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1	Crystallization of emulsified anhydrous milk fat: the role of confinement and of
2	minor compounds. A DSC study
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20	
21	Abbreviations: AMF Anhydrous Milk Fat; DSC Differential Scanning Calorimetry; FA Fatty
22	Acid; p-NMR pulsed Nuclear Magnetic Resonance; SC Sodium Caseinate; SDS Sodium
23	Dodecyl Sulfate; SFC solid fat content; TAG TriAcylGlycerol; XRD X-ray diffraction; 3:0
24	propionic acid; 18:1 oleic acid; 16:0 palmitic acid; PPP tripalmitin

Abstract

We examined the crystallization and melting of anhydrous milk fat (AMF)-in-water 26 emulsions stabilized by sodium caseinate. Various additives at low concentrations (< 5 27 wt%), differing in their hydrocarbon chain length (propionic vs. palmitic acid), unsaturation 28 29 (palmitic vs. oleic acid), and esterification state (palmitic acid vs. tripalmitin) were used to modulate AMF crystallization kinetics. Three emulsions with different average droplet 30 diameters were cooled down from 60 °C to 4 °C. Fat crystallization was followed by DSC 31 32 under dynamic (cooling) and static (isothermal) conditions. Propionic acid did not have any noticeable effect. Oleic acid favored supercooling and the formation of unstable 33 polymorphs at short times but its impact faded after 48 h of isothermal storage. The impact 34 of palmitic acid was related to its amphiphilic properties and vanished after 48 h. 35 Tripalmitin influenced crystallization via volume effects that were persistent. It formed 36 37 mixed crystals which extended the melting range of AMF.

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40 **1. Introduction**

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Numerous food products are based on oil-in-water emulsions (O/W) (salad sauces, 42 milk...) or water-in-oil-emulsions (W/O) (butter, margarines...). In all cases, two 43 immiscible fluids are involved, leading to metastable systems. Once fabricated, emulsions 44 evolve towards the total phase separation of the two fluids under the effect of coalescence 45 and Ostwald ripening, over time scales that may vary from hours to years (Bibette, Leal-46 47 Calderon, Schmitt, & Poulin, 2002). The kinetic stability and shelf life of emulsions is determined by the nature and concentration of the surface-active species (amphiphilic 48 49 polymers, proteins, low molecular weight surfactants) that are adsorbed at the oil-water interface. It is well known that the presence of crystals in the dispersed phase of O/W 50 emulsions may have profound consequences for their stability because of specific 51 52 instabilities caused by protruding crystals (Fredrick, Walstra, & Dewettinck, 2010). Also, 53 crystallization impacts the rheological properties. When fat droplets are attached to a gel 54 protein network formed in the continuous phase, they act as active fillers and droplets 55 crystallization may strongly enhance the gel firmness (Dickinson, 2012).

Milk fat is of special relevance in dairy products and this is why there is abundant 56 literature about its crystallization, both in bulk phase and in the emulsified state. 57 Crystallization of triacylglycerols (TAGs) is a two-stage process involving nucleation and 58 59 growth (Metin & Hartel, 2005). The presence of impurities like dust particles which are almost inevitable in natural fats, or the contact with solid substrates (scraped surface of 60 61 a heat exchanger, for example) during fat processing may favor a heterogeneous nucleation mechanism by catalyzing the formation of the first fat nuclei. Emulsification of 62 63 anhydrous milk fat (AMF) in an aqueous phase leads to a divided state of the fat, its confinement in drops, more or less pronounced depending on their average size, and the 64

65 existence of a water/oil interface, covered by amphiphilic species. All these parameters 66 can modify the fat crystallization scenario in terms of nucleation, growth, polymorphism and/or microstructure (Lopez, Lavigne, Lesieur, Keller, & Ollivon, 2001a; McClements, 67 68 2012). The impurities initially present in bulk fat become distributed within drops in the emulsified state. The smaller the average droplet size, the greater the number of drops 69 and the smaller their volume. As a result, the probability of finding impurities likely to 70 induce heterogeneous nucleation in a drop decreases and it is generally accepted that 71 72 nucleation tends towards the homogeneous mode (McClements, 2012). This is why, in order to induce crystallization, it is necessary to impose a higher degree of supercooling 73 74 as the average droplet size decreases (Walstra, Wouters, & Geurts, 2006; Lopez, Lesieur, Bourgaux, Keller, & Ollivon, 2001b). Lopez et al. (2002) combined optical 75 microscope observations between crossed analyzers and Differential Scanning 76 77 Calorimetry (DSC) measurements coupled with X-ray diffraction (XRD) to study AMF 78 crystallization in emulsified systems. They demonstrated that the average drop diameter 79 affects the crystal size and/or the structure of the crystal network because of defects in 80 the organization of TAG molecules in the crystals, either directly due to the curvature of the oil/water interface from which crystals are supposed to grow, or indirectly due to the 81 faster relaxation that can induce the formation of crystals of smaller size. Moreover, a 82 higher disorder and/or a smaller size of TAG crystals within the emulsion droplets was 83 84 observed, due to the faster supercooling relaxation induced by the decrease of droplet size. 85

Milk fat crystals can exist at the nanoscale in different crystalline forms, called polymorphs. The 3 main polymorphic forms encountered in TAGs are the α -hexagonal form, the β ' orthorhombic form, and the β triclinic form. They correspond to the lateral arrangement and degree of tilt of fatty acid (FA) chains in space (Sato, 1999). A fourth

90 unstable polymorph, called sub- α , can be observed in AMF at very low temperatures (<-8 °C) for rapid cooling rates (>|2.5 °C/min|) (ten Grotenhuis, van Aken, van Malssen, & 91 Schenk, 1999). In milk fat, different TAGs often associate and crystallize together to form 92 mixed crystals (Martini, Herrera, & Hartel, 2002). The α polymorph is likely to provide this 93 94 type of crystals since it is loose enough to incorporate TAGs of different FA compositions. The β' form also allows such compositional versatility (Walstra et al., 2006). TAGs also 95 self-organize themselves in a so-called longitudinal arrangement. They form stacks, 96 97 generally over two (2L), three (3L) or six chain lengths (6L), depending on the nature of FAs that compose them (Sato and Ueno, 2011). 98

99 Although the structure of AMF is largely dictated by its TAG composition and the thermal history, minor exogenous compounds or compounds formed during dairy 100 101 processes can also influence the crystallization scenario and the physical characteristics 102 of the fat (Smith, Bhaggan, Talbot, & Malssen, 2011; Talbot, Smith, & Bhaggan, 2012; 103 Rønholt, Mortensen, & Knudsen, 2013; Sato, Bayés-García, Calvet, Cuevas-Diarte, & 104 Ueno, 2013; Bayés-García et al., 2015; Patel & Dewettinck, 2015; Bayard, Leal-Calderon, 105 & Cansell, 2017). Minor compounds can accelerate or inhibit both nucleation and crystalline growth (Metin & Hartel, 2005; Smith et al., 2011; Rønholt et al., 2013; Sato et 106 al., 2013; Bayés-García et al, 2015; Bayard et al., 2017), and their influence can be 107 108 antagonistic, accelerating one while slowing the other (Talbot et al., 2012). In addition, 109 minor compounds can influence the polymorphic behavior, in particular by inhibiting some phase transitions. They could also block growth sites during recrystallization in 110 111 polymorphic phase transitions involving crystal melting (Smith et al., 2011; Talbot et al., 112 2012). Various factors determine the impact of modulators on crystallization and on the 113 resulting physical properties of AMF like their chemical nature (Wright & Marangoni, 114 2003; Smith et al., 2011), their concentration (Vanhoutte, Dewettinck, Foubert,

Vanlerberghe, & Huyghebaert, 2002; Foubert, Vanhoutte, & Dewettinck, 2004, Bayard et
al., 2017), and the processing conditions (Sato et al., 2013; Kaufmann, Kirkensgaard,
Andersen, & Wiking, 2013). It is worth noticing that most studies related to the impact of
minor compounds in AMF have been carried out in bulk systems.

In this paper, we examine AMF crystallization under confined conditions, *i.e.*, in 119 120 emulsion droplets, in the absence and in the presence of modulators at a concentration less than or equal to 5 wt%. At such low concentrations, the presence of minor 121 122 compounds is not expected to modify the nanostructure of AMF crystals (Bayard, Kaufmann, Leal-Calderon, & Cansell, 2021). However, depending on the nature of their 123 124 hydrocarbon chains, modulators can be water-soluble, can self-crystallize or may exhibit surface-active properties and their impact on AMF crystallization can combine volume 125 and/or interfacial effects that have not been addressed so far. To illustrate this diversity, 126 127 3 FAs, namely propionic, palmitic and oleic acids, and a TAG, tripalmitin, were selected. In a previous study devoted to bulk AMF crystallization, it was demonstrated that 128 129 propionic and oleic acids behaved as inhibitors, whilst palmitic acid and tripalmitin were 130 crystallization promotors (Bayard et al., 2017). The question arises as to whether these properties are retained in the emulsified state. The average droplet size of the emulsions 131 132 was tuned by varying the emulsification conditions so that 3 mean diameters were obtained. Fast cooling induced the formation of fat crystals whose kinetic evolution was 133 followed under isothermal and non-isothermal conditions. The crystallization state of AMF 134 was assessed by DSC during a fast-cooling step from 60 °C to 4 °C and after a storage 135 136 period of 90 min and 48 h under isothermal conditions, at 4 °C. Crystallization was also visualized *in situ* using the pending drop method. Overall, our approach aims at providing 137 138 insight into the effect of average droplet size of the emulsion and of the chemical nature

of minor compounds on supercooling phenomena and on the crystallization scenarioduring isothermal storage.

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142 **2. Materials and Methods**

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- 144 *2.1. Materials*

AMF was supplied by Corman SA (Goé, Belgium) and was used without further 145 purification. AMF was composed of approximately 6% short chain FAs (strictly less than 146 8 carbons), 20% midsize chains (with more than half of myristic acid), and 72% long 147 chains mainly represented by oleic acid (20 wt%), palmitic acid (32 wt%), and stearic acid 148 (9 wt%). Unsaturated chains represented 27% of the total FAs, oleic acid being the most 149 abundant one. The melting domain of AMF ranged from -40 °C to +37 °C. A low melting 150 151 AMF fraction (full melting for T> 10 °C; chain composition: C4-C6=7 wt%; C8 to C14= 19 152 wt%; C16= 23 wt%, C18:1= 32 wt%, C18= 7 wt%) of AMF provided by Corman (Goé, 153 Belgium) was used for surface tension measurements. Sodium caseinate (SC) with a 154 minimal protein content of 88% and a maximum moisture content of 6% was purchased from Armor Protéines (France). All chemicals adopted as crystallization modulators were 155 from Sigma-Aldrich (Saint-Quentin Fallavier, France): palmitic acid (16:0, purity> 98%, M 156 157 = 256 g/mol), propionic acid (3:0, purity > 99.5%, M = 74 g/mol), oleic acid (18:1, purity >95%, M = 282 g/mol) and tripalmitin (PPP, purity >85%, 100% triglyceride and 158 palmitic content >90%, M = 807 g/mol). Sodium Dodecyl Sulfate (SDS) was used to dilute 159 160 emulsions before droplet size measurements and sodium hydroxide was used to adjust 161 the pH.

The effect of modulators on AMF crystallization was studied on 3 emulsions of different diameters. The concentration of the modulators with respect to AMF mass varied between 0.5 and 5 wt%. Blends of AMF and crystallization modulators were first heated at 85 °C to obtain a homogeneous melt. This procedure that consists in heating the fat at least 10 °C above the temperature of the species with the higher melting point (16:0, 63 °C) is usually considered as sufficient to erase any crystal memory.

The emulsions formulated in this study were of the O/W type and were stabilized 170 171 by SC. The fat phase, either neat AMF or AMF+modulator, was the dispersed phase (40 wt%) and the continuous phase (60 wt%) was an aqueous solution at 3.33 wt% SC (2 172 173 wt% SC with respect to the whole emulsion mass). The aqueous and fat phases were mixed in an IKA LR 1000 laboratory reactor (Germany) equipped with an Ultra-Turrax® 174 turbulent mixer with a S25-KV-25F rotor-stator dispersing unit, thermostatically controlled 175 176 at 60 °C. This setup prevented crystallization during shearing and provided agitation to the system. The mixture to be emulsified (1.2 L) was subjected to an intense shear at 177 178 25,000 rpm for 10 min. The obtained emulsion was then stored at 60 °C for a period no 179 longer than 2h. From this primary emulsion, 2 daughter emulsions with reduced droplet size were obtained using a Niro Soavi Panda, GEA (Italy) high pressure homogenizer at 180 two homogenizing pressures, 50 and 500 bars. The obtained emulsions were stored at 181 182 60 °C, between 1 and 8h, until use. Emulsions resulting from the 3 homogenization levels 183 (0, 50 and 500 bars) will be referred to as LD ("Large Diameter"), MD ("Medium Diameter") and SD ("Small Diameter"). 184

A specific protocol was adopted for the emulsion containing 3:0. Due to their high polarity, short-chain FAs exhibit high water solubility (>3 mol/L) (Bell, 1973). Solubilization of 3:0 in water led to a decrease in pH that reached the isoelectric point of SC (close to 4.8) and caused protein precipitation, which precluded emulsification. Thus, the pH of the

189 aqueous phase was adjusted by adding sodium hydroxide, prior to the emulsification. The 190 concentration of sodium hydroxide in the aqueous phase was fixed at 0.042 mol/L. After emulsification, the final pH of the aqueous phase was equal to 6.3, close to that measured 191 for emulsions based on neat AMF (6.7 \pm 0.1), and on AMF mixed with modulators with 192 very low water solubility: AMF+16:0 at 2.5 wt% (6.6 ± 0.1), and AMF+18:1 at 2.5 wt% (6.5 193 \pm 0.1). Concentrations of 3:0 above 0.5 wt% were not probed as they required larger 194 195 amounts of sodium hydroxide likely to induce fast triglyceride hydrolysis because of the 196 initial basic pH conditions and to substantially increase the ionic strength in the aqueous 197 phase.

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199 2.3. Emulsion characterization

Emulsions were imaged with a phase contrast microscope (BX53F, Olympus, Japan). To facilitate the observation, they were diluted with a SDS aqueous solution at 8.10⁻³ mol/L. This surfactant is known to dissociate droplets potentially aggregated *via* protein bridging, which enables the observation of single droplets (Demetriades & McClements, 2000). Images were acquired using a 10.6-megapixel digital camera (SC100, Olympus, Japan).

The size distributions were measured using a Mastersizer 3000 apparatus (Malvern Instruments, UK). Static light-scattering data were transformed into size distribution using Mie Theory. The droplet refractive index was 1.46 and that of the aqueous phase was 1.33. The emulsions were submitted to a 100-fold dilution with an aqueous phase at 8.10⁻³ mol/L SDS in order to disaggregate droplets. A small volume of sample was then introduced under stirring at 1,200 rpm in the dispersion unit. Each measurement was performed in triplicate. The droplet distributions were described in

213 terms of their volume-averaged diameter, D_v , surface-averaged diameter, D_s , and 214 polydispersity, *P*, defined as:

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$$D_v = \frac{\sum N_i D_i^4}{\sum N_i D_i^3};$$
 $D_s = \frac{\sum N_i D_i^3}{\sum N_i D_i^2};$ $P = \frac{1}{\overline{D}} \frac{\sum N_i D_i^3 (\overline{D} - D_i)}{\sum N_i D_i^3}$ Equation (1)

where \overline{D} is the median diameter, *i.e.* the value for which the cumulative undersized volume fraction is equal to 50%, and N_i is the total number of particles with diameter D_i .

219 2.4. Differential scanning calorimetry

Thermal analyses were conducted on a differential scanning calorimeter (DSC 220 8500, Perkin Elmer, UK) in aluminum pans of 10 µL hermetically sealed. An empty, 221 hermetically sealed aluminum pan was used as reference. Calibration was made with 222 indium (melting point = 156.6 °C, melting enthalpy = ΔH_m = 28.45 J/g), and cyclohexane 223 224 (melting point = 6.47 °C, ΔH_m = 31.8 J/g) at scanning rates of ± 5 °C/min. All DSC 225 experiments were carried out in the temperature range from 4 °C to 60 °C. The heat flow was expressed in W/g of fat. For the sake of comparison, several plots were 226 superimposed in the same graph. To facilitate the readability, the graphs were shifted 227 vertically. The baseline corresponding to a zero-heat flow is given by the enthalpic signal 228 between 40 and 50 °C. 229

The melting or crystallization enthalpy variation, ΔH , was determined from the total area of the thermographs between 4 °C and T_{offset} , the temperature at which the DSC signal reached the baseline in melting experiments, or T_{onset} , the initial crystallization temperature in cooling experiments (non-isothermal crystallization). When thermograms presented several endothermic or exothermic events, the characteristic temperatures were taken at the maximum of the peaks. DSC thermograms were measured in triplicate for all crystallization experiments and for melting experiments performed with emulsions containing neat AMF. A single melting experiment was carried out for emulsions containing a modulator. In this case, for the statical analysis, we adopted the same relative standard deviation (SD/mean) as that of the emulsion based on neat AMF with comparable droplet size.

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242 *2.5.* Thermal programs

The following operating conditions allowed to assess the evolution of fat crystallization under dynamic (cooling) and static (isothermal) conditions (Figure SI1, supplementary information):

- Controlled cooling ramp from 60 °C to 4 °C at -5 °C/min followed by an isothermal stage
at 4 °C for 90 min and by a melting ramp at +5 °C/min with continuous recording of the
heat flow by DSC.

- Quench at 4 °C, followed by storage for 48 h at 4 °C. Samples introduced in sealed aluminum pans of 10 μ L were warmed at 60 °C and were then located in a thermostatically controlled chamber at 4 °C for 48 h. After that delay, the thermal properties of the crystallized state were characterized by DSC following a melting ramp at +5 °C/min.

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255 *2.6. Surface tension measurements*

Surface tension measurements at the oil/water interface were performed at 25 °C using the rising drop method (Tracker" apparatus from Teclis Instruments, France). Single fat droplets of 5 μ L were formed at the tip of a steel needle and images were recorded at regular time intervals with a digital camera. A low melting AMF fraction that was fully molten at the operating temperature was adopted for the measurements. The aqueous

261 phase contained SC at 0.33 wt%. The drop shape was fitted considering the Young-262 Laplace equation, the surface tension, γ being the unique free parameter. The same 263 setup was used to visually follow AMF crystallization. In this case, a larger AMF drop of 264 15 μ L was formed at the tip of the needle to facilitate the observation and the 265 concentration of the modulators 16:0 and PPP was 0.25 wt%.

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267 2.7. Statistical analysis

268 Droplet diameters, onset, offset and peak temperatures, melting and crystallization 269 enthalpies from DSC curves were expressed as mean values with standard deviation 270 (mean ± SD). Comparisons were analyzed using a one-way analysis of variance followed 271 by a pairwise t-test using R version 3.1.2. Differences with P-values <0.05 were 272 considered to be statistically significant.

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3. Results and discussion

We aim at assessing the influence on AMF crystallization of the droplet size and 275 of the nature of the modulators. Interestingly, the additives under study cover different 276 situations regarding their localization, either at the oil/water interface or in the droplets' 277 278 core, which will allow a modulation of AMF crystallization based on bulk or interfacial 279 effects. Both 18:1 and 16:0 FAs are surface-active and mainly oil-soluble (water solubility <10⁻⁶ mol/L (Bell, 1973)); 3:0 and PPP are not expected to have any interfacial activity. 280 As noted in Section 2.2, 3:0 is highly soluble in water and is distributed between the oil 281 282 and aqueous phase. Being nearly insoluble in water, PPP is primarily present in the fat globules. 283

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285 *3.1 Main characteristics of the emulsions*

286 Emulsions were characterized by droplet sizing based on light-scattering. The 287 main characteristics are provided in Table 1. The size distributions can be found in Figure SI2 of supplementary information. Irrespective of the formulation, the 3 levels of 288 homogenization applied (0 bar, 50 bars, 500 bars) allowed to obtain emulsions whose 289 290 droplets had significantly different diameters but all exhibiting a roughly monomodal 291 distribution. The non-homogenized LD emulsion based on neat AMF had a volume-292 averaged diameter, D_v of 6.0 μ m, close to the average diameter in raw milk (Michalski, Ollivon, Briard, Leconte, & Lopez, 2004). The homogenization at 50 bars (MD emulsion) 293 294 led to an average diameter of 2.2 µm. Homogenization at 500 bars (SD emulsion) resulted 295 in a submicron-sized emulsion of 0.8 µm (LD emulsion). Observations under the 296 microscope (Figure SI2-A in supplementary information) were consistent with the results 297 obtained via droplet sizing. The size distributions and average diameters obtained in the presence of the modulators are almost identical to those reported for neat AMF except 298 for the LD emulsions containing 18:1 and 16:0 that have slightly smaller diameters 299 300 compared to neat AMF (Table 1). This can be interpreted as being due to the surfaceactivity of both 18:1 and 16:0. Interfacial tension measurements were carried out using 301 302 the rising drop method. The concentration of SC in the aqueous phase was set at 0.33 wt% to avoid drop detachment due to excessively low interfacial tension (<4 mN/m). To 303 304 avoid the formation of fat crystals likely to perturb measurements, neat AMF was replaced by a low melting AMF fraction. Studies on competitive adsorption in model emulsion 305 306 systems stabilized by pure milk proteins have shown that the presence of oil-soluble or 307 water-soluble surfactants during emulsification leads to a reduction in surface tension, in 308 the protein surface coverage and in the average droplet size of the resulting emulsions, 309 irrespective of the oil and of the protein nature (De Feijter, Benjamins, & Tamboer, 1987). The values obtained 1 h after forming the drop were 10.8 ± 0.5 mN/m, 6.2 ± 0.5 310

mN/m and $8.2 \pm 0.5 mN/m$ for neat fat, and fat containing 18:1 and 16:0, respectively. It can be argued that SC which stabilizes emulsions is at least partially displaced by these two FAs, leading to lower surface tensions and thus smaller average diameters.

314 For any surface-active species, the mass, *m*, required to fully cover the fat/water 315 interface is given by the following relationship:

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$$m = \Gamma \frac{6m_{AMF}}{D_S \rho_{AMF}}$$
 Equation (2)

where Γ is the surface coverage, m_{AMF} is the AMF mass and ρ_{AMF} is the AMF density (0.92 g/cm³). For SC, Γ is close to 1.2 mg/m² (Robson and Dalgleish, 1987). Thus, for the emulsion with smallest average diameter homogenized at 500 bars, the amount of SC adsorbed at the interface represents about 34% of the mass initially introduced. This indicates that the protein concentration is, in any case, sufficient to saturate the interface generated by the emulsification process.

Assuming that the surface coverage of 16:0 is Γ = 1.70 mg/m² (equivalent to 25 323 324 $Å^2$ /molecule) (Karleskind, 1992) and considering the average diameters D_s reported on Table 1, from Eq. 2, we can estimate that the interface mobilizes at the most 15%, 34% 325 326 and 99% of the initial amount of this surfactant in LD, MD and SD emulsions, respectively. 327 16:0 is thus expected to localize at both the water/oil interface and in the volume of the 328 fat drops. The same statement holds for 18:1. Assuming Γ = 1.65 mg/m² (28.6 329 Å²/molecule) (Karleskind, 1992), the interface mobilizes at the most 14%, 34% and 97% of this surfactant in LD, MD and SD emulsions, respectively. 330

Coalescence of the drops, if any, is supposed to increase the average diameter of the emulsions and an evolution of the thermograms would then be expected. The size distributions of emulsions stored at 4 °C for 48 h and rewarmed at room temperature were measured and were strictly identical to those obtained right after emulsification (Results

not shown). Thus, for any emulsion, it can be stated that variations in thermal profiles
over time are not due to a change in droplet size distribution.

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338 *3.2* Non-isothermal crystallization following a controlled cooling ramp

339 *3.2.1 Neat AMF*

340 Crystallization of emulsions following a controlled ramp at -5 °C/min between 60 341 and 4 °C, was studied by DSC and compared to bulk AMF crystallized at the same rate 342 (Figure 1). Such approach allows to reveal supercooling effects induced by the dispersion 343 of matter, with a shift of the exothermic signal to lower temperatures as the size of the 344 drops decreases.

The onset crystallization temperature, T_{onset} , of bulk AMF is equal to 18.4 ± 0.9 °C. 345 As the average droplet size of the emulsions decreases from 6.0 to 0.8 μ m, T_{onset} 346 significantly shifts from 15.7 ± 0.2 °C to 12.4 ± 0.4 °C (Figure 1, Table 2). This observation 347 348 about the impact of emulsification and of the droplet size is consistent with previous studies (Michalski et al., 2004; Lopez et al., 2002; Truong, Bansal, Sharma, Palmer, & 349 Bhandari, 2014). The dispersion of fat in drops leads to its confinement. The substrates 350 of heterogeneous nucleation initially present in bulk AMF are therefore also confined in 351 the drops. Once the nucleation stage is overcome, crystal growth is limited by the finite 352 extension of the drops. This explains the higher degree of supercooling in emulsions 353 354 compared to bulk AMF. In addition, for the same fat fraction, when the average diameter of the drops decreases, substrates of heterogeneous nucleation are increasingly 355 confined, *i.e.* the volume fraction of the drops that contain them decreases, which 356 357 necessarily slows down crystallization.

The number of DSC peaks and their position are also influenced by the emulsification and by the size of the drops (Figure 1). In Table 2, we report the position

360 of the most intense ones. At a cooling rate of -5 °C/min, bulk AMF crystallizes following a two-stage process, and the corresponding peaks are centered at 17.2 °C and 11.4 °C. 361 The thermogram obtained for LD emulsion exhibits 3 peaks: a low-amplitude event 362 around 14 °C and two peaks centered at 12.5 °C and 8.6 °C. For MD emulsion, the 363 thermal profile comprises a main peak centered at 8.6 °C, with a low-intense shoulder at 364 higher temperatures. Finally, the thermogram of SD emulsion contains a single peak 365 centered at 7.3 °C. To sum up, reducing the size of the drops in emulsions has the effect 366 367 of directing crystallization towards a single thermal event. In submicron SD emulsion, the different AMF molecular fractions no longer crystallize separately as in bulk phase or in 368 369 large droplets, but jointly.

In bulk AMF, crystalline growth preferentially results from the accretion of 370 triglycerides with similar molecular size and shape. The crystallization process 371 372 segregates triglycerides according to the chain length of their esterified FAs and their saturation level (Lavigne and Ollivon, 1997; Lopez, 2020). Therefore, several exothermic 373 374 peaks are observed, each corresponding to a different molecular fraction, generally 375 referred to as low-, middle- and high-melting TAGs. Conversely, in MD and SD emulsions, crystal growth generates mixed crystals involving different molecules, formed during a 376 single and more intense exothermic event, reflecting a loss in molecular selectivity. Within 377 378 a single droplet, the diffusion-limited process ensuring crystal growth is limited by the 379 finite extension of the drops and molecules with similar molecular structure are quickly depleted. In the meanwhile, the expansion of the supercooling domain (lower T_{onset}) tends 380 381 to diversify triglycerides available for adsorption on the growing crystalline faces. It can 382 be assumed that fast crystallization in emulsions promotes growth in the α polymorphic 383 form which is loose enough to allow the incorporation of triglycerides of different chain 384 lengths.

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386 *3.2.2. Impact of the modulators*

387 All systems containing modulators are compared to control emulsions containing388 neat emulsified AMF.

The thermal profiles obtained for the emulsions with and without 3:0 at 0.5 wt% in 389 390 AMF are similar (Figure 2-A). For a given droplet size, comparable exothermic enthalpies, 391 centered on the same temperatures are obtained (Table 2). Bayard et al. (2017) showed 392 that 3:0 has an inhibitory effect on bulk AMF crystallization. They used pulsed lowresolution nuclear magnetic resonance (p-NMR) to measure the solid fat content (SFC) 393 394 following a fast quench from 80 °C to 25 °C. AMF crystallization exhibited a sigmoidal curve with an induction time followed by a sudden SFC increase whose average slope 395 was indicative of a steady crystallization process. The induction time was defined as the 396 397 delay between the start of the quench and the onset of the SFC signal. The addition of 398 3:0 at 1 wt% in AMF had the effect, among others, to considerably prolong the induction 399 time for crystallization and to slightly decrease the final SFC value. The inhibiting effect 400 was explained as being mainly due to the entropy of mixing. The presence of a low molecular weight solute tends to lower the melting temperature following Raoult's law 401 402 (Atkins & Paula, 2014). In the present study, the partition of 3:0 between the oil droplets 403 and the aqueous phase makes the impact on AMF crystallization negligible.

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Figure 2-B shows the evolution of the heat flow for emulsions based on 18:1 at 2.5 wt% in AMF. This modulator was shown to exhibit an inhibitory effect in bulk AMF by p-NMR, revealed by a higher induction time and a lower maximal crystallization rate compared to plain AMF (Bayard et al., 2017). The second crystallization peak, centered around 12.5 °C for AMF emulsion, disappears in the presence of 18:1. Decreasing the droplet size shifts the single crystallization peak to lower temperatures compared with neat AMF (Table 2), revealing the supercooling effect of 18:1.The TAG fraction involved in this large exothermic peak crystallizes at lower temperatures, together with a lower melting fraction, probably forming mixed crystals. Two hypotheses can be drawn to explain the origin of these observations:

Interfacial effect. A fraction of 18:1 is adsorbed at the oil/water interface where it
forms a brush of chains oriented towards the oil phase. The disorder induced by the
double bond in *cis* configuration makes the brush structure incompatible with the ordered
packing of saturated AMF chains. As the average diameter decreases, the interfacial
surface area of the drops is augmented and the inhibitory effect of 18:1 becomes
increasingly pronounced.

421 - Volume effect. As in the continuous phase, 18:1 hinders the formation of crystal
422 nuclei (Bayard et al., 2017). Nucleation being the kinetically determining step, the
423 inhibitory effect of 18:1 is amplified when the volume of the drops decreases.

424

425 Figure 2-C corresponds to emulsified systems containing 16:0 at 2.5 wt% in AMF. For the 3 sizes, the presence of 16:0 significantly increases *T*_{onset} compared to the control 426 emulsion (Table 2). The promoting effect of 16:0 in emulsified systems is consistent with 427 428 what was observed in bulk AMF by p-NMR, *i.e.* a lower induction time and a higher maximal crystallization rate (Bayard et al., 2017). Moreover, the final SFC was not 429 affected by the addition of 16:0 (Bayard et al., 2017). The addition of 16:0 leads to new 430 431 exothermic events. In MD emulsion, a peak centered at 8.8 °C and a weak exothermic signal between 18 °C and 14 °C appear, while the control emulsion shows a single 432 433 crystallization peak. For SD emulsion, a thermal event, not very intense and quite broad, 434 is distinguishable around 13 °C. Those new events suggest that 16:0 favors a segregation

of TAGs, with different fractions crystallizing separately. The value of *T*onset obtained with 435 436 16:0 for the 3 emulsions slightly varies from 17 to 18 °C depending on the average droplet size (Table 2). This result suggests that nucleation is no longer hindered by the decrease 437 438 in drop size. It can be assumed that 16:0 accelerates nucleation by acting through the oil/water interfaces, where it concentrates. Saturated chains of 16:0 would crystallize at 439 440 the interface and would promote crystallization by acting as templates. It is well known 441 that crystal growth is favored if the alkyl chains within the nuclei are of similar length to 442 those of the main fraction of the fat (Basso et al., 2010). The templating effect can be due to the fact that 16:0 chains are abundant in AMF. Since the amount of interface becomes 443 444 larger in finer emulsions, nucleation does not require deeper supercooling anymore. Nevertheless, a mode of action via the core of the drops cannot be discarded. Indeed, 445 the amphiphilic nature of 16:0 may allow this FA to self-assemble in the melted fat. Due 446 447 to the stacking of the aliphatic chains, the structures resulting from the self-assembly would constitute numerous nucleation sites. Reverse micelles with the hydrophobic 448 449 "brush" facing the oil phase can potentially be formed and may accelerate nucleation 450 (McClements, 2004).

451

To better understand the extent to which the interfacial and bulk properties of 16:0 are involved in accelerating AMF crystallization, the impact of 16:0 was compared to that of PPP which is not surface-active, while comprising the same FA chain. Emulsions containing 16:0 or PPP were prepared at 5 wt% in AMF. Their thermal profiles are presented in Figure 2-D. The modulator content is higher than previously, which will allow in the meanwhile to assess the influence of the modulator concentration.

458 Doubling 16:0 concentration greatly modifies the AMF crystallization profile 459 (Figures 2-C and 2D). Regardless of the droplet size, an increase in *T_{onset}* is observed

(Table 2). In addition, crystallization involves only one mean intense peak, instead of the
two peaks observed at lower 16:0 concentration. Since the size distributions are
comparable for both concentrations, it may be concluded that a high 16:0 concentration
promotes AMF crystallization in the droplets core.

For LD emulsion, substitution of 16:0 by PPP induces a significant change in the 464 thermal pattern with the appearance of a new peak centered at 14 °C, nearly 3 °C above 465 the main peak of AMF+16:0 and a shoulder around 17 °C (Figure 2-D). A second peak, 466 467 less intense, is centered around 8 °C (this peak was already present in the control emulsion). For SD emulsion, PPP shifts the main crystallization peak by + 2.4 °C 468 469 compared to the emulsion based on neat AMF. An intermediate pattern is observed for MD emulsion. Irrespectively of the droplet size, PPP modulates the crystallization by 470 471 increasing *T*_{onset}.

472 To further assess the origin of the promoting effect of 16:0 and PPP, a 15 µL drop of AMF, melted at 80 °C, was formed in a 0.33% SC solution, thermostatically controlled 473 474 at 25 °C in a rising drop tensiometer. Figure 3-A shows a sampling of the acquired images. 475 As the crystals form, they prevent light from passing through the fat phase, causing dark spots to appear in the drop. This experiment confirms that 16:0 and PPP are 476 477 crystallization promoters, as drops containing them darken faster than the control system 478 based on neat AMF. It also allows to appreciate the shape, size and number of crystals 479 and to see where they preferentially form, in the drop core or at the interface. Figure 3-B shows a magnification of the images obtained after 12 minutes of crystallization. The 480 481 control drop does not yet show any crystal, while the first crystals are visible in the 482 presence of 16:0 and PPP. The crystals in the drop containing PPP tend to form a 483 sediment (indicated by the arrow). In Figure 3-A, at times between 24 and 36 min, the 484 upper part of the drop appears slightly less turbid than the lower part, again reflecting

crystals sedimentation. On the other hand, in the presence of 16:0 (Figure 3-B), the crystals formed remain attached to the water/oil interface and the blackening of the drop is spatially homogeneous (Figure 3-A). This blackening is also faster than in the drop containing PPP. The crystals form a thick layer at the interfaces and thus opacify the drop rapidly. This experiment supports the conclusion that 16:0 accelerates crystallization *via* the interface, while PPP acts *via* the core of the drop.

491 From the whole set of observations, it can be concluded that different promoting 492 mechanisms are at play for these two modulators:

- Crystallization of 16:0 occurs first at the interfaces, where it tends to adsorb and 493 494 pack due to its amphiphilic nature. The finer the drops, the greater the amount of interface and thus the fraction of adsorbed 16:0, which could justify the increase in intensity of the 495 early exothermic peak (Figure 2-D). However, the increasing curvature may hinder 496 497 crystallization since the peak maximum shifts towards lower temperatures as the drop 498 size decreases. Tonset tends to increase with the concentration of 16:0 in AMF, presumably 499 due to an increase of the interfacial concentration of this modulator. Nevertheless, a 500 volume effect cannot be excluded since the concentration of free 16:0 in the core of the drops is also likely to increase, especially in emulsions with large average droplet 501 diameter. 502

503 - PPP crystallizes concomitantly with a fraction of high-melting AMF TAGs. It has 504 been shown that adding PPP at low concentration does not modify the nanostructure of 505 AMF crystals (Bayard et al., 2021). This modulator would mobilize saturated AMF TAGs 506 comprising comparable chain lengths to form mixed crystals. The expansion of the 507 supercooling domain (decrease of T_{onset}) as the average droplet size decreases gives a 508 hint that that its effect has a bulk (volume) origin.

509

510 3.3 Melting behavior of emulsions after isothermal crystallization at 4 °C for 90 min and

511 **48** h

512 3.3.1 Neat AMF

513 Following the crystallization ramp at -5 °C/min, the samples reached the target temperature, 4 °C, after 11 min and were subjected to a short (90 min) isothermal period 514 at this temperature. Alternatively, samples were guenched from 60 °C to 4 °C in 515 approximately 2-3 min and were stored for 48 h at 4 °C in a thermostatically controlled 516 517 chamber. The crystallization state was then characterized by DSC following a melting ramp at +5 °C/min. In Figure SI3 (supplementary information), the thermal melting profiles 518 519 for the 3 emulsions are superimposed for the 2 cold storage times: 90 min (A) and 48 h (B). Crystallization keeps on between these 2 storage periods, as evidenced by the 520 overall heat flow observed in the thermograms between 4 °C and 15 °C which is lower 521 522 after 48 h of crystallization than after 90 min. For instance, an intense endothermic peak around 21 °C, not observed after 90 min, is present after 48 h. This reflects an evolution 523 524 of the crystals from a less stable form (melting between 4 and 15 °C) at short times, to 525 more stable forms (melting around 21 °C) when the isothermal period becomes longer. It is likely that the medium melting fraction of AMF undergoes a polymorphic transition 526 between 90 min and 48 h of crystallization. In the remainder, we will mainly comment the 527 results obtained after 48 h of storage for both AMF and AMF+modulator systems. 528

529 Whatever the average droplet size, the melting profiles of AMF emulsions stored 530 for 48 h at 4 °C exhibit 3 endothermic events (Figure 4 and Figure SI4 in supplementary 531 information):

- A peak at temperatures below 15 °C, reflecting the fusion of the most unstable
 forms, probably 3L-α forms (Michalski et al., 2004);

- An intense peak between 15 °C and 25 °C, attributable to the melting of 3L and 535 2L α and β' forms (Lopez et al., 2001b);

536 - A thermal event around 26 °C that would correspond to the melting of a $2L-\beta'$ 537 form (Michalski et al., 2004). This event is more apparent on the thermograms of LD and 538 MD emulsions. The curvature and confinement imposed by the submicron-sized drops in 539 SD emulsion inhibit the polymorphic transition leading to the formation of this crystals 540 fraction.

541

542

3.3.2 Impact of the modulators

543 The melting profiles obtained after 48 h at 4 °C, for the 3 droplet sizes (Figure 4-A) show that the emulsions with and without 3:0 exhibit similar enthalpic behaviors, again 544 suggesting that this modulator initially dissolved at 0.5 wt% has no impact on the 545 546 formation of the polymorphs, mainly because of its high-water solubility. Likewise, the 547 endothermic peaks with and without 18:1 are guite close (Figure 4-B, Table 2). The 548 melting profiles are weakly modified by the presence of 16:0 at 2.5 wt% in AMF (Figure 549 4-C, Table 2). Thus, after a sufficiently long crystallization time, the unstable polymorphs formed at short times evolve towards more stable ones, regardless of the drop size. 550

551

Both 16:0 and PPP at 5 wt% in AMF lead to the appearance of endothermic peaks at temperatures higher than those of the control (Figure 4-D, Table 2). For 16:0, a weak endothermic event is observed between 35 °C and 40 °C in LD emulsion and between 35 °C and 47 °C in MD emulsion. For PPP, the melting range of AMF extends to about 45 °C, a variation of +10 °C in T_{offset} compared to neat AMF. The heat flow is significant between 35 °C and 45 °C. The enthalpy of fusion in this range cannot be due to PPP alone. Indeed, in Figure SI5 (supplementary information), we show the signal obtained

559 for LD emulsion as well as its integral from 4 to 55 °C. The SFC of bulk AMF is 52% at 4 °C (Bayard et al., 2017). Assuming that PPP is fully crystallized, the expected fraction 560 of the enthalpic signal between 35 °C and 45 °C should represent about 10% of the 561 562 cumulative signal, which is half of the actual value obtained in Figure SI5. In addition, this signal appears in the continuity of thermal events occurring at lower temperatures and 563 not as an individual peak. It can be concluded that this endothermic heat flow corresponds 564 to the melting of mixed crystals comprising PPP and AMF TAGs. Conversely, in the 565 system containing 16:0, the isolated peak at high temperature, around 40 °C, and its low 566 intensity suggest that the signal is due to the melting of 16:0 alone, which would be 567 568 segregated from the rest of the fat. The melting temperature of pure 16:0 is 63 °C, but in the presence of liquid oil, the entropy of mixing lowers its melting temperature. 569

570

571 *3.4 Analysis of the phase change enthalpies*

A detailed analysis of the total phase change enthalpy ΔH between 4 and 45 °C 572 573 (Table 2) was carried out with the aim of assessing the crystallization kinetics. Figure SI6-A and B (supplementary information) display the average values obtained with neat AMF 574 575 and with all samples including neat AMF and AMF+modulators, respectively. We consider the absolute value of the crystallization enthalpy, ΔH_{cryst} , measured following a cooling 576 577 ramp in order to facilitate its comparison with the melting enthalpies measured after a 578 storage of 90 min, ΔH_{90min} , and 48 h, ΔH_{48h} . The crystallization enthalpy is lower than the melting enthalpies. This can be explained by the fast cooling that occurred in 11 minutes, 579 resulting in partial crystallization, and by the formation of metastable polymorphs, whose 580 enthalpies are known to be lower than those of stable ones (Ravotti, Worlitschek, Pulham, 581 582 & Stamatiou, 2020). After a given storage time, with only a few exceptions, the total melting enthalpies follow the same hierarchy: $\Delta H_{LD} > \Delta H_{MD} > \Delta H_{SD}$. 583

All other factors being equal, the average values obtained with AMF alone and with all samples are almost identical (the differences between figures SI6-A and B are not statistically significant). The modulators modify the crystallization scenario at short times as revealed by the thermograms reported in Figures 2, 4 and SI4 but the global enthalpy is a "mean field" parameter that does not allow to differentiate the impact of the modulators.

590 Crystallization still evolves after 90 minutes. This can be concluded after noticing 591 that ΔH_{90min} is lower than ΔH_{48h} . Several factors may explain this lower enthalpy: (i) a 592 lower solid content; (ii) the presence of metastable forms with lower melting enthalpy than 593 more stable polymorphs formed during storage; (iii) transitions towards more stable forms 594 during fusion trigger an exothermic signal that lowers the total enthalpy (Figure SI4).

595 After 48 h, it is not clear whether a steady state has been reached. The enthalpy of SD emulsions remains significantly lower than in the other 2 emulsions. It can be 596 597 argued that the crystallization kinetics is slower in SD emulsion. However, an alternative 598 interpretation can be put forward. According to the classical theory of homogeneous 599 nucleation, the Gibbs energy difference associated to the formation of a solid germ from 600 liquid molecules comprises a volume (bulk) term linked to the difference in the standard 601 chemical potential of liquid and solid phases and an interfacial contribution accounting for solid-liquid interfacial tension (Abraham, 1974). Within this model, the growth of a germ 602 603 leading to crystal formation is only possible if it forms spontaneously with a size larger than a critical threshold value, r*. It can be hypothesized that the confinement in very tiny 604 droplets modifies the thermodynamic conditions. Grossier and Veesler (2008) have 605 606 calculated the change in free energy associated to the formation of a nucleus containing 607 solute molecules in a supersaturated finite system (droplet). As the nucleus grows, solute 608 molecules in the liquid phase are depleted, which tends to reduce supersaturation.

609 Because of this, at low initial supersaturations, it is impossible for a critical nucleus to 610 form. A critical nucleus can form only if supersaturation exceeds a threshold value. In this case, once the critical size has been reached, the nucleus continues to grow until it 611 612 reaches a second critical size associated with a potential well in the energy curve. Growth beyond this "equilibrium" size is not favored. These thermodynamic effects can therefore 613 limit the size that a nucleus can attain within a finite reservoir like tiny droplets, or can 614 615 even prevent nucleation from supersaturated or undercooled solutions that would yield crystals in bulk conditions. 616

617

618 **4. Conclusion**

In this paper, we studied the impact of confinement and of the presence of 619 additives on the crystallization and melting of milk fat. Differential scanning calorimetry 620 621 was used to assess the kinetics and the nature of thermal events occurring during crystallization under non-isothermal and isothermal conditions. The impact of 622 623 confinement was addressed by fabricating oil-in-water emulsions and by tuning their 624 average drop size. Upon cooling, in the absence of additives, the crystallization onset temperature tended to decrease as the average drop size was reduced. In addition to this 625 effect usually observed in emulsions based on crystallizable oils, we showed that unstable 626 crystals formed in the early stages and that crystallization was uncomplete even after 48 627 628 h of low temperature storage in submicron emulsions.

Additives likely to be formed in dairy processes, propionic, palmitic and oleic acids or added like tripalmitin were dissolved in milk fat at a concentration less than or equal to 5%. In emulsions, propionic acid showed no effect. Due to its high polarity, this additive was transferred to a large extent in the aqueous phase. Oleic acid favored supercooling and promoted unstable forms in the early stages of crystallization but its impact became

insignificant after 48 h of cold isothermal storage. Two crystallization promoters, palmitic
acid and tripalmitin, were also studied. The impact of palmitic acid was related to its
interfacial (amphiphilic) properties and vanished considerably after 48 h. Conversely,
tripalmitin influenced crystallization *via* volume effects that were persistent. It formed
mixed crystals which considerably extended the melting range of the milk fat.

639 We hope the results provided in this article will provide a useful guidance for 640 mastering crystallization in dairy products.

641

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645	Table Captions
646	Table 1: Average diameter and polydispersity of AMF-in-water emulsions with and without
647	modulators obtained at various homogenization pressures.
648	
649	Table 2: Characteristic temperatures and enthalpies of AMF-in-water emulsions with and
650	without modulators obtained at various homogenization pressures. All data derive from
651	DSC experiments.
652	
653	Figure captions
654	
655	Figure 1: DSC crystallization thermograms of bulk AMF and of AMF-in-water emulsions
656	following a controlled cooling ramp from 60 °C to 4 °C at -5 °C/min. Heat flows are related
657	to the mass of AMF contained in the sample. Thermograms are off set vertically for clarity.
658	
659	Figure 2: DSC crystallization thermograms of emulsions containing various modulators
660	following a controlled cooling from 60 °C to 4 °C at -5 °C/min. The thermogram of the
661	emulsion comprising neat AMF is provided for comparison.
662	
663	Figure 3: A: Images of AMF drops (15 $\mu L)$ during crystallization at 25 °C. From left to right:
664	neat AMF (control), AMF+0.25 wt% PPP; AMF+0.25 wt% 16:0. B: Focus on time t=12
665	min. Each of the modulators was added in molten AMF.
666	
667	Figure 4: DSC melting thermograms of emulsions containing various modulators
668	measured after 48 h of isothermal storage at 4 °C. The thermogram of the emulsion
669	comprising neat AMF is provided for comparison.
	28

670 Supplementary data captions

Figure SI1: Thermal programs implemented to assess the evolution of fat crystallization
under dynamic (cooling, solid line) and static (isothermal, dotted line) conditions.

673

Figure SI2: (A) Micrographs of neat AMF-in-water emulsions obtained at various homogenization pressures. The emulsions were diluted prior to the observation. The scale bar represents 50 μ m. (B) Size distribution of AMF-in-water emulsions with and without modulators obtained at various homogenization pressures; the darkest lines correspond to neat AMF; in the bottom graph, the lightest lines correspond to PPP and those with intermediate intensity correspond to 16:0.

680

Figure SI3: Melting thermograms after 90 min (A) and 48 h (B) of isothermal storage at
4 °C of neat AMF-in-water emulsions of various average diameters.

683

Figure SI4: DSC melting thermograms of emulsions containing various modulators measured after 90 min of isothermal storage at 4 °C. The thermogram of the emulsion comprising neat AMF is provided for comparison.

687

Figure SI5: DSC thermogram of LD emulsion following a ramp at +5 °C/min after
isothermal storage at 4 °C for 48h. The fat phase contains 5 wt% PPP in AMF. Left axis:
heat flow; right axis: cumulative curve (enthalpy).

691

Figure SI6: Crystallization and melting enthalpies of (A) emulsions based on neat AMF
and (B) all emulsions comprising neat AMF and AMF+modulators. The values reported
here are averaged over all data provided in Table 2.

Crystallization enthalpy (absolute value)

Melting enthalpy after 90 min

Melting enthalpy after 48h

695

- 696 Mean values under the vertical bars are significantly different (p < 0.05) using a one-way
- 697 analysis of variance followed by a pairwise t-test.

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Tables

Table 1

	Homogenization	Emulsion	D v ^{1,4}	D s ^{2,4}	Polydispersity ^{3,4}	
	pressure (bars)		(μm)	(μm)	(%)	
	0	Large	6.0 ± 0.2^{a}	3.4 ± 0.1 ^a	48 ± 1 ^a	
Neat AMF	50	Medium	2.2 ± 0.3^{b}	1.3 ± 0.1 ^b	63 ± 1 ^b	
	500	Small	$0.8 \pm 0.0^{\circ}$	$0.5 \pm 0.0^{\circ}$	80 ± 3°	
	0	Large	6.2 ± 0.1ª	3.6 ± 0.3^{a}	48 ± 3ª	
	50	Medium	2.2 ± 0.0^{b}	1.4 ± 0.0^{b}	57 ± 2 ^b	
0.5 WI /8 3.0	500	Small	$0.7 \pm 0.0^{\circ}$	$0.5 \pm 0.0^{\circ}$	73 ± 2 ^c	
	0	Large	4.8 ± 0.1ª	3.0 ± 0.5^{a}	47 ± 4ª	
2 5 wt% 18-1	50	Medium	2.2 ± 0.0^{b}	1.3 ± 0.0^{b}	61 ± 3 ^b	
2.3 WI /0 10.1	500	Small	$0.7 \pm 0.0^{\circ}$	$0.4 \pm 0.0^{\circ}$	75 ± 1°	
	0	Large	4.4 ± 0.1ª	2.9 ±0.1ª	45 ± 3 ^a	
2 5 wt% 16.0	50	Medium	2.2 ± 0.0^{b}	1.3 ± 0.1 ^b	61 ± 4 ^b	
2.5 WI /8 10.0	500	Small	$0.7 \pm 0.0^{\circ}$	$0.5 \pm 0.0^{\circ}$	71 ± 2°	
	0	Large	4.3 ± 0.1ª	2.7 ± 0.1 ^a	48 ± 1 ^a	
5 wt9/ 16.0	50	Medium	1.8 ± 0.2 ^b	1.1 ± 0.1 ^b	61 ± 2 ^b	
5 WI /8 TO.U	500	Small	$0.6 \pm 0.0^{\circ}$	$0.4 \pm 0.0^{\circ}$	71 ± 6°	
	0	Large	6.2 ± 0.1ª	3.7 ± 0.2^{a}	48 ± 2 ^a	
	50	Medium	2.7 ± 0.1 ^b	1.6 ± 0.0^{b}	64 ± 2 ^b	
	500	Small	$0.8 \pm 0.0^{\circ}$	$0.6 \pm 0.0^{\circ}$	78 ± 0°	

¹ D_v: volume-averaged diameter $\frac{\sum N_i D_i^4}{\sum N_i D_i^3}$ where N_i is the total number of particles with diameter D_i

² D_s: surface-averaged diameter, $\frac{\sum N_i D_i^3}{\sum N_i D_i^2}$ where N_i is the total number of particles with diameter D

diameter D_i

³ *P*: polydispersity, $\frac{1}{\overline{D}} \frac{\sum N_i D_i^3 (\overline{D} - D_i)}{\sum N_i D_i^3}$ where \overline{D} is the median diameter, *i.e.* the value for which the cumulative undersized volume fraction is equal to 50%, and N_i is the total number of particles with diameter D_i

⁴ Values are expressed as mean \pm standard deviations, n = 5 independent preparations. Within a column, mean values with unlike superscript letters are significantly different (p < 0.05) using a one-way analysis of variance followed by a pairwise t-test.

		Crystallization				Melting after 90 min				Melting after 48h		
Emulsion		T _{onset} ^{1,4}	7	r _{peak} ²	∆H ^{3,4}	7	- 2 peak	T _{offset} ¹	∆H ^{3,4}	T _{peak} ²	T _{offset} ¹	∆H ^{3,4}
		°C	°C	°C	J/g of fat	°C	°C	°C	J/g of fat	°C	°C	J/g of fat
	bulk	18.4 ± 0.9	16.8 ± 0.3	10.2 ± 0.3								
Noot AME	LD	15.7 ± 0.2ª°	12.5 ± 0.1ª°	8.6 ± 0.6	-37.1 ± 3.9 ^{ab} °	19.6 ± 0.5ª°	32.6 ± 0.4 ^a °	36.4 ± 0.3 ^a °	59.2 ± 9.6 ^a °	21.5 ± 0.5ª°	36.8 ± 0.3 ^{a°}	66.2 ± 12.8°°
	MD	13.7 ± 0.9ª*	8.6 ± 0.7 ^{ab} *		-33.5 ± 3.1 ^{ab} °	14.9 ± 0.2ª*	31.6 ± 1.8 ^{a°*}	36.3 ± 0.5 ^a °	44.5 ± 7.5 ^{a°}	21.6 ± 0.2ª°	36.5 ± 0.1ª°	61.2 ± 3.3 ^a °
	SD	$12.4 \pm 0.4^{a#}$	$7.3 \pm 0.3^{a#}$		$-23.8 \pm 4.6^{ab*}$	$13.3 \pm 0.3^{a#}$	$30.5 \pm 0.1^{a*}$	36.1 ± 0.5 ^{a°}	41.4 ± 8.4 ^a °	21.7 ± 0.2 ^{a°}	36.5 ± 0.5 ^{a°}	53.3 ± 9.0 ^a °
	LD	15.1 ± 0.1ª	12.3 ± 0.1ª	8.6 ± 0.1^{a}	-32.8 ± 1.0ª	19.2 ± 0.5ª	33.4 ± 0.4^{a}	36.4 ± 0.3 ^a	52.6 ± 8.5ª	21.6 ± 0.5ª	36.0 ± 0.3^{a}	57.5 ± 11.1ª
AMF+	MD	13.4 ± 0.3ª	9.1 ± 0.7^{a}		-35.0 ± 1.4^{abc}	14.8 ± 0.2ª	31.0 ± 1.8 ^a	36.2 ± 0.5ª	46.2 ± 7.8 ^a	20.8 ± 0.2 ^b	36.2 ± 0.1^{a}	57.4 ± 3.1ª
0.5 wt% 3:0	SD	11.5 ± 0.1ª	6.7 ± 0.4^{a}		-28.4 ± 0.5^{b}	13.4 ± 0.3^{a}	30.6 ± 0.1^{a}	36.2 ± 0.5 ^a	31.8 ± 6.5^{a}	21.6 ± 0.2 ^a	36.0 ± 0.5^{a}	44.1 ± 7.4^{a}
	LD	15.8 ± 0.1ª	9.0 ± 0.3^{d}		-36.5 ± 1.1 ^{ab}	15.2 ± 0.4 ^b	33.0 ± 0.4^{a}	36.0 ± 0.2 ^a	62.1 ± 10.1ª	21.0 ± 0.5ª	36.2 ± 0.3^{a}	64.1 ± 12.4ª
AMF+	MD	13.3 ± 0.4ª	7.6 ± 0.3 ^b		-30.6 ± 4.8^{b}	13.2 ± 0.2 ^b	33.0 ± 1.9ª	35.6 ± 0.5ª	41.2 ± 6.9 ^a	21.2 ± 0.2 ^{ab}	36.2 ± 0.1^{a}	51.8 ± 2.8^{b}
2.5 wt% 18:1	SD	10.8 ± 2.5ª	4.7 ± 0.1^{b}		-21.8 ± 3.6 ^a	12.6 ± 0.3^{a}	30.0 ± 0.1^{a}	35.2 ± 0.5 ^a	34.9 ± 7.1^{a}	20.4 ± 0.2^{b}	35.6 ± 0.5^{a}	48.1 ± 8.1 ^ª
	LD	17.0 ± 0.1 ^b	12.5 ± 0.1^{a}		-34.7 ± 4.1ª	13.0 ± 0.3 ^c	33.2 ± 0.4ª	35.8 ± 0.2 ^a	42.2 ± 6.8 ^a	21.0 ± 0.5ª	36.2 ± 0.3^{a}	52.2 ± 10.1ª
AMF+	MD	18.1 ± 0.1 ^b	12.5 ± 0.7 ^c	8.8 ± 0.1^{a}	-36.0 ± 2.8^{bc}	12.8 ± 0.2 ^b	32.8 ± 1.9^{a}	35.6 ± 0.5 ^a	38.7 ± 6.5ª	21.2 ± 0.2 ^{ab}	36.2 ± 0.1^{a}	64.3 ± 3.5 ^a
2.5 wt% 16:0	SD	17.0 ± 1.4 ^b	4.7 ± 0.4^{b}		-27.2 ± 3.6 ^{ab}	12.6 ± 0.3^{a}	30.2 ± 0.1^{a}	34.8 ± 0.5 ^a	40.0 ± 8.1^{a}	20.8 ± 0.2^{b}	35.6 ± 0.5^{a}	49.7 ± 8.4^{a}
	LD	23.3 ± 0.4 ^c	11.2 ± 0.1^{c}		-41.4 ± 3.6^{bc}	16.0 ± 0.4^{b}	32.6 ± 0.4^{a}	35.6 ± 0.2 ^a	50.1 ± 8.1 ^a	21.0 ± 0.5ª	41.0 ± 0.4^{b}	59.6 ± 11.5ª
AMF+	MD	21.6 ± 0.6 ^c	19.2 ± 0.5^{d}	9.0 ± 0.6^{a}	-36.0 ± 3.2^{bc}	17.2 ± 0.3 ^c	33.4 ± 1.9ª	35.2 ± 0.5ª	45.7 ± 7.7ª	21.5 ± 0.2ª	49.0 ± 0.1 ^c	59.8 ± 3.2ª
5 wt% 16:0	SD	21.3 ± 1.0 ^c	16.3 ± 0.4^{d}	$4.6 \pm 0.1^{\circ}$	-23.8 ± 4.3 ^{ab}	13.0 ± 0.3^{a}	30.6 ± 0.1 ^a	35.4 ± 0.5 ^ª	$48.4 \pm 9.8^{\circ}$	21.8 ± 0.2^{a}	36.0 ± 0.5^{a}	42.5 ± 7.2 ^ª
	LD	21.3 ± 0.4^{d}	14.0 ± 0.1^{b}	8.1 ± 0.1 ^a	-42.9 ± 2.2 ^c	16.0 ± 0.4 ^b	38.8 ± 0.4 ^b	41.8 ± 0.3 ^b	54.2 ± 8.8 ^a	21.2 ± 0.5 ^a	$46.0 \pm 0.4^{\circ}$	71.1 ± 13.7 ^a
AMF+	MD	18.4 ± 0.2 ^b	13.5 ± 0.1 ^c	7.7 ± 0.1^{b}	-38.4 ± 1.2 ^c	16.8 ± 0.3 ^c	39.2 ± 2.3 ^b	45.6 ± 0.6 ^b	49.2 ± 8.3ª	21.4 ± 0.2 ^{ab}	46.0 ± 0.1^{b}	63.7 ± 3.4ª
5 wt% PPP	SD	15.1 ± 0.6 ^b	9.7 ± 0.4 ^c		-36.2 ± 1.8 ^c	17.4 ± 0.4 ^b	33.2 ± 0.1 ^b	45.6 ± 0.6 ^b	43.3 ± 8.8 ^a	21.6 ± 0.2ª	45.5 ± 0.6 ^b	49.8 ± 8.4 ^a

Table 2

¹ T_{onset} : initial crystallization temperature upon cooling; T_{offset} : upper melting temperature.

² When thermograms present several endothermic or exothermic events, T_{peak} is taken at the maximum of each peak. Only the position of the most intense peaks is reported.

 ${}^{3}\Delta H$: melting or crystallization enthalpy variations, determined from the total area of the thermographs between 4 °C and Tonset or Toffset, the temperature at which the DSC signal leaves or reaches the baseline.

⁴ Values are expressed as mean \pm standard deviations. n = 3 independent repetitions for all crystallization experiments and for melting experiments with emulsions containing meat AMF. n=1 for melting experiments with emulsions containing modulators; for the statical analysis, the same relative standard deviations (SD/mean) as those of the emulsions based on neat AMF with comparable average droplet sizes were adopted. For a given droplet size, mean values with unlike superscript letters are significantly different (p < 0.05) using a one-way analysis of variance followed by a pairwise t-test. Within the neat AMF group, mean values with unlike symbols were significantly different (p < 0.05).

Figures



Figure 1







Α



Figure 3



Figure 4