversion of BHB into AcAc or
199 lactate into pyruvate).

200 Taken altogether, challenges in glucose availability after an acute brain injury, combined with the
201 possibility of the brain to metabolize efficiently KB and lactate, offer new perspectives. A direct
202 modulation of neuronal metabolism by these alternative energy substrates could be of major
203 interest in the search for therapeutic avenues in the context of acute brain injury.

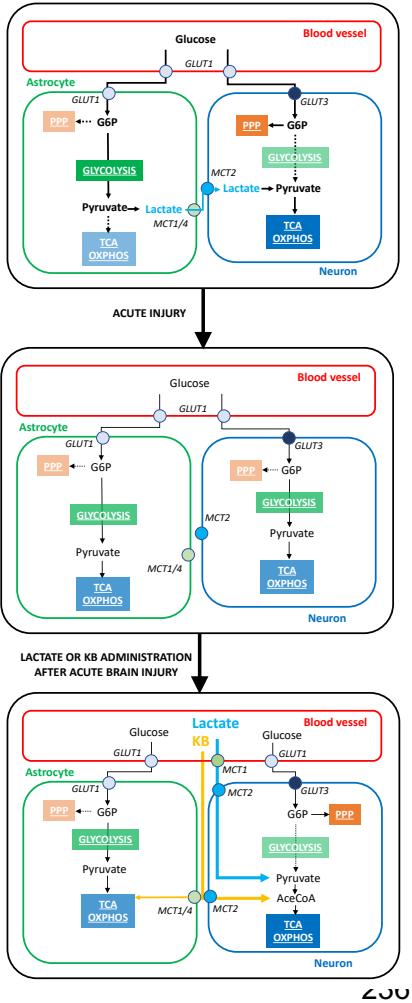


Figure 2: Compartmentalization of cerebral glucose metabolism and consequence in brain injury as well as for neuroprotection strategies. A- In the brain, glucose from the bloodstream enters both astrocytes and neurons. It is metabolized to pyruvate in the cytoplasm (glycolysis) then supplies ATP via pyruvate oxidation into the mitochondria (TCA cycle + OXPHOS). However, astrocytes have a greater glycolytic capacity compared to neurons, which are highly oxidative cells (Herrero-Mendez et al. 2009). Astrocytes are known to produce high level of lactate, which can be further transferred to neurons where it can be used as an energy substrate through the TCA and OXPHOS. Glucose can also enter directly into the neuron. Low glycolysis in neurons allows glucose conversion into the pentose phosphate pathway PPP to sustain antioxidant protection. B- In acute brain injury condition, the low level of circulating glucose can not meet cerebral energy needs and the metabolic cooperation between astrocyte and neurons can not be maintained. Glucose is mainly used for oxidative metabolism in both cell types to maintain ATP level. C- In order to overcome the energy deficit induced by acute brain injury, energy substitution strategies could be adopted. Lactate or exogenous ketone bodies enter directly into brain cells but predominantly in neurons (Pan et al. 2002, Duarte et al. 2015, Scafidi et al. 2022). Lactate is then converted into pyruvate, while ketone bodies are precursors of acetyl-CoA. Pyruvate and Acetyl-CoA are then oxidized through TCA cycle and OXPHOS to provide energy. Saved glucose can be redirected to the PPP pathway to reestablish the antioxidant defense.

GLUT, glucose transporter; KB, ketone bodies; MCT, monocarboxylate transporter; OXPHOS, oxidative phosphorylation; PPP, pentose phosphate pathway; TCA, tricarboxylic acid cycles.

237 Alternative fuels and the pentose-phosphate pathway

238 Neuronal glycolysis is controlled by the enzymatic complex APC/C-Cdh1 that degrades the
 239 proglycolytic enzyme 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase-3 (PFKFB3). The
 240 enzyme PFKFB3 is less abundant in neurons than in astrocytes, resulting in a lower glycolytic
 241 rate and in the redirection of glucose-6-phosphate through the PPP.^{30,92} Astrocytes, on the other
 242 hand, exhibit a greater PFKFB3 activity, resulting in higher glycolytic rate and lactate production.
 243 Hence, this compartmentalization suggests that neurons use glucose preferentially for regulation
 244 of their redox state rather than for energy requirements (Bolanos 2016). In stressed states, such
 245 after an oxygen-glucose deprived condition, the reperfused neurons showed elevated PFKFB3
 246 expression (Li et al. 2019). Such changes directed neuronal glucose metabolism from PPP to

247 aerobic glycolysis compared to normal neurons, resulting in increased ROS production and
248 apoptosis during reperfusion. Interestingly, both KB and lactate are oxidative substrates and
249 provide acetylCoA or pyruvate, respectively, while contributing to the conversion of cytosolic
250 NAD⁺ into NADH. An increase in the NAD⁺/NADH ratio will inhibit glycolysis and redirect glucose
251 through the PPP, and therefore will increase the pool of reduced glutathione. Indeed, it has been
252 shown that both alternative fuels can decrease ROS damages réf 12 + Kim, do Y., et al. Ketone
253 bodies are protective against oxidative stress in neocortical neurons. J. Neurochem. 2007;
254 101:1316–1326. [PubMed: 17403035] Maalouf M, et al. Ketones inhibit mitochondrial production
255 of reactive oxygen species production following glutamate excitotoxicity by increasing NADH
256 oxidation. Neuroscience. 2007; 145:256–264. [PubMed: 17240074] Jarrett S, et al. The ketogenic
257 diet increases mitochondrial glutathione levels. J. Neurochem. 2008; 106:1044–1051. [PubMed:
258 18466343] + [Joseph Lemire 1, Christopher Auger, Ryan Mailloux, Vasu D Appanna.
259 Mitochondrial lactate metabolism is involved in antioxidative defense in human astrocytoma cells.
260 J Neurosci Res . 2014 Apr;92(4):464-75. doi: 10.1002/jnr.23338. Epub 2014 Jan 22.] whereas
261 the activity of G6PD, the key enzyme of the first (and irreversible) step of the PPP, was
262 significantly increased with lactate in neuronal cultures Pötzsch A, Zocher S, Bernas SN, Leiter
263 O, Rünker AE, Kempermann G. L-lactate exerts a pro-proliferative effect on adult hippocampal
264 precursor cells *in vitro*. iScience. 2021 Feb 3;24(2):102126. doi: 10.1016/j.isci.2021.102126.
265 PMID: 33659884; PMCID: PMC7895751.

266

267 **Lactate and ketone bodies to control neuroinflammation**

268 Emerging data suggest that lactate and ketone bodies have anti-inflammatory properties, but the
269 underlying mechanisms remain unclear. Microglia are key regulators of brain homeostasis and
270 act as Janus-faced cells regarding neuroinflammation.⁹³ After an ischemic event, microglia are
271 activated and release pro-inflammatory factors, such as TNF- α , IL-1 β , IL-6 and IFN- γ , exhibiting

272 a pro-inflammatory or “M1” microglial phenotype.⁹⁴ On the other hand, microglia can also promote
273 tissue and vascular remodeling via expression of anti-inflammatory factors such as IL-10, VEGF,
274 TGF- β and BDNF, exhibiting an anti-inflammatory or “M2” phenotype. These microglial
275 phenotypes are tightly linked to their metabolism, and lactate was shown to switch microglial
276 profiles from a glycolytic M1 to an oxidative M2 phenotype, hence promoting microglial energy
277 production.⁹⁵ Lactate has been shown to inhibit microglia-mediated neuroinflammation through
278 the Akt-pathway leading to elongation of microglial processes [Hong, Hongxiang; Su, Jianbin;
279 Zhang, Yi; Xu, Guanhua; Huang, Chao; Bao, Guofeng; Cui, Zhiming. A novel role of lactate:
280 Promotion of Akt-dependent elongation of microglial process. International Immunopharmacology
281 June 2023 119 DOI: 10.1016/j.intimp.2023.110136]. BHB was shown to exert similar effects
282 through induction of microglial ramifications and of an anti-inflammatory response in a Akt-
283 dependant manner.⁹⁶ There is also evidence that a ketogenic diet reduces ROS generation and
284 suppresses inflammasome activation.⁹⁷ In another study, ketogenic diet and BHB post-treatment
285 reduced infarct volume and suppressed the over-activation of microglia in a mouse model of MCA
286 occlusion reference 85.

287 Together with glucose redirection through the PPP, these anti-inflammatory effects might
288 constitute an elegant hypothesis to explain some of the clinical benefits observed with alternative
289 fuel supplementation in acute brain injury.

290

291 **Increase in cerebral blood flow**

292 In an *in vitro* study on rat hippocampal and neocortical slices, cerebral vasomotor tone was
293 influenced by the metabolic state of the tissue. In hypoxic conditions, astrocyte glycolysis and
294 lactate production were increased, causing vasodilation via an attenuation of prostaglandin
295 uptake.^{6,68} In a cohort of patients with severe acute brain injury (SAH and TBI), a lactate infusion
296 increased extracellular glucose levels and improved cerebral blood flow estimated by transcranial
297 Doppler.⁵⁰ Explanation of this mechanism is not well understood. Lactate could act as a

298 transmitter of metabolic information by modulating prostaglandin action and cerebral
299 vasodilatation causing cerebral blood flow to increase, regulates the NAD⁺/NADH redox ratio by
300 conversion to pyruvate, and/or activates a G-protein coupled receptor in neurons, astrocytes,
301 and capillaries (Gordon 2008, Begersen 2012, Lauritzen 2014). Interestingly, KB can also increase
302 cerebral blood flow as shown in mice by MRI⁶⁹, and in human healthy subjects by PET scan⁶⁵ and
303 the Kety-Schmidt technique⁷⁰.

304

305 **Control of brain edema**

306 Intracranial hypertension is one of the major challenges faced by neurocritical care clinicians
307 following TBI, massive SAH or hypoxic-ischemic brain injury. To control intracranial pressure
308 (ICP), conventional treatment is based on a stepwise approach mostly described in TBI and
309 involves deep sedation and analgesia, correct positioning of the patient, and hypertonic sodium
310 chloride or mannitol infusions.⁷¹ If the efficacy of osmotic therapy to control ICP is well established,
311 it is also associated with side effects, among which are hyperchloremic acidosis and fluid
312 overload.⁷² Based on its favorable molecular profile, sodium lactate has been identified as a
313 potential agent to control ICP while avoiding these deleterious effects. It was first tested in a
314 randomized trial in humans against mannitol in the treatment of intracranial hypertension during
315 severe TBI. In this study, lactate was associated with a greater and more prolonged decrease in
316 ICP than mannitol.⁵¹ This result has since been replicated, and was also shown to prevent
317 intracranial hypertensive episodes and fluid overload.^{49,73,74} Moreover, half-molar sodium lactate
318 administered to patients with severe head trauma was associated with a mild metabolic alkalosis,
319 counteracting the hyperchloremic acidosis associated with “traditional” osmotic therapy.⁵² This
320 control of brain edema might explain why lactate has been associated with increased tissue
321 oxygenation. In rat models of TBI, exogenous sodium lactate increased cerebral tissue oxygen
322 pressure, possibly via improvement of oxygen diffusion due to reduced tissue edema.^{75,76} The
323 antiedematous effect of sodium lactate might be induced by an increased expression of VRAC,

324 KCC2 and AQP4, channels involved in cell volume regulation and free water movements across
325 membranes, rather than by an osmotic effect.⁷⁵ Concerning KB, their neuroprotective effects
326 were examined in rats with transient occlusion of middle cerebral artery. The administration of
327 BHB (30 mg.kg⁻¹.h⁻¹) significantly reduced cerebral infarct area and edema formation.
328 <https://doi.org/10.1254/jip.89.36>. Réf 81

329

330 **Reduction of ischemic lesion volume**

331 As described just before, KG have been associated with reductions in infarct volume after middle
332 cerebral artery occlusion⁸¹⁻⁸⁵ but also after TBI.^{82,83} *In vitro* experiments on cultured cortical
333 neurons showed that a BHB-based treatment prevents neuronal death induced by glucose
334 deprivation or hypoxia by stimulation of the autophagic flux thus preventing autophagosome
335 accumulation abd improving neuronal survival.⁸⁴ Moreover, both lactate and ketone bodies can
336 limit excitotoxicity, reducing neuronal death as well as lesion volume. In a glutamate excitotoxicity
337 context, pyruvate coming from lactate is metabolized to produce ATP. This ATP in turn activates
338 the PI3K pathway leading to KATP channels opening. This results in the hyperpolarization of the
339 neuron, decreasing neuronal excitability and leading to a neuroprotective effect [P. Jourdain^{1,2},
340 K. Rothenfusser^{1,2}, C. Ben-Adiba², I. Allaman¹, P. Marquet^{2,3,4,5} & P. J. Magistretti^{1,2,6}. Dual
341 action of L-Lactate on the activity of NR2B-containing NMDA receptors: from potentiation to
342 neuroprotection Scientific Reports Open Access Volume 619 February 2016 Article number
343 21250]. The neuroprotective effect of ketone bodies, targeting glutamate excitotoxicity, has also
344 been shown in rat neocortical neurons [Maalouf M, Sullivan PG, Davis L, Kim DY, Rho JM.
345 Ketones inhibit mitochondrial production of reactive oxygen species production following
346 glutamate excitotoxicity by increasing NADH oxidation. Neuroscience. (2007) 145:256–64. doi:
347 10.1016/j.neuroscience.2006.11.065]. In this deleterious context, ketone bodies can compete
348 with the Cl⁻ anion and inactivate the vesicular glutamate transporters (VGLUTs), decreasing

349 presynaptic glutamate release [Juge N, Gray JA, Omote H, Miyaji T, Inoue T, Hara C, et al.
350 Metabolic control of vesicular glutamate transport and release. *Neuron*. (2010) 68:99–112. doi:
351 10.1016/j.neuron.2010.09.00]. A summary of *in vivo* studies showing a reduction of the infarct
352 volume after KB administration can be found in Table 2 of the following review
353 (<https://doi.org/10.1038/sj.jcbfm.9600>).

354 Lactate was also shown to decrease brain lesion volume in several rat models after acute brain
355 injuries such as neonatal hypoxia-ischemia réf 12 et 80, TBI and stroke,^{12,77–80} ajouter Castillo
356 2015 JCBFM.

357

358 **Improvement of recovery and survival**

359 Both substrates have been associated with improvement of cognitive function. In a rat model of
360 neonatal hypoxia-ischemia, lactate administration was associated with short and long term
361 improvement of motor and cognitive functions (réf 12). Improvement of long term cognitive
362 functions was also observed after lactate injections in rat models of TBI or stroke.^{87–89} Ketone
363 bodies might as well promote cognitive recovery after TBI since they act as histone deacetylase
364 inhibitors, an enzyme involved in memory and cognitive impairment in neurodegenerative
365 diseases.⁴⁴ A ketogenic diet or BHB administered post-treatment improved neurological function
366 in a mouse model of MCA occlusion.⁸⁵ Finally, in a recent meta-analysis of 49 studies on murine
367 models of TBI, ketones were associated with an improved survival, particularly in the adult
368 subgroup.⁹⁰ Interestingly, the plasma level of ketosis was the only predictor of the neuroprotective
369 effect. Another meta-analysis of studies on animal models of stroke described a protective effect
370 of ketosis on outcome⁹¹ while a recent review summarized the different studies showing the
371 neuroprotective effet of KB in acute brain injuries (different models, different species, different
372 ages) <https://doi.org/10.1038/sj.jcbfm.9600543>.

373

374 **Lactate and ketone bodies as pharmacological agents**

375 Lactate can be administered as an intravenous solution of sodium lactate.¹ All interventional
376 studies on TBI used solutions with an osmolality ranging from 500 mmol/L to 1000 mmol/L, the
377 latter being approximately iso-osmolar to 20% mannitol.^{48–54} In two studies, investigators injected
378 sodium lactate directly in the brain using a microdialysis technique using 4 and 8 mmol/L
379 microdialysis solutions.^{55,56} Both methods of lactate administration aimed to improve brain
380 function and reduce damage associated with head trauma. However, they differed since systemic
381 injection involves administering lactate intravenously and therefore distributing the compound
382 throughout the body. The objective of this method is to provide an additional energy substrate to
383 the entire body including brain cells. Lactate can be used by neurons to produce energy, even
384 when brain metabolism is disrupted by head trauma. In contrast, cerebral lactate injection involves
385 directly administering lactate into the brain, usually by microdialysis or intracerebral injection. This
386 method helps to deliver a high concentration of lactate directly to brain cells. Currently, there is
387 no clear consensus on the optimal method of administration of lactate.

388 Ketone bodies can be administered through various formulations. Ketogenic diets are based on
389 carbohydrate deprivation, either by fasting or by enteral formulas containing medium-chain
390 triglycerides and low amounts of carbohydrates. An exclusive ketogenic diet such as used to treat
391 epilepsy in children takes 3 to 5 days to reach therapeutic blood levels. These delays render the
392 ketogenic diet of limited interest in acute neurocritical care. Moreover, ketogenic diets are difficult
393 to prepare and compliance is a challenge.⁵⁷ Ketone bodies may also be infused as intravenous
394 sodium BHB or sodium acetoacetate solutions. Finally, enteral ketone bodies are now available
395 in the form of ketone esters (such as 1,3-butanediol monoester of beta-hydroxybutyrate or
396 glyceryl-tris-3-hydroxybutyrate, two precursors of ketone bodies). The compound (R)-3-
397 hydroxybutyl (R)-3-hydroxybutyrate is now considered a safe food supplement for humans by the
398 U.S. Food and Drug Administration. Ketone esters induce a rapid and significant ketosis after a
399 single oral intake and as such, represent an interesting therapeutic avenue in acute care.^{57,58}

400

401 **Clinical trials**

402 Based on such preclinical evidence, the idea of providing fuel support to brain-injured patients
403 progressively emerged and a few clinical trials started. Bouzat *et al.* were the first to infuse
404 hypertonic lactate in patients with severe TBI undergoing CMD monitoring. They observed a
405 significant increase in extracellular cerebral glucose.⁴⁹ Overall, these data suggest that both
406 endogenous and exogenous lactate are associated with an increased cerebral glucose
407 availability. This “excess” glucose may then be redirected for other metabolic purposes, such as
408 the neuronal PPP.⁵⁶

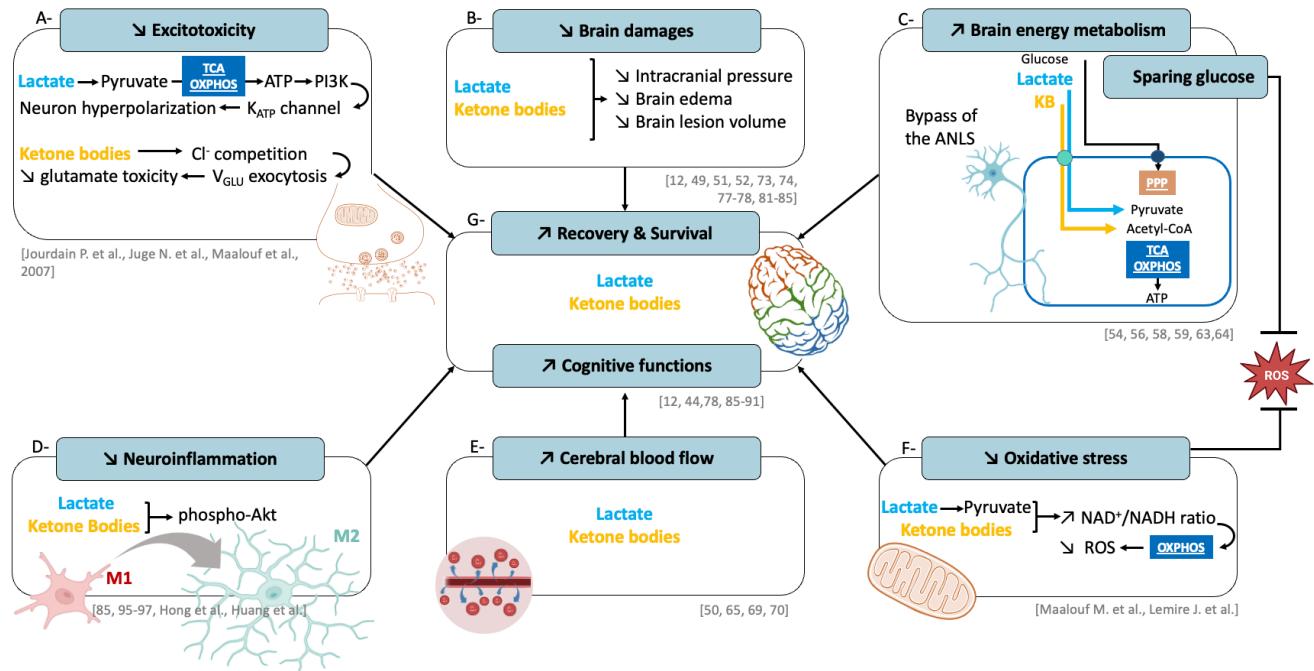
409 In human, clinical trials of lactate administration after TBI or SAH....Developer un peu plus les
410 essais cliniques TBI, SAH et lactate et what about KB ou ester de cétones ?

411

412 **Conclusion and Perspectives**

413 After being considered as unwanted metabolic wastes, lactate and KB are now widely viewed as
414 integral parts of brain metabolism. Data accumulating over the last thirty years support their
415 important role in energy production during rest states, as well as alternative substrates in case of
416 glucose shortage or unavailability. They have also been tested as therapeutics in animal models
417 and human cohorts with acute brain injury with beneficial results concerning excitotoxicity,
418 neuroinflammation, cerebral blood flow control, reduction of brain lesion volume, improvement of
419 cognitive function and survival (**Figure 3**). In addition, KB and lactate share a common
420 mechanism: modulation of brain metabolism. Indeed, when administered, lactate and ketone
421 bodies exert a glucose-sparing effect leading to neuronal glycolytic regulation and redirection of
422 glucose through the PPP which helps to fight against ROS produced during oxidative stress
423 (**Figure 3**). Overall, data in animal models and small human cohorts call for a confirmation of their
424 benefits in large therapeutic trials targeting acute neurocritical care populations. After decades of

425 preclinical research and encouraging new clinical data, we are at the dawn of a new period during
 426 which lactate and ketone bodies might become an integral part of neurocritical care therapeutics.



427

428 **Figure 3: Neuroprotection by exogenous lactate and ketone bodies, as alternative energy substrates,**
 429 **in a context of brain injury.** A- Lactate counteracts glutamate excitotoxicity through the PI3K pathway
 430 leading to neuronal hyperpolarization. Ketone bodies also decrease excitotoxicity through a competition
 431 with Cl⁻, responsible of a decrease in v_{GLU} exocytosis. B- Both exogenous lactate and ketone bodies limit
 432 brain damages by decreasing intracranial pressure, brain edema as well as brain lesion volume. C- Lactate
 433 and ketone bodies increase brain energy metabolism by providing respectively pyruvate and acetyl-CoA to
 434 feed the TCA cycle and OXPHOS. D- Both alternative substrates also switch microglia from an inflammatory
 435 M1 to a anti-inflammatory M2 phenotype, through a stimulation of the Akt pathway. E- Lactate and ketone
 436 bodies increase cerebral blood flow. F- Both lactate and ketone bodies regulate the oxidative stress by
 437 increasing the NAD⁺/NADH ratio. G- Altogether, beneficial effects of exogenous lactate and ketone bodies
 438 lead to increase recovery and survival, as well as improvement of cognitive functions.
 439 ANLS, astrocyte-neurone lactate shuttle; OXPHOS, oxidative phosphorylation; PPP, pentose phosphate
 440 pathway; ROS, reactive oxygen species; TCA, tricarboxylic acid; v_{GLU}, vesicular glutamate transporters.
 441

442 Disclosure/conflict of interest

443 All authors have no disclosures or conflict of interests.

444 References

- 446 1. Oddo M, Villa F, Citerio G. Brain multimodality monitoring: an update. *Curr Opin Crit Care*.
447 Apr 2012;18(2):111-8.

448 2. Bouzat P, Sala N, Payen JF, Oddo M. Beyond intracranial pressure: optimization of
449 cerebral blood flow, oxygen, and substrate delivery after traumatic brain injury. *Ann*
450 *Intensive Care. ul* 2013;3(1):23.

451 3. Hermanides J, Hong YT, Trivedi M, Outtrim J, Aigbirhio F, Nestor PJ, et al. Metabolic
452 derangements are associated with impaired glucose delivery following traumatic brain
453 injury. *Brain J Neurol. Dec 2021;144(11):3492-504.*

454 4. Patet C, Quintard H, Suys T, Bloch J, Daniel RT, Pellerin L, et al. Neuroenergetic
455 Response to prolonged cerebral glucose depletion after severe brain injury and the role of
456 lactate. *J Neurotrauma. Oct 2015;32(20):1560-6.*

457 5. Robbins NM, Swanson RA. Opposing effects of glucose on stroke and reperfusion injury:
458 acidosis, oxidative stress, and energy metabolism. *Stroke. jun2014;45(6):1881-6.*

459 6. Attwell D, Buchan AM, Charpak S, Lauritzen M, MacVicar BA, Newman EA. Glial and
460 neuronal control of brain blood flow. *Nature. Nov 2010;468(7321):232-43.*

461 7. Herrero-Mendez A, Almeida A, Fernández E, Maestre C, Moncada S, Bolaños JP. The
462 bioenergetic and antioxidant status of neurons is controlled by continuous degradation of a
463 key glycolytic enzyme by APC/C-Cdh1. *Nat Cell Biol. Jun 2009;11(6):747-52.*

464 8. Oddo M, Schmidt JM, Mayer SA, Chioléro RL. Glucose control after severe brain injury.
465 *Curr Opin Clin Nutr Metab Care. Mar 2008;11(2):134.*

466 9. Carre E, Ogier M, Boret H, Montcriol A, Bourdon L, Risso JJ. Metabolic crisis in severely
467 head-injured patients: Is ischemia just the tip of the iceberg? *Front Neurol [Internet]. 2013*
468 [Jul 2023];4.

- 469 10. Pulsinelli WA, Waldman S, Rawlinson D, Plum F. Moderate hyperglycemia augments
470 ischemic brain damage: a neuropathologic study in the rat. Neurology. Nov
471 1982;32(11):1239-46.
- 472 11. MacDougall NJJ, Muir KW. Hyperglycaemia and infarct size in animal models of middle
473 cerebral artery occlusion: systematic review and meta-analysis. J Cereb Blood Flow Metab
474 Off J Int Soc Cereb Blood Flow Metab. Mar 2011;31(3):807-18.
- 475 12. Roumes H, Dumont U, Sanchez S, Mazuel L, Blanc J, Raffard G, et al. Neuroprotective
476 role of lactate in rat neonatal hypoxia-ischemia. J Cereb Blood Flow Metab. Feb
477 2021;41(2):342-58.
- 478 13. Badjatia N, Topcuoglu MA, Buonanno FS, Smith EE, Nogueira RG, Rordorf GA, et al.
479 Relationship between hyperglycemia and symptomatic vasospasm after subarachnoid
480 hemorrhage. Crit Care Med. Jul 2005;33(7):1603-9; quiz 1623.
- 481 14. Bellolio MF, Gilmore RM, Ganti L. Insulin for glycaemic control in acute ischaemic stroke.
482 Cochrane Database Syst Rev. an 2014;(1):CD005346.
- 483 15. Piironen K, Putala J, Rosso C, Samson Y. Glucose and Acute Stroke. Stroke. Mar
484 2012;43(3):898-902.
- 485 16. Van den Berghe G, Wouters P, Weekers F, Verwaest C, Bruyninckx F, Schetz M, et al.
486 Intensive Insulin Therapy in Critically Ill Patients. N Engl J Med. ov 2001;345(19):1359-67.
- 487 17. Gray CS, Hildreth AJ, Sandercock PA, O'Connell JE, Johnston DE, Cartlidge NEF, et al.
488 Glucose-potassium-insulin infusions in the management of post-stroke hyperglycaemia:
489 the UK Glucose Insulin in Stroke Trial (GIST-UK). Lancet Neurol. mai 2007;6(5):397-406.
- 490 18. Hermanides J, Plummer MP, Finnis M, Deane AM, Coles JP, Menon DK. Glycaemic
491 control targets after traumatic brain injury: a systematic review and meta-analysis. Crit
492 Care Lond Engl. Jan 2018;22(1):11.
- 493 19. Vespa P, Bergsneider M, Hattori N, Wu HM, Huang SC, Martin NA, et al. Metabolic crisis

- 494 without brain ischemia is common after traumatic brain injury: A combined microdialysis
495 and positron emission tomography study. *J Cereb Blood Flow Metab.* Jun
496 2005;25(6):763-74.
- 497 20. Venturini S, Bhatti F, Timofeev I, Carpenter KLH, Hutchinson PJ, Guilfoyle MR, et al.
498 Microdialysis-based classifications of abnormal metabolic states after traumatic brain
499 injury: A systematic review of the literature. *J Neurotrauma.* Feb 2023;40(3-4):195-209.
- 500 21. Stein NR, McArthur DL, Etchepare M, Vespa PM. Early cerebral metabolic crisis after TBI
501 influences outcome despite adequate hemodynamic resuscitation. *Neurocrit Care.* Aug
502 2012;17(1):49-57.
- 503 22. Abi-Saab WM, Maggs DG, Jones T, Jacob R, Srihari V, Thompson J, et al. Striking
504 differences in glucose and lactate levels between brain extracellular fluid and plasma in
505 conscious human subjects: effects of hyperglycemia and hypoglycemia. *J Cereb Blood
506 Flow Metab Off J Int Soc Cereb Blood Flow Metab.* Mar 2002;22(3):271-9.
- 507 23. Vespa PM, McArthur D, O'Phelan K, Glenn T, Etchepare M, Kelly D, et al. Persistently low
508 extracellular glucose correlates with poor outcome 6 months after human traumatic brain
509 injury despite a lack of increased lactate: a microdialysis study. *J Cereb Blood Flow Metab
510 Off J Int Soc Cereb Blood Flow Metab.* Jul 2003;23(7):865-77.
- 511 24. Stovell MG, Helmy A, Thelin EP, Jalloh I, Hutchinson PJ, Carpenter KLH. An overview of
512 clinical cerebral microdialysis in acute brain injury. *Front Neurol [Internet].* 2023 [juill
513 2023];14.
- 514 25. Boumezbeur F, Petersen KF, Cline GW, Mason GF, Behar KL, Shulman GI, et al. The
515 contribution of blood lactate to brain energy metabolism in humans measured by dynamic
516 ¹³C nuclear magnetic resonance spectroscopy. *J Neurosci Off J Soc Neurosci.* Oct
517 2010;30(42):13983-91.
- 518 26. Owen OE, Morgan AP, Kemp HG, Sullivan JM, Herrera MG, Cahill GF. Brain metabolism

- 519 during fasting*. J Clin Invest. Oct 1967;46(10):1589-95.
- 520 27. Patet C, Suys T, Carteron L, Oddo M. Cerebral lactate metabolism after traumatic brain
521 injury. Curr Neurol Neurosci Rep. Feb 2016;16(4):31.
- 522 28. White H, Venkatesh B. Clinical review: Ketones and brain injury. Crit Care. 2011;15(2):219.
- 523 29. Pellerin L, Magistretti PJ. Glutamate uptake into astrocytes stimulates aerobic glycolysis: a
524 mechanism coupling neuronal activity to glucose utilization. Proc Natl Acad Sci. Oct
525 1994;91(22):10625-9.
- 526 30. Almeida A, Jimenez-Blasco D, Bolaños JP. Cross-talk between energy and redox
527 metabolism in astrocyte-neuron functional cooperation. Bolaños JP, éditeur. Essays
528 Biochem. Mar 2023;67(1):17-26.
- 529 31. Bak LK, Walls AB. CrossTalk opposing view: lack of evidence supporting an astrocyte-to-
530 neuron lactate shuttle coupling neuronal activity to glucose utilisation in the brain. J
531 Physiol. Feb 2018;596(3):351-3.
- 532 32. Barros LF, Weber B. CrossTalk proposal: an important astrocyte-to-neuron lactate shuttle
533 couples neuronal activity to glucose utilisation in the brain. J Physiol. Feb
534 2018;596(3):347-50.
- 535 33. Barros LF, Weber B. Rebuttal from L. F. Barros and B. Weber. J Physiol.
536 Feb 2018;596(3):355-6.
- 537 34. Pellerin L, Bouzier-Sore A, Aubert A, Serres S, Merle M, Costalat R, et al. Activity-
538 dependent regulation of energy metabolism by astrocytes: An update. Glia. Sept
539 2007;55(12):1251-62.
- 540 35. Roumes H, Pellerin L, Bouzier-Sore AK. Astrocytes as metabolic suppliers to support
541 neuronal activity and brain functions. Bolaños JP, éditeur. Essays Biochem. Mar
542 2023;67(1):27-37.
- 543 36. Fletcher WM, Hopkins FG. Lactic acid in amphibian muscle1. J Physiol. Mar

- 544 1907;35(4):247-309.
- 545 37. Wyss MT, Jolivet R, Buck A, Magistretti PJ, Weber B. In vivo evidence for lactate as a
546 neuronal energy source. *J Neurosci Off J Soc Neurosci*. May 2011;31(20):7477-85.
- 547 38. Ide K, Schmalbruch IK, Quistorff B, Horn A, Secher NH. Lactate, glucose and O₂ uptake in
548 human brain during recovery from maximal exercise. *J Physiol*. Jan 2000;522(Pt
549 1):159-64.
- 550 39. van Hall G, Strømstad M, Rasmussen P, Jans O, Zaar M, Gam C, et al. Blood lactate is an
551 important energy source for the human brain. *J Cereb Blood Flow Metab Off J Int Soc*
552 *Cereb Blood Flow Metab*. Jun 2009;29(6):1121-9.
- 553 40. Oddo M, Levine JM, Frangos S, Maloney-Wilensky E, Carrera E, Daniel RT, et al. Brain
554 lactate metabolism in humans with subarachnoid hemorrhage. *stroke*. May
555 2012;43(5):1418-21.
- 556 41. Schurr A, Payne RS, Miller JJ, Tseng MT, Rigor BM. Blockade of lactate transport
557 exacerbates delayed neuronal damage in a rat model of cerebral ischemia. *Brain Res*. 23
558 Mar 2001;895(1-2):268-72.
- 559 42. Guzmán M, Blázquez C. Is there an astrocyte-neuron ketone body shuttle? *Trends*
560 *Endocrinol Metab TEM*. 2001;12(4):169-73.
- 561 43. Cox PJ, Kirk T, Ashmore T, Willerton K, Evans R, Smith A, et al. Nutritional ketosis alters
562 fuel preference and thereby endurance performance in athletes. *Cell Metab*. Aug
563 2016;24(2):256-68.
- 564 44. Oddo M, Vespa P, Menon DK. Boosting the injured brain with supplemental energy fuels.
565 *Intensive Care Med*. Jun 2019;45(6):872-5.
- 566 45. Xin L, Ipek Ö, Beaumont M, Shevlyakova M, Christinat N, Masoodi M, et al. Nutritional
567 ketosis increases NAD+/NADH ratio in healthy human brain: An in vivo study by 31P-MRS.
568 *Front Nutr [Internet]*. 2018 [juin 2023];5.

- 569 46. Poff AM, Moss S, Soliven M, D'Agostino DP. Ketone supplementation: Meeting the needs
570 of the brain in an energy crisis. *Front Nutr* [Internet]. 2021 [juin 2023];8: 47. van Gemert
571 LA, de Galan BE, Wevers RA, Ter Heine R, Willemse MA. Lactate infusion as
572 therapeutical intervention: a scoping review. *Eur J Pediatr.* Jun 2022;181(6):2227-35.
- 573 48. Bisri T, Utomo B, Fuadi I. Exogenous lactate infusion improved neurocognitive function of
574 patients with mild traumatic brain injury. *Asian J Neurosurg.* Jun 2016;11(02):151-9.
- 575 49. Bouzat P, Sala N, Suys T, Zerlauth JB, Marques-Vidal P, Feihl F, et al. Cerebral metabolic
576 effects of exogenous lactate supplementation on the injured human brain. *Intensive Care
577 Med.* Mar 2014;40(3):412-21.
- 578 50. Carteron L, Solari D, Patet C, Quintard H, Miroz JP, Bloch J, et al. Hypertonic lactate to
579 improve cerebral perfusion and glucose availability after acute brain injury. *Crit Care Med.*
580 Oct 2018;46(10):1649-55.
- 581 51. Ichai C, Armando G, Orban JC, Berthier F, Rami L, Samat-Long C, et al. Sodium lactate
582 versus mannitol in the treatment of intracranial hypertensive episodes in severe traumatic
583 brain-injured patients. *Intensive Care Med.* Mar2009;35(3):471-9.
- 584 52. Ichai C, Payen JF, Orban JC, Quintard H, Roth H, Legrand R, et al. Half-molar sodium
585 lactate infusion to prevent intracranial hypertensive episodes in severe traumatic brain
586 injured patients: a randomized controlled trial. *Intensive Care Med.*
587 Aug2013;39(8):1413-22.
- 588 53. Quintard H, Patet C, Zerlauth JB, Suys T, Bouzat P, Pellerin L, et al. Improvement of
589 neuroenergetics by hypertonic lactate therapy in patients with traumatic brain injury is
590 dependent on baseline cerebral lactate/pyruvate ratio. *J Neurotrauma.* Apr
591 2016;33(7):681-7.
- 592 54. Wolahan SM, Mao HC, Real C, Vespa PM, Glenn TC. Lactate supplementation in severe
593 traumatic brain injured adults by primed constant infusion of sodium L-lactate. *J Neurosci*

- 594 Res. Apr 2018;96(4):688-95.
- 595 55. Gallagher CN, Carpenter KLH, Grice P, Howe DJ, Mason A, Timofeev I, et al. The human
596 brain utilizes lactate via the tricarboxylic acid cycle: a ¹³C-labelled microdialysis and high-
597 resolution nuclear magnetic resonance study. Brain. Oct 2009;132(10):2839-49.
- 598 56. Jalloh I, Helmy A, Shannon RJ, Gallagher CN, Menon DK, Carpenter KLH, et al. Lactate
599 uptake by the injured human brain: Evidence from an arteriovenous gradient and cerebral
600 microdialysis study. J Neurotrauma. Dec 2013;30(24):2031-7.
- 601 57. Clarke K, Tchabanenko K, Pawlosky R, Carter E, Todd King M, Musa-Veloso K, et al.
602 Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult
603 subjects. Regul Toxicol Pharmacol RTP. Aug2012;63(3):401-8.
- 604 58. Suissa L, Kotchetkov P, Guigonis JM, Doche E, Osman O, Pourcher T, et al. Ingested
605 ketone ester leads to a rapid rise of acetyl-CoA and competes with glucose metabolism in
606 the brain of non-fasted mice. Int J Mol Sci. Jan 2021;22(2):524.
- 607 59. Glenn TC, Kelly DF, Boscardin WJ, McArthur DL, Vespa P, Oertel M, et al. Energy
608 Dysfunction as a predictor of outcome after moderate or severe head injury: Indices of
609 oxygen, glucose, and lactate metabolism. J Cereb Blood Flow Metab. Oct
610 2003;23(10):1239-50.
- 611 60. Bouzier AK, Thiaudiere E, Biran M, Rouland R, Canioni P, Merle M. The Metabolism of [3-
612 ¹³C]lactate in the rat brain is specific of a pyruvate carboxylase-deprived compartment. J
613 Neurochem. 2000;75(2):480-6.
- 614 61. Hyacinthe JN, Buscemi L, Lê TP, Lepore M, Hirt L, Mishkovsky M. Evaluating the potential
615 of hyperpolarised [1-¹³C] L-lactate as a neuroprotectant metabolic biosensor for stroke.
616 Sci Rep. Mar 2020;10(1):5507.
- 617 62. Drenick EJ, Alvarez LC, Tamasi GC, Brickman AS. Resistance to symptomatic insulin
618 reactions after fasting. J Clin Invest. Oct1972;51(10):2757-62.

- 619 63. Bouzier-Sore AK, Voisin P, Canioni P, Magistretti PJ, Pellerin L. Lactate is a preferential
620 oxidative energy substrate over glucose for neurons in culture. *J Cereb Blood Flow Metab.*
621 Nov 2003;23(11):1298-306.
- 622 64. Bouzier-Sore AK, Voisin P, Bouchaud V, Bezancon E, Franconi JM, Pellerin L.
623 Competition between glucose and lactate as oxidative energy substrates in both neurons
624 and astrocytes: a comparative NMR study. *Eur J Neurosci.* 2006;24(6):1687-94.
- 625 65. Svart M, Gormsen LC, Hansen J, Zeidler D, Gejl M, Vang K, et al. Regional cerebral
626 effects of ketone body infusion with 3-hydroxybutyrate in humans: Reduced glucose
627 uptake, unchanged oxygen consumption and increased blood flow by positron emission
628 tomography. A randomized, controlled trial. *PLoS One.* 2018;13(2):e0190556.
- 629 66. Horowitz T, Doche E, Philip M, Cammilleri S, Suissa L, Guedj E. Regional brain glucose
630 metabolism is differentially affected by ketogenic diet: a human semiquantitative positron
631 emission tomography. *Eur J Nucl Med Mol Imaging.* juin 2023;50(7):2047-55.
- 632 67. Hutchinson PJ, O'Connell MT, Seal A, Nortje J, Timofeev I, Al-Rawi PG, et al. A combined
633 microdialysis and FDG-PET study of glucose metabolism in head injury. *Acta Neurochir*
634 (*Wien*). 1 janv 2009;151(1):51-61.
- 635 68. Gordon GRJ, Choi HB, Rungta RL, Ellis-Davies GCR, MacVicar BA. Brain metabolism
636 dictates the polarity of astrocyte control over arterioles. *Nature.* 11 déc
637 2008;456(7223):745-9.
- 638 69. Ma D, Wang AC, Parikh I, Green SJ, Hoffman JD, Chlipala G, et al. Ketogenic diet
639 enhances neurovascular function with altered gut microbiome in young healthy mice. *Sci*
640 *Rep.* 27 avr 2018;8(1):6670.
- 641 70. Hasselbalch SG, Madsen PL, Hageman LP, Olsen KS, Justesen N, Holm S, et al.
642 Changes in cerebral blood flow and carbohydrate metabolism during acute
643 hyperketonemia. *Am J Physiol-Endocrinol Metab.* 1 mai 1996;270(5):E746-51.

- 644 71. Carney N, Totten AM, O'Reilly C, Ullman JS, Hawryluk GWJ, Bell MJ, et al. Guidelines for
645 the Management of Severe Traumatic Brain Injury, Fourth Edition. Neurosurgery. 1 janv
646 2017;80(1):6-15.
- 647 72. Anstey JR, Taccone FS, Udy AA, Citerio G, Duranteau J, Ichai C, et al. Early Osmotherapy
648 in Severe Traumatic Brain Injury: An International Multicenter Study. J Neurotrauma. 1 janv
649 2020;37(1):178-84.
- 650 73. Bernini A, Miroz JP, Abed-Maillard S, Favre E, Iaquaniello C, Ben-Hamouda N, et al.
651 Hypertonic lactate for the treatment of intracranial hypertension in patients with acute brain
652 injury. Sci Rep. 22 févr 2022;12(1):3035.
- 653 74. Ichai C, Orban JC, Fontaine E. Sodium lactate for fluid resuscitation: the preferred solution
654 for the coming decades? Crit Care Lond Engl. 7 juill 2014;18(4):163.
- 655 75. Duhaut DE, Heurteaux C, Gandin C, Ichai C, Quintard H. The Antiedematous Effect of
656 Exogenous Lactate Therapy in Traumatic Brain Injury: A Physiological and Mechanistic
657 Approach. Neurocrit Care. déc 2021;35(3):747-55.
- 658 76. Millet A, Cuisinier A, Bouzat P, Batandier C, Lemasson B, Stupar V, et al. Hypertonic
659 sodium lactate reverses brain oxygenation and metabolism dysfunction after traumatic
660 brain injury. Br J Anaesth. juin 2018;120(6):1295-303.
- 661 77. Alessandri B, Schwandt E, Kamada Y, Nagata M, Heimann A, Kempski O. The
662 Neuroprotective Effect of Lactate Is Not Due to Improved Glutamate Uptake after
663 Controlled Cortical Impact in Rats. J Neurotrauma. 10 août 2012;29(12):2181-91.
- 664 78. Berthet C, Lei H, Thevenet J, Gruetter R, Magistretti PJ, Hirt L. Neuroprotective role of
665 lactate after cerebral ischemia. J Cereb Blood Flow Metab Off J Int Soc Cereb Blood Flow
666 Metab. nov 2009;29(11):1780-9.
- 667 79. Roumes H, Pellerin L, Bouzier-Sore AK. Effet neuroprotecteur du lactate dans l'hypoxie-
668 ischémie cérébrale néonatale. médecine/sciences. nov 2020;36(11):973-6.

- 669 80. Tassinari ID, Andrade MKG, da Rosa LA, Hoff MLM, Nunes RR, Vogt EL, et al. Lactate
670 Administration Reduces Brain Injury and Ameliorates Behavioral Outcomes Following
671 Neonatal Hypoxia–Ischemia. *Neuroscience*. 10 nov 2020;448:191-205.
- 672 81. Suzuki M, Suzuki M, Kitamura Y, Mori S, Sato K, Dohi S, et al. β -Hydroxybutyrate, a
673 Cerebral Function Improving Agent, Protects Rat Brain Against Ischemic Damage Caused
674 by Permanent and Transient Focal Cerebral Ischemia. *Jpn J Pharmacol*. 2002;89(1):36-43.
- 675 82. Prins ML, Sm L, Ls F, Da H. Increased cerebral uptake and oxidation of exogenous
676 betaHB improves ATP following traumatic brain injury in adult rats. *J Neurochem* [Internet].
677 août 2004 [cité 26 juin 2023];90(3). Disponible sur:
678 <https://pubmed.ncbi.nlm.nih.gov/15255945/>
- 679 83. Prins ML, Fujima LS, Hovda DA. Age-dependent reduction of cortical contusion volume by
680 ketones after traumatic brain injury. *J Neurosci Res*. 1 nov 2005;82(3):413-20.
- 681 84. Camberos-Luna L, Gerónimo-Olvera C, Montiel T, Rincon-Heredia R, Massieu L. The
682 Ketone Body, β -Hydroxybutyrate Stimulates the Autophagic Flux and Prevents Neuronal
683 Death Induced by Glucose Deprivation in Cortical Cultured Neurons. *Neurochem Res*.
684 mars 2016;41(3):600-9.
- 685 85. Lin C, Wang S, Xie J, Zhu J, Xu J, Liu K, et al. Ketogenic diet and β -Hydroxybutyrate
686 alleviate ischemic brain injury in mice via an IRAKM-dependent pathway. *Eur J Pharmacol*.
687 20 juill 2023;955:175933.
- 688 86. Schurr A, Payne RS, Miller JJ, Rigor BM. Brain lactate, not glucose, fuels the recovery of
689 synaptic function from hypoxia upon reoxygenation: an in vitro study. *Brain Res*. 2 janv
690 1997;744(1):105-11.
- 691 87. Berthet C, Castillo X, Magistretti PJ, Hirt L. New Evidence of Neuroprotection by Lactate
692 after Transient Focal Cerebral Ischaemia: Extended Benefit after Intracerebroventricular
693 Injection and Efficacy of Intravenous Administration. *Cerebrovasc Dis*. 14 nov
694 2012;34(5-6):329-35.

- 695 88. Crespy T, Durost M, Fricault P, Lemasson B, Bouzat P, Barbier EL, et al. Hypertonic
696 Sodium Lactate to Alleviate Functional Deficits Following Diffuse Traumatic Brain Injury:
697 An Osmotic or a Lactate-Related Effect? *Neurocrit Care.* juin 2021;34(3):795-803.
- 698 89. Rice AC, Zsoldos R, Chen T, Wilson MS, Alessandri B, Hamm RJ, et al. Lactate
699 administration attenuates cognitive deficits following traumatic brain injury. *Brain Res.* 22
700 févr 2002;928(1-2):156-9.
- 701 90. Gambardella I, Ascione R, D'Agostino DP, Ari C, Worku B, Tranbaugh RF, et al.
702 Systematic Review - Neuroprotection of ketosis in acute injury of the mammalian central
703 nervous system: A meta-analysis. *J Neurochem.* 2021;158(2):105-18.
- 704 91. Gibson CL, Murphy AN, Murphy SP. Stroke outcome in the ketogenic state - a systematic
705 review of the animal data. *J Neurochem.* nov 2012;123(0 2):52-7.
- 706 92. Rodriguez-Rodriguez P, Fernandez E, Bolaños JP. Underestimation of the Pentose–
707 Phosphate Pathway in Intact Primary Neurons as Revealed by Metabolic Flux Analysis. *J*
708 *Cereb Blood Flow Metab.* 1 déc 2013;33(12):1843-5.
- 709 93. Augusto-Oliveira M, Arrifano GP, Lopes-Araújo A, Santos-Sacramento L, Takeda PY,
710 Anthony DC, et al. What Do Microglia Really Do in Healthy Adult Brain? *Cells.* 22 oct
711 2019;8(10):1293.
- 712 94. Ma Y, Wang J, Wang Y, Yang GY. The biphasic function of microglia in ischemic stroke.
713 *Prog Neurobiol.* oct 2017;157:247-72.
- 714 95. Monsorno K, Buckinx A, Paolicelli RC. Microglial metabolic flexibility: emerging roles for
715 lactate. *Trends Endocrinol Metab TEM.* mars 2022;33(3):186-95.
- 716 96. Huang C, Wang P, Xu X, Zhang Y, Gong Y, Hu W, et al. The ketone body metabolite β -
717 hydroxybutyrate induces an antidepression-associated ramification of microglia via HDACs
718 inhibition-triggered Akt-small RhoGTPase activation. *Glia.* févr 2018;66(2):256-78.
- 719 97. Guo M, Wang X, Zhao Y, Yang Q, Ding H, Dong Q, et al. Ketogenic Diet Improves Brain

720 Ischemic Tolerance and Inhibits NLRP3 Inflammasome Activation by Preventing Drp1-
721 Mediated Mitochondrial Fission and Endoplasmic Reticulum Stress. *Front Mol Neurosci.* 20
722 mars 2018;11:86.
723
724

725 **Figure legend**

726

727

728

729

730

- 731 Scafidi, S., J. Jernberg, G. Fiskum and M. C. McKenna (2022) Metabolism of Exogenous [2,4-
732 (13)C]beta-Hydroxybutyrate following Traumatic Brain Injury in 21-22-Day-Old Rats: An Ex Vivo
733 NMR Study. **Metabolites** 12(8).10.3390/metabo12080710
- 734 Roumes, H., U. Dumont, S. Sanchez, L. Mazuel, J. Blanc, G. Raffard, J. F. Chateil, L. Pellerin
735 and A. K. Bouzier-Sore (2021) Neuroprotective role of lactate in rat neonatal hypoxia-ischemia.
736 **J Cereb Blood Flow Metab** 41(2): 342-358.10.1177/0271678X20908355
- 737 Roumes, H., C. Jolle, J. Blanc, I. Benkhaled, C. P. Chatain, P. Massot, G. Raffard, V.
738 Bouchaud, M. Biran, C. Pythoud, N. Deglon, E. R. Zimmer, L. Pellerin and A. K. Bouzier-Sore
739 (2021) Lactate transporters in the rat barrel cortex sustain whisker-dependent BOLD fMRI signal
740 and behavioral performance. **Proc Natl Acad Sci U S A** 118(47).10.1073/pnas.2112466118
- 741 Netzahualcoyotzi, C. and L. Pellerin (2020) Neuronal and astroglial monocarboxylate
742 transporters play key but distinct roles in hippocampus-dependent learning and memory
743 formation. **Prog Neurobiol** 194: 101888.10.1016/j.pneurobio.2020.101888
- 744 Li, Z., B. Zhang, W. Yao, C. Zhang, L. Wan and Y. Zhang (2019) APC-Cdh1 Regulates
745 Neuronal Apoptosis Through Modulating Glycolysis and Pentose-Phosphate Pathway After
746 Oxygen-Glucose Deprivation and Reperfusion. **Cellular and Molecular Neurobiology** 39(1):
747 123-135.10.1007/s10571-018-0638-x
- 748 Bak, L. K. and A. B. Walls (2018) CrossTalk opposing view: lack of evidence supporting an
749 astrocyte-to-neuron lactate shuttle coupling neuronal activity to glucose utilisation in the brain. **J**
750 **Physiol** 596(3): 351-353.10.1113/JP274945
- 751 Barros, L. F. and B. Weber (2018) CrossTalk proposal: an important astrocyte-to-neuron lactate
752 shuttle couples neuronal activity to glucose utilisation in the brain. **J Physiol** 596(3): 347-
753 350.10.1113/JP274944
- 754 Bolanos, J. P. (2016) Bioenergetics and redox adaptations of astrocytes to neuronal activity. **J**
755 **Neurochem** 139 Suppl 2: 115-125.10.1111/jnc.13486
- 756 Duarte, J. M., F. M. Girault and R. Gruetter (2015) Brain energy metabolism measured by (13)C
757 magnetic resonance spectroscopy in vivo upon infusion of [3-(13)C]lactate. **J Neurosci Res**
758 93(7): 1009-1018.10.1002/jnr.23531
- 759 Roy, M., M. C. Beauvieux, J. Naulin, D. El Hamrani, J. L. Gallis, S. C. Cunnane and A. K.
760 Bouzier-Sore (2015) Rapid adaptation of rat brain and liver metabolism to a ketogenic diet: an
761 integrated study using (1)H- and (13)C-NMR spectroscopy. **J Cereb Blood Flow Metab** 35(7):
762 1154-1162.10.1038/jcbfm.2015.29
- 763 Suzuki, A., S. A. Stern, O. Bozdagi, G. W. Huntley, R. H. Walker, P. J. Magistretti and C. M.
764 Alberini (2011) Astrocyte-neuron lactate transport is required for long-term memory formation.
765 **Cell** 144(5): 810-823.S0092-8674(11)00132-2 [pii]
766 10.1016/j.cell.2011.02.018
- 767 Herrero-Mendez, A., A. Almeida, E. Fernandez, C. Maestre, S. Moncada and J. P. Bolanos
768 (2009) The bioenergetic and antioxidant status of neurons is controlled by continuous

769 degradation of a key glycolytic enzyme by APC/C-Cdh1. **Nat Cell Biol** 11(6): 747-
770 752.10.1038/ncb1881
771 Pan, J. W., R. A. de Graaf, K. F. Petersen, G. I. Shulman, H. P. Hetherington and D. L.
772 Rothman (2002) [2,4-13 C2]-beta-Hydroxybutyrate metabolism in human brain. **J Cereb Blood
773 Flow Metab** 22(7): 890-898.10.1097/00004647-200207000-00014
774 Bouzier, A. K., E. Thiaudiere, M. Biran, R. Roulard, P. Canioni and M. Merle (2000) The
775 metabolism of [3-(13)C]lactate in the rat brain is specific of a pyruvate carboxylase-deprived
776 compartment. **J Neurochem** 75(2): 480-486
777 Pellerin, L. and P. J. Magistretti (1994) Glutamate uptake into astrocytes stimulates aerobic
778 glycolysis: a mechanism coupling neuronal activity to glucose utilization. **Proc Natl Acad Sci U
779 S A** 91(22): 10625-10629.10.1073/pnas.91.22.10625
780 Swanson, R. A., S. M. Sagar and F. R. Sharp (1989) Regional brain glycogen stores and
781 metabolism during complete global ischaemia. **Neurol Res** 11(1): 24-
782 28.10.1080/01616412.1989.11739856
783