

Importance of the vegetation-groundwater-stream continuum to understand transformation of biogenic carbon in aquatic systems — A case study based on a pine-maize comparison in a lowland sandy watershed (Landes de Gascogne, SW France)

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Abstract

- During land-aquatic transfer, carbon (C) and inorganic nutrients (IN) are transformed in soils,
- 26 groundwater, and at the groundwater-surface water interface as well as in stream channels and stream
- 27 sediments. However, processes and factors controlling these transfers and transformations are not well
- 28 constrained, particularly with respect to land use effect. We compared C and IN concentrations in shallow
- 29 groundwater and first-order streams of a sandy lowland catchment dominated by two types of land use:
- 30 pine forest and maize cropland. Contrary to forest groundwater, crop groundwater exhibited oxic

conditions all-year round as a result of higher evapotranspiration and better lateral drainage that decreased the water table below the organic-rich soil horizon, prevented the leaching of soil-generated dissolved organic carbon (DOC) in groundwater, and thus limited consumption of dissolved oxygen (O_2) . In crop groundwater, oxic conditions inhibited denitrification and methanogenesis resulting in high nitrate (NO₃; on average $1{,}140 \pm 485 \,\mu\text{mol}\,\,L^{-1}$) and low methane (CH₄; $40 \pm 25 \,\text{nmol}\,\,L^{-1}$) concentrations. Conversely, anoxic conditions in forest groundwater led to lower NO_3^- (25 ± 40 µmol L⁻1) and higher CH_4 (1,770 ± 1,830 nmol L⁻¹) concentrations. The partial pressure of carbon dioxide (pCO₂; $30,650 \pm 11,590$ ppmv) in crop groundwater was significantly lower than in forest groundwater ($50,630 \pm 26,070$ ppmv), and was apparently caused by the deeper water table delaying downward diffusion of soil CO₂ to the water table. In contrast, pCO₂ was not significantly different in crop $(4,480 \pm 2,680 \text{ ppmv})$ and forest $(4,900 \pm 4,500 \text{ ppmv})$ ppmv) streams, suggesting faster degassing in forest streams resulting from greater water turbulence. Although NO₃ concentrations indicated that denitrification occurred in riparian-forest groundwater, crop streams nevertheless exhibited important signs of spring and summer eutrophication such as the development of macrophytes. Stream eutrophication favored development of anaerobic conditions in crop stream sediments, as evidenced by increased ammonia (NH₄⁺) and CH₄ in stream waters and concomitant decreased in NO₃ concentrations as a result of sediment denitrification. In crop streams, dredging and erosion of streambed sediments during winter sustained high concentration of particulate organic C, NH₄⁺ and CH₄. In forest streams, dissolved iron (Fe²⁺), NH₄⁺ and CH₄ were negatively correlated with O₂ reflecting the gradual oxygenation of stream water and associated oxidations of Fe²⁺, NH₄⁺ and CH₄. The results overall showed that forest groundwater behaved as source of CO₂ and CH₄ to streams, the intensity depending on the hydrological connectivity among soils, groundwater, and streams. CH₄ production was prevented in cropland in soils and groundwater, however crop groundwater acted as a source of CO2 to streams (but less so than forest groundwater). Conversely, in streams, pCO₂ was not significantly affected by land use while CH₄ production was enhanced by cropland. At the catchment scale, this study found substantial biogeochemical heterogeneity in C and IN concentrations between forest and crop waters, demonstrating the importance of including the full vegetation-groundwater-stream continuum when

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- estimating land-water fluxes of C (and nitrogen) and attempting to understand their spatial and temporal dynamics.
- 59 Keywords: carbon dioxide, methane, groundwater, stream, land use, pine, maize

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1. Introduction

62 Despite their small surface area worldwide (Downing et al., 2012), inland waters have been recognized as 63 key component of the global carbon (C) cycle, constituting a preferential pathway of dissolved and particulate C transport from terrestrial ecosystems to the coastal ocean (Cole et al., 2007; Meybeck, 1982; 64 Ludwig et al., 1996b, 1996a; Meybeck, 1987). Inland waters act as significant sources of carbon dioxide 65 (CO₂) and methane (CH₄) to the atmosphere because inland waters are generally supersaturated by CO₂ 66 67 and CH₄ compared to the overlying atmosphere (Abril et al., 2014; Bastviken et al., 2011; Borges et al., 2015; Lauerwald et al., 2015; Raymond et al., 2013; Stanley et al., 2016). 68 Inland waters and specifically small streams are tightly connected to their catchment characteristics such 69 70 as hydrology and land use, as they receive large inputs of C from land (mainly from soils and groundwater), which in turn control the stream biogeochemical processes and the water composition 71 72 (Aitkenhead et al., 1999; Deirmendjian and Abril, 2018; Hotchkiss et al., 2015; Johnson et al., 2006; Jones 73 and Mulholland, 1998; McClain et al., 2003; Polsenaere and Abril, 2012; Bodmer et al., 2016; Findlay et 74 al., 2001; Lehrter, 2006). Groundwater discharge has been recognized as an important source of CO₂ in 75 riverine systems, especially in small streams and headwaters (Deirmendjian and Abril, 2018; Hotchkiss et 76 al., 2015; Johnson et al., 2008; Kokic et al., 2015; Marx et al., 2017; Raymond et al., 2013; Wallin et al., 77 2013). On the contrary to riverine CO₂, riverine CH₄ is likely to originate from wetlands that generally 78 combine a strong hydrological connectivity with riverine waters and a high productivity (Abril et al., 79 2014; Abril and Borges, 2018). Although some studies found low CH₄ concentrations in the groundwater of Belgium (up to 1.1 µmol L⁻¹; Borges et al., 2018; Jurado et al., 2017), other studies found high CH₄ 80

concentrations in the groundwater of Great Britain (up to 295 µmol L⁻¹; Bell et al., 2017) and in the Appalachian basin of the USA (up to 2,8000 umol L⁻¹; Molofsky et al., 2016). Actually, soil moisture, which controls oxic/anoxic conditions in soil, is the main determinant of terrestrial CO_2 or CH_4 production in soil. As a consequence, CH₄ emissions from soils are high under strictly anaerobic conditions in waterlogged soils whereas CO₂ emissions from soils are high under aerobic conditions in drier soils (Christensen et al., 2003; Moore and Knowles, 1989). Croplands affect water mass balance at the plot scale, especially through irrigation and extraction of groundwater, which results in declining water table in many regions worldwide (Foley et al., 2005; Gleick, 2003; Jackson et al., 2001; Postel, 1999; Rosegrant et al., 2002). Investigating spatial dynamics of CO₂ and CH₄ in groundwater in relation with land use is critical better understanding processes governing their terrestrial production and leaching to groundwater. Croplands cover about 40% of the terrestrial ice-free surface and are often associated with degradation of both ground and surface water quality (Asner et al., 2004; Clague et al., 2015; Foley et al., 2005; Hiscock et al., 1991; Ramankutty and Foley, 1999). Intensive agriculture led to an increase of nitrate (NO₃) entering ground and surface water environments that has fueled aquatic primary production in surface waters and led to low CO₂ and high CH₄ concentrations, the latter being related to enhanced organic matter delivery in sediments (Borges et al., 2018; Carpenter et al., 1998; Clague et al., 2015; Crawford et al., 2016; Jordan and Weller, 1996; Smith, 2003; Zhou et al., 2017). Additionally, aquatic primary production in crop streams is enhanced as a result of low light limitation (clearing of riparian vegetation), and the excessive transport of sediment-bound organic matter and nutrients to surface waters (Bernot et al., 2010; Lamba et al., 2015; Ramos et al., 2015; Young and Huryn, 1999). Soil erosion rates in agricultural landscapes are one to two times larger than those in areas with native vegetation (Montgomery, 2007; Quinton et al., 2010). Indeed, riparian forest is usually considered stream buffer zones that attenuate stream bank erosion and NO₃ inputs from croplands (Balestrini et al., 2016; Cey et al., 1999; Christensen et al., 2013; Stott, 2005; Wynn and Mostaghimi, 2006). Denitrification represents a permanent removal pathway that limits the extent and impact of NO₃ contamination by transforming NO₃

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to inert dinitrogen (N_2) . However, incomplete denitrification can produced nitrous oxide (N_2O) , a major anthropogenic ozone-depleting substance (Ravishankara et al., 2009). On the contrary to croplands, forests are known to export fewer nutrients by limiting runoff and leakage of nutrients (Canton et al., 2012; Onderka et al., 2010).

Land use effects on both water composition and biogeochemical processes have been studied in streams and groundwater (Barnes and Raymond, 2010, 2009; Bernot et al., 2010; Bodmer et al., 2016; Jeong, 2001; Lehrter, 2006; Masese et al., 2017; Raymond and Cole, 2003; Rodrigues et al., 2018; Salvia-Castellví et al., 2005; Vidon et al., 2008; Wilson and Xenopoulos, 2009; Young and Huryn, 1999; Zhang et al., 2018), but land use studies with simultaneous groundwater and stream sampling are more scarce (Bass et al., 2014; Borges et al., 2018; Hu et al., 2016). The objective of this study was to understand how two contrasting types of land use (pine forest and maize cropland) affected C and inorganic nutrient (IN) concentrations in shallow groundwater and in first-order streams of a sandy lowland catchment. We hypothesized that the biogeochemical variability between crop groundwater and forest groundwater was due to agricultural practices that affect N inputs (fertilizer) and water mass balance (irrigation and drainage). We hypothesized that the biogeochemical variability between crop and forest streams originate from differential lateral export of C and IN from two distinct sources (i.e., crop groundwater and forest groundwater) because of a strong hydrological connection between groundwater and streams in the studied catchment.

2. Materials and Methods

2.1. Study site

The Leyre catchment (2,100 km²) is located in the southwestern part of France. This is a flat coastal plain with a mean slope lower than 0.125% and a mean altitude lower than 50 m (Jolivet et al., 2007). The lithology is relatively homogeneous and composed of sandy permeable surface layers dating from the

Plio-Quaternary period (Legigan, 1979; Bertran et al., 2009, 2011). The soils are podzols characterized by a low pH (\approx 4), low nutrient availability, low cationic exchange capacity, and high organic C content that can reach 50 g per kg of soil (Augusto et al., 2010; Lundström et al., 2000). In Leyre sandy podzols, the low clay and silt content causes a low soil water retention (Augusto et al., 2010). The study area was a vast wetland until the 19th century, when a wide forest of maritime pine was sown following a landscape drainage campaign resulting from an imperial decree of Napoleon III in 1857 (Jolivet et al., 2007). Currently, the catchment is mainly occupied by C₃ pine forest (approximately 85%), with a modest proportion of C₄ maize cropland (approximately 15%) (Fig. 1; Jolivet et al., 2007). Following catastrophic forest wildfires, the maize croplands were installed during the second half of the 20th century. Consequently, their spatial distribution was not based on soil properties, as confirmed by the similar mean values of soil texture in local croplands and forests (Augusto et al., 2010; Jolivet et al., 2003). During the maize cropping season (usually May to November), farmers irrigate the plots by pumping shallow groundwater (~1-5 m deep) almost daily to maintain adequate soil moisture status, whereas maritime pine stands are never irrigated (Govind et al., 2012). As N is not limiting for tree growth in our study region (Trichet et al., 2009), forests are never fertilized with N. Conversely, croplands generally receive two N fertilizer applications annually, a first at the beginning of May (30–50 kg N ha⁻¹), and second at the beginning of June with 200–250 kg N ha⁻¹ (Canton et al., 2012; Jambert et al., 1997; Ulrich et al., 2002). Additionally, in order to maintain soil pH in the 5.5–6.0 range, local maize croplands are limed with crushed limestone (CaCO₃) containing a small portion of dolomite (CaMg(CO₃)₂) (10 t ha⁻¹ right after forest conversion and then 0.5 t ha⁻¹ an⁻¹; Jolivet et al., 2003). The climate is oceanic with a mean annual air temperature of 13°C and a mean annual precipitation of 930 mm (Moreaux et al., 2011). Owing to the low slope, the low soil water retention and the high permeability of the soil (i.e., hydraulic conductivity is approximately 40 cm h⁻¹, Corbier et al., 2010), the percolation of rain water is fast (55 cm h⁻¹ on average, Vernier and Castro, 2010). Consequently, surface runoff does not occur as the excess of rainfall percolates into the soil and recharges the shallow groundwater, causing the

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water table to rise. The sandy permeable surface layers contain a free and continuous water table that is strongly interconnected with the superficial river network. This is facilitated by a dense network of drainage ditches, initiated in the 19th century and currently maintained by forest managers in order to enhance tree regeneration and growth (Thivolle-Cazat and Najar, 2001). During the sampling period, channels of some crop streams were dredged before they began to flow again. This was done to optimize local cropland drainage and to feed croplands with IN and organic residuals found in the stream sediments. To increase soil permeability and to optimize lateral drainage in local maize croplands, farmers practice subsoiling and agricultural ditches are generally deeper (2.0–2.5m) than forest ditches (1.0m).

2.2. Sampling strategy

We defined order 0 as groundwater and order 1 as streams and ditches either having no tributaries or being seasonally dry (from June to December during our sampling period). We selected 17 sampling stations (5 shallow groundwater and 12 first-order streams) within the Leyre catchment (Tab. 1; Fig. 1). The groundwater sampling stations were located in maize cropland (n=2), pine forest (n=2; one is the Bilos station (FR-Bil) of the ICOS Research infrastructure) and in a riparian forest adjacent to a maize cropland (n=1; Tab. 1; Fig. 1). The stream sampling stations were chosen based on the different proportions of croplands in their respective catchments (Tab. 1; Fig. 1). Groundwater was sampled for temperature, electrical conductivity (EC), pH, dissolved oxygen (O₂), methane (CH₄), partial pressure of CO₂ (pCO₂), total alkalinity (TA), dissolved inorganic carbon (DIC), stable isotope composition of the dissolved inorganic carbon (δ^{13} C-DIC), dissolved organic carbon (DOC), ammonia (NH₄⁺), nitrate (NO₃⁻) and dissolved iron (Fe²⁺). For groundwater, we took the precaution to renew the water in the piezometers by pumping with a submersible pump before sampling. Groundwater was then sampled once the stabilization (approximately 10 min) of groundwater temperature, pH, EC and O₂ monitored with portable probes was observed. Streams were sampled for the

same parameters, plus total suspended matter (TSM), particulate organic carbon (POC) and the POC content of the TSM (POC%).

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2.3. Field measurements and laboratory analyses

Groundwater and streams were sampled at approximately monthly time intervals between Jan. 2014 and Jul. 2015 (Tab. S1). In total, throughout the sampling period, we sampled 55 groundwaters and 137 stream waters. The pCO₂ in groundwater and streams was measured directly using an equilibrator (Frankignoulle and Borges, 2001; Polsenaere et al., 2013) following the procedure of Deirmendjian and Abril (2018). We stored the total alkalinity (TA) samples in polypropylene bottles after filtration using a syringe equipped with glass fiber filters (GF/F; 0.7 µm). TA was then analyzed on filtered samples by automated electro-titration on 50 mL samples with 0.1N HCl as the titrant. The equivalence point was determined from pH between 4 and 3 with the Gran method (Gran, 1952). Precision based on replicate analyses was better than $\pm 5 \mu \text{mol L}^{-1}$. For samples with a very low pH (<4.5), we bubbled the water with atmospheric air in order to degas the CO₂. Consequently, the initial pH increased above 5, and the TA titration was then performed (Abril et al., 2015). We calculated DIC from pCO₂, TA, and temperature measurements using the carbonic acid dissociation constants of Millero (1979) and the CO₂ solubility from Weiss (1974), using the CO₂SYS software (Lewis et al., 1998). The δ^{13} C-DIC samples were collected using 120 mL glass serum bottles sealed with a rubber stopper and treated with 0.3 mL of HgCl₂ at 20 g L⁻¹ to avoid any microbial respiration during storage. Vials were carefully sealed such that no air remained in contact with samples and were stored in the dark to prevent photo-oxidation. The δ^{13} C-DIC measurements were performed with the headspace technique using an isotope ratio mass spectrometer coupled to an elemental analyser (EA-IRMS, Micromass IsoPrime) equipped with a manual gas injection port as described in Gillikin and Bouillon (2007).

203 CH₄ was also measured using a headspace technique in 60 mL glass serum bottles. The headspace was 204 created with 10 mL of N₂ gas. We then injected 0.5 mL of the headspace in a gas chromatograph equipped 205 with a flame ionization detector (GC-FID). 206 DOC samples were obtained after filtration in the field through pre-combusted GF/F (0.7 µm). DOC filtrates were stored in pre-combusted Pyrex vials (25 mL), acidified with 50 µL of 37% HCl to reach pH 207 208 2, and kept at 4 °C in the laboratory before analysis. The DOC concentrations were measured with a 209 SHIMADZU TOC 500 analyzer (in TOC-IC mode), using a technique based on thermal oxidation after a DIC removal step (Sharp, 1993). The repeatability was better than 0.1 mg L⁻¹. 210 211 The water for TSM and POC measurements was filtered through pre-weighed and pre-combusted GF/F 212 glass fiber filters (0.7 µm). The filters were dried at 60 °C and stored in the dark, and subsequently, TSM 213 was determined by gravimetry. POC was measured using the same filter. The filters were acidified in 214 crucibles with 2N HCl to remove carbonates and were then dried at 60 °C to remove inorganic carbon and 215 most of the remaining acid and water (Etcheber et al., 2007). POC content was measured by combustion 216 (1500 °C) using a LECO CS 200 analyzer and the CO₂ formed was determined quantitatively by infrared absorption. POC in μ mol L⁻¹ and POC% were then calculated. The uncertainty was $\pm 0.05\%$ of TSM. 217 218 For IN determination, water was filtered through a 0.20 µm cellulose acetate syringe membrane. Subsamples for Fe²⁺ were acidified with 37% HCl to prevent precipitation of iron oxide, whereas 219 subsamples for NH₄ and NO₃ were not acidified but kept frozen until later analyses. Then, NH₄, NO₃, 220 and Fe²⁺ were analyzed by colorimetry according to standard techniques. NH₄⁺ was analyzed following the 221 222 procedure of Harwood and Kühn (1970). NO₃ was analyzed by flow injection analysis following the procedure of Anderson (1979). Fe²⁺ was analyzed using the ferrozine method (Stookey, 1970). Precision 223 was $\pm 10\%$ for NH₄⁺ and NO₃, and was $\pm 5\%$ for Fe²⁺. 224 EC, temperature, O₂, and pH were measured using portable probes (WTW®). Before each field trip, the 225 226 pH probe was calibrated using two NBS buffer solutions (4 and 7), the oxygen polarographic probe was

calibrated to 100% in a humid atmosphere and the conductivity probe was calibrated using a salinity standard.

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2.4. Statistical analyses

K-means clustering analysis (MacQueen, 1967) was used to classify waters either as forest-dominated or as cropland-affected (Tab. 1). Indeed, K-means clustering analysis allows partitioning a dataset into k groups (i.e., clusters) pre-specified by the analyst (MacQueen, 1967). Contrary to forest waters at our study site, crop waters exhibit disproportionately higher NO₃ concentration as a result of N fertilizer use on maize cropland (Canton et al., 2012; De Wit et al., 2005; Jambert et al., 1997, 1994). Consequently, in the K-means clustering analysis we used NO₃ concentration data as a proxy to establish a statistical distinction between forest and crop waters (Tab. 1). K-means clustering analysis was performed one time with the groundwater dataset (but excluding the riparian groundwater) and a second time with the firstorder streams dataset. We excluded data from riparian groundwater because we have considered riparian groundwater as a cluster itself (Tab. 1). Principal component analysis (PCA) was used to condense multivariate information on correlated biogeochemical parameters to a set of uncorrelated variables called principal components (further referred to as dimensions). PCA was performed one time with a dataset consisting of each measured parameter in groundwater (but excluding the riparian groundwater) and a second time with the corresponding firstorder streams dataset. PCA was performed separately for groundwater and streams because particulate parameters were not present in groundwater. If PCA were not performed separately for groundwater and streams, all data from groundwater would have been removed from the analysis (indeed, if one parameter is missing for a given sampling station, the sampling station is entirely deleted from the PCA). In addition, performing the PCA separately for groundwater and streams led to information that was more robust with respect to the biogeochemical variability induced by land use, in either groundwater or streams. However,

to observe whether the two groundwater (crop and forest) and two streams (crop and forest) sources could be distinguished mathematically in one PCA, we performed an additional PCA with data from both groundwater and streams that excluded particulate parameters from the analysis. All concentrations data were log-transformed prior to PCA. The PCAs showed the biogeochemical variability across forest, cropland, and hydrological seasons in either groundwater or first-order streams.

Later, non-parametric bivariate analyses (Mann-Whitney statistical tests) were used to estimate if hydrological seasons or increasing stream order significantly influenced the concentration of a biogeochemical parameter. Linear regressions were performed to model the relationships between two variables by fitting a linear equation to observed data.

K-means clustering analysis (package Stats) and PCA analysis (package FactoMineR for analysis and package factoextra for visualization; Kassambra and Mundt, 2017; Lê et al., 2008) were performed with R software version 3.1.4 (R Core Team, 2018). Mann-Whitney tests and linear regressions were performed with Graph Pad Prism version 7 software.

3. Results

3.1. Hydrology

In previous work based on the same dataset, but excluding cropland sampling stations, we identified two major hydrological seasons (Deirmendjian and Abril, 2018). One defined a high flow period as two relatively short flood events that occurred in Jan. 2014–Mar. 2014 and in Feb. 2015–Mar. 2015, whereas we defined the base flow period as two longer periods of low flow occurring in Apr. 2014–Jan. 2015 and Apr. 2015–Jul. 2015. During high flow, the average and the maximum river flows were 50 m³ s⁻¹ and 119 m³ s⁻¹, respectively. During base flow, the average and the minimum river flow were 10 m³ s⁻¹ and 5 m³ s⁻¹, respectively. The water tables in the forest, riparian forest, and cropland exhibited similar temporal fluctuations but with a different intensity, and the forest had an overall higher water table depth than the

cropland (Fig. 2). The water table in the riparian area exhibited intermediate depth between the forest and cropland sites (Fig. 2). As surface runoff was negligible in the studied sandy and flat catchment, most of the stream water likely originated from groundwater discharge.

To investigate the temporal variability of the studied biogeochemical parameters, we chose to rely on hydrological regimes (high flow and base flow periods) rather than on temperature periods (seasons). At our study site climate was oceanic (by definition very temperate) and the amplitude of the water temperature was not as high as the amplitude of the river flow. As an example, Leyre River (main stem) flow could be up to 119 m³ s⁻¹ and could be down to 5 m³ s⁻¹, whereas the highest water temperature amplitude occurred in forest streams and was 6.4–25.8 °C (Tab. 2). Additionally, most of the lateral export occurred during the short periods of high flow (up to 90% for DOC, Deirmendjian et al., 2018). Thus, characterizing biogeochemical variability and biogeochemical processes in relation with land use during this hydrological period was important. Furthermore, the seasonality induced by water temperature was to a certain extent included in the defined hydrological regimes since the high flow period was associated with lower water temperatures while the base flow period was associated with higher water temperatures (Tabs. 2-4).

3.2. K-means clustering analysis

For both hydrological seasons (i.e., high and base flow), we partitioned each sampling stations (excepting riparian groundwater), into either cropland-affected or forest-dominated waters (Tab. 1). K-means clustering analysis produced satisfactory results. Logically, groundwater located in cropland was classified as crop water whereas groundwater located in forest was classified as forest water (Tab. 1). Stream sampling stations having more than 30% of croplands in their respective catchment were always classified as crop waters (Tab. 1). Stream sampling stations having less than 8% of croplands in their respective catchment were always classified as forest waters excepting two times (Tab. 1). These two stream

sampling stations were located a few kilometers downstream from important maize croplands.

Specifically, one station was a ditch strongly vegetated during the base flow period that showed signs of N fertilizer uptake from upstream cropland and, therefore, this ditch was logically classified as a crop station during base flow (Tab. 1). One station was a stream that exhibited a high water flow during the high flow period, which probably increased the upstream cropland influences during this hydrological period and, therefore, this stream was logically classified as a crop station during high flow (Tab. 1).

Excepting one strictly forested headwater, the other sampled streams were not strictly forested or cropped (Tab. 1). Consequently, as explained further, some biogeochemical variability between forest and crop streams was introduced by simple water mixing from two distinct sources: forest groundwater and crop

groundwater. We used the term crop stream to indicate a stream classified as a crop-affected one, although

3.3. Land use influence on water composition of shallow groundwater

such stream was a forest stream affected by cropland rather that a strictly crop stream.

PCA on the groundwater dataset revealed that groundwater biogeochemical variability was strongly dependent on land use (maize cropland vs. pine forest) and hydrological seasons (base flow vs. high flow) (Figs. 3a, b). The first three PCA dimensions covered 44%, 17.5% and 10.5% of the total variance within the dataset, respectively (Figs. 3a, b).

PCA dimension 1 clearly separated forest groundwater from crop groundwater based on two groups of variables negatively correlated with one another (Fig. 3a). One group of variables was characterized crop groundwater and was composed of EC, NO₃-, δ¹³C-DIC, and O₂, whereas the second group of variables was characterized forest groundwater and was composed of DIC, pCO₂, CH₄, Fe²⁺, and NH₄+ (Fig. 3a). Indeed, we observed that the yearly average of EC (+270 μS cm⁻¹), NO₃- (+1,115 μmol L⁻¹), δ¹³C-DIC (+6.9‰), and O₂ (+200 μmol L⁻¹) were higher in crop groundwater than in forest groundwater and were significantly and positively affected by cropland cover (Tab. 2; Figs. 4b, c, f, j). Conversely, we observed

higher DIC (+1,010 μ mol L⁻¹), pCO₂ (+19,985 ppmv), CH₄ (+1,730 μ mol L⁻¹), Fe²⁺ (+14.1 μ mol L⁻¹), and 323 NH_4^+ (+4.1 µmol L⁻¹) in forest groundwater than in crop groundwater; these were significantly and 324 positively affected by forest cover (Tab. 2; Figs. 4d, e, g, h, k). In riparian groundwater, EC, NO₃, δ¹³C-325 DIC, O₂, DIC, pCO₂, and CH₄ exhibited intermediate values between the groundwater of forest and crop 326 sites, whereas Fe²⁺ and NH₄⁺ were low and close to those found in crop groundwater (Tab. 2; Figs. 4b, c, 327 328 d, e, f, g, h, j, k). 329 In crop groundwater, EC, NO_3^- , δ^{13} C-DIC, and O_2 were not significantly affected by hydrological seasons (Figs. 3a, S2c, h, j, k, l). However, δ^{13} C-DIC (+1.1%), NO₃⁻ (+120 μ mol L⁻¹), and DOC (+130 μ mol L⁻¹) 330 were slightly higher (but not significantly) during base flow compared to high flow (Tab. 3; Figs. S2c, h, j, 331 332 k, l). In forest groundwater, pCO₂, DIC, CH₄, and DOC were significantly affected by hydrological 333 seasons (Tab. 3; Figs. S2g, h, k, l). DOC (+1,490 µmol L⁻¹) was significantly higher during high flow, whereas pCO₂ (+30,980 ppmv), DIC (+1,330 μ mol L⁻¹) and CH₄ (+1,780 nmol L⁻¹) were significantly 334 335 higher during base flow (Tab. 3; Figs. S2 and S4g, h, k, l). In crop and riparian groundwater, we also 336 observed higher pCO₂ and DIC values during base flow, but with lower intensities than in forest 337 groundwater (Tab. 3; Figs. S2h, k).

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3.4. Land use influence on water composition of first-order streams

Figures 3c-d present PCA based on first-order streams data set. The first three PCA dimensions covered 28.6%, 18.5% and 13.8% of the total variance within the dataset, respectively (Figs. 3c, d).

Interestingly, the PCA based on the first-order streams dataset did not clearly separate crop streams from forest streams as it did for groundwater dataset (Figs. 3a-d). This implied lower spatial variability in streams in relation to land use in than in groundwater (Tabs. 2-4; Fig. 4). Nevertheless, a land use gradient was observed on PCA dimension 2 (Figs. 3c, d). PCA dimension 2 was best defined by a group of variables composed of EC, CH₄, NO₃-, NH₄+, DOC, TSM, and POC, which collectively characterized crop

streams (Figs. 3c, d). On a yearly average basis, significantly higher EC (+105 µS cm⁻¹), CH₄ (+220 µmol L^{-1}), NO_3^{-1} (+265 µmol L^{-1}), NH_4^{+1} (+4.3 µmol L^{-1}), DOC (+135 µmol L^{-1}), TSM (+3.3 mg L^{-1}), and POC(+70 μmol L⁻¹) were observed in crop streams compared to forest streams (Tab. 2; Figs. 4b, c, d, g, l, m, o). High CH₄, NH₄⁺, and DOC concentrations were characteristics of forest groundwater, but in streams, these parameters were characteristic of crop streams (Figs. 3a-d). In addition, pCO₂, DIC, δ^{13} C-DIC and O₂ were not able to separate crop streams from forest streams as they did for groundwaters (Figs. 3a-d). On a yearly average basis, no significant differences were observed between crop and forest streams for these four parameters (Figs. 3f, h, j, k). The relatively low temporal variability between high and base flow periods observed in both crop and forest streams for the studied parameters did not allow the PCA based on stream data set to clearly separate base flow samples from high flow samples (Tab. 4; Figs. 3c-d, S3, S4). Nevertheless, in crop streams, pH (+0.2) and δ^{13} C-DIC (+3.8%) were significantly higher during base flow while pCO₂ (-1,160 ppmv), NO₃⁻ (-130 µmol L⁻¹) and DOC (-230 µmol L⁻¹) were significantly lower during the same period (Tab. 4; Figs. S3a, j, c, h, l, m, o). In streams, TSM and POC were significantly higher during high flow but with a higher intensity in crop streams than in forest streams (Tab. 4; Fig. S3m, o). Interestingly, in both forest and crop streams, POC% was not significantly affected by hydrological regime (Tab. 4; Fig. S3n).

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3.5. Upstream-downstream distribution of biogeochemical parameters

To explore the influence of land use on water composition at the groundwater-stream continuum, we observed the upstream-downstream (groundwater-stream) distribution of biogeochemical parameters along forest and crop continuums (Tab. 2; Fig. 4). Along both types of continuum, some parameters (i.e., pCO₂, TA, DIC, δ^{13} C-DIC, pH, O₂) exhibited the same upstream-downstream distribution, whereas other

parameters (i.e., EC, NO₃, NH₄⁺, Fe²⁺, CH₄, DOC) exhibited a different upstream-downstream distribution (Tab. 2; Fig. 4).

In crop and forest continuums, we observed strong spatial patterns for pCO₂, TA, DIC, δ¹³C-DIC, and pH:

pCO₂ and DIC decreased while TA remained more or less constant, and δ^{13} C-DIC and pH increased (Tab. 2; Figs. 4a, h, i, j, k). However, a larger decrease in pCO₂ levels in the forest continuum suggested a more intense degassing in forest streams (Tab. 2; Fig. 4h). We also observed an increase in O₂ in both types of continuum, which could result from stream ventilation, although with a higher intensity in forest continuum (Tab. 2; Figs. 4f, h).

EC decreased downstream in the crop continuum, but did not in the forest continuum where EC remained very stable and much lower than in the crop continuum (Tab. 2; Fig. 4b). NO₃⁻ decreased downstream between groundwater and streams in croplands, and in contrast, in forests NO₃⁻ increased downstream between groundwaters and streams (Tab. 2; Fig. 4c). NH₄⁺, Fe²⁺, and CH₄ decreased in the forest continuum but they increased in the crop continuum (Tab. 2; Figs. 4d, e, g). DOC significantly decreased in the forest continuum but remained stable in the crop continuum (Tab. 2; Fig. 4l). TSM and POC were significantly higher in crop relative to forest streams; however, similarly high POC% (28%) was observed in both types of streams (Tab. 2; Fig. 4m, n, o).

3.6. Biogeochemistry dynamics in the groundwater-stream continuum

PCA with groundwater and streams datasets indicated mathematically that streams were fed with two distinct sources: forest groundwater mostly characterized by high pCO₂, DIC, and CH₄ concentrations and crop groundwater mostly characterized by high NO₃⁻ concentrations (Figs. 5). Forest and crop streams were characterized by higher O₂, δ^{13} C-DIC and pH values than in groundwater (Figs. 5). In this PCA, the distinction between forest and crop streams was primarily a function of NO₃⁻, crop streams points were moved upward along dimension 2 (Figs. 5). Throughout the sampling period, we observed a negative

394 linear relationship (R²=0.6, p<0.001, n=192) between CO₂ and O₂ for all sampled groundwater and 395 streams (Fig. 6a). On the one side, stream samples were mostly characterized by high O₂ (mean was 290 396 umol L⁻¹) and low CO₂ (mean was 4,480 ppmv), excepting some forest streams during summer that were characterized by low O₂ (down to 110 µmol L⁻¹) and high CO₂ (up to 27,200 ppmv) (Tab. 2; Figs. 6a, S4f, 397 398 h). On the other side, anoxic conditions associated with high CO₂ (mean was 50,630 ppmv) were 399 characteristic of forest groundwater, whereas crop groundwater exhibited O₂ (mean was 220 µmol L⁻¹) and 400 CO₂ (mean was 30,650 ppmy) intermediate between streams and forest groundwater (Tab. 2; Fig. 6a). In 401 forest groundwater, DOC was negatively and linearly correlated with CO_2 (R²=0.4, p < 0.001, n=22) 402 suggesting that part of groundwater CO₂ came from degradation of groundwater DOC (Fig. 6b). A 403 comparison of CO₂ and CH₄ for all sampled groundwaters and streams showed that a large portion of the 404 CO₂ and CH₄ in forest streams could come from forest groundwater discharge (Tab. 2; Fig. 6e). In crop streams, CH₄, NH₄⁺, and Fe²⁺ could not originate from crop groundwater discharge since they had much 405 lower CH₄, NH₄⁺, and Fe²⁺ concentrations than crop streams (Tab. 2; Fig. 6e, 7e, 7f). We observed a 406 407 positive linear relationship between CH₄ and NH₄⁺ (R²=0.4, p<0.001, n=53) in crop streams, 408 demonstrating that these two compounds may come from the same source (Fig. 7d). Conversely, a comparison of O₂ and NH₄⁺ or Fe²⁺ in forest streams indicated that NH₄⁺ and Fe²⁺ were mostly discharged 409 through forest groundwater (Fig. 7e, f). In forest streams, the negative linear relationship between O₂ and 410 NH_4^+ (R²=0.1, p-value<0.05, n=70), Fe²⁺ (R²=0.5, p-value<0.001, n=77), or CH_4 (R²=0.1, p-value<0.001, 411 412 n=77) suggested oxidation of these reduced compounds in the stream water column (Fig. 7e, f). We 413 observed a gradient of NO₃ concentration, from high values to low values, between crop groundwater (mean was 1,140 μmol L⁻¹), to riparian groundwater and crop streams (310 and 340 μmol L⁻¹, 414 415 respectively), to forest streams (75 µmol L⁻¹) and to forest groundwater (25 µmol L⁻¹) (Tab. 2; Fig. 7a, b, 416 c). In crop streams, a large share of riverine NO₃ could be discharged through crop groundwater. 417 Conversely, NO₃ concentration in forest streams could not be explained by NO₃ concentration in forest 418 groundwater (Tab. 2; Fig. 7a, b, c, d). In crop groundwater, high NO₃ concentrations were associated with

low CH₄ concentrations. In crop streams, high NO₃ concentrations could be related to high CH₄ concentrations (Fig. 7c).

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4. Discussion

4.1. Water table depth in relation to land use

At the studied catchment scale, lithology, topography, soils, and precipitation are more or less uniform (Augusto et al., 2010; Jolivet et al., 2003). At the plot scale, spatial variability of water table depth in relation to land use was thus necessarily dependent on how water outputs (drainage, evapotranspiration or groundwater storage) of the water mass balance were human-affected (Govind et al., 2012; Stella et al., 2009). Local forests are never irrigated, conversely, irrigation with extraction of groundwater (that decreases groundwater storage) in local croplands could strongly bias the water mass balance at the plot scale since about half of the water diverted for irrigation is rapidly consumed through evapotranspiration (e.g., Jackson et al; 2001). Additionally, evapotranspiration in maize croplands is typically higher than in forests owing to the larger stomatal conductance that makes the exchange of C and water between the biosphere and the atmosphere much easier (Govind et al., 2012; Stella et al., 2009). Other studies have shown that the combination of subsoiling practices (increasing soil permeability) with deep agricultural ditches in croplands also affected water mass balance at the plot scale by enhancing lateral drainage of groundwater (Evans et al., 1996; Robinson et al., 1985). From an 8 year survey of local cropland, Juste et al (1982) showed that lateral drainage strongly affected the water mass balance at the plot scale as it represented an annual mean of 637 mm (70% of the amount of precipitation), whereas precipitation was estimated at 922 mm. At the forest plot scale, lateral drainage represented an annual mean of 182 mm (20% of the amount of precipitation), whereas precipitation was estimated at 895mm (Deirmendjian et al, 2018). At the study site, deeper water table in croplands was thus a consequence of a higher evapotranspiration and more lateral drainage than in forest. As explained further, water table depth is an

important determinant for understanding the biogeochemical variability in groundwater in relation to land use.

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4.2. Dynamic of O_2 , DOC, DIC and δ^{13} C-DIC in groundwater: a combination of hydrological,

physical and metabolic processes

In other aquifer systems worldwide, several studies have observed a significant positive correlation between groundwater O₂ concentration and depth to water (Datry et al., 2004; Foulquier et al., 2010; Goldscheider et al., 2006; McMahon and Chapelle, 2008; Pabich et al., 2001). Where the water table is close to the soil surface, groundwater O₂ consumption is likely rapid because of incomplete degradation of soil-generated labile DOC in the unsaturated zone. On the contrary, where the water table is far from the soil surface, strong oxygen depletion in the vicinity of the water table does not occur since the higher residence time of infiltrating water results in almost complete degradation of soil-generated DOC in the unsaturated zone (Malard and Hervant, 1999; Starr and Gillham, 1993). A regional study in forest soils of Switzerland (Hagerdon et al., 2000) and a study compiling a global database of soil carbon (Camino-Serrano et al., 2014) both found that soil-generated DOC was preferentially mobilized under reducing conditions in soils because of dissolution of Fe oxides. Deeper water tables in croplands do not reach topsoil that exhibits high labile organic C content. Thus, reducing conditions in topsoil and the leaching of soil-generated DOC are prevented as is the consumption of the groundwater O₂ stock, as occurs in forests during high flow stages (Tab. 3; Fig. S2l; Deirmendjian et al. 2018). Therefore, groundwater pCO₂ was higher in the forest during high flow than it was in cropland and riparian sites (Tab.3; Figs. 4h, S2l, h). This also explains the negative correlation between DOC and CO₂ observed only in forest groundwater (Fig. 6b). During base flow, we observed a clear land use spatial pattern among cropland, riparian forest, and forest sites (Tab. 3; Fig. S2h). We hypothesize that this difference was a consequence of a less intense soil respiration in croplands during summer. From simultaneous eddy covariance measurements over pine forests and maize croplands of the study area, Stella et al (2009) confirmed that ecosystem respiration was

lower in croplands than in forests over the whole year. However, Stella et al (2009) also showed that ecosystem respiration was larger during the growing season of the maize, because of increased of soil respiration in response to the higher soil water content caused by irrigation. In forest sites, groundwater pCO₂ increases during the summer because soil CO₂ diffuses downward and then is dissolved into the water table (Deirmendjian et al., 2018; Tsypin and Macpherson, 2012). A deeper water table in cropland suggests a less efficient CO₂ transfer from soil air to water table. Higher soil moisture in croplands due to irrigation probably delays soil CO₂ diffusion to groundwater. The δ^{13} C-DIC signature of forest groundwater was typical of a signature that originated from respiration of soil organic matter derived from C₃ plants (O'Leary, 1988; Vogel et al., 1993). The studied forest soils have no natural carbonate minerals (Augusto et al., 2010) and DIC originating from silicate weathering has the same isotopic signature as DIC originating from soil respiration (Das et al., 2005; Polsenaere and Abril, 2012; Wachniew, 2006). Crop groundwater had a heavier δ^{13} C-DIC signature than forest groundwater and this discrepancy resulted from distinct processes. Liming in cropland brings artificial carbonates into crop soil and DIC originating from carbonate weathering produced DIC with a δ^{13} C value of approximately half of that of soil CO₂ as carbonate rocks have a δ^{13} C of approximately 0 %, making δ^{13} C-DIC less negative (Clark and Fritz, 1997; Salomons and Mook, 1986). Irrigation with extraction of groundwater could also increase the δ^{13} C-DIC signature by enhancing the degassing rate of 12 CO₂ relative to $^{13}\text{CO}_2$ (Deirmendjian and Abril, 2018; Polsenaere and Abril, 2012). Changes in the $\delta^{13}\text{C-DIC}$ signature could also originate from respiration of soil organic matter derived from maize, a C₄ plant with a heavier δ¹³C signature than C₃ forest plants (O'Leary, 1988; Vogel et al., 1993), as observed in the study region (Quénéa et al., 2006). Indeed, after three decades of cultivation, the remaining carbon from the forest pool was mostly recalcitrant and its degradation probably did not affect the δ^{13} C-DIC pool (Jolivet et al., 1997).

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4.3. Dynamics of IN and CH₄ in groundwater: the influence of groundwater O₂

Subsurface and groundwater redox zonation is driven by the spatial and temporal distribution of O₂ that serves as the primary terminal electron acceptor during the degradation of organic C. In crop groundwater, high O₂ concentrations inhibited methanogenesis, as this process is strictly anaerobic and thus resulted in very low CH₄ concentrations (Tab. 2; Figs. 4f, g; Borges et al., 2018; Ciais et al., 2010; Jurado et al., 2017; Klüber and Conrad, 1998). Conversely, forest and riparian groundwater exhibited anoxic conditions that allowed methanogenesis to occur and created higher CH₄ concentration in forest sites compared to cropland sites (Tab. 2; Figs. 4f, g). High water table stages in forested areas cause anoxia in soils, forcing plants and microorganisms to switch to anaerobic metabolism (Naumburg et al., 2005; Bakker et al., 2006, 2009). Thus, in riparian and forest areas, we expected a positive relationship between water table and groundwater CH₄ but, to the contrary, we observed a negative relationship between these two parameters $(R^2 = 0.25, p < 0.05; data not shown)$. This implies that methanogenesis primarily occurs in deeper layers of forest soils, especially in summer. Fe²⁺ and NH₄⁺ accumulates in forest groundwater because anoxic conditions inhibit nitrification and iron oxidation (Tab. 2; Figs. 4d, e; Jambert et al., 1994; Widdel et al., 1993). In groundwater, anoxic conditions enable heterotrophic denitrification, whereas an O₂ threshold of 30–60 umol L⁻¹ completely inhibits heterotrophic denitrification (Balestrini et al., 2016; Cey et al., 1999; Christensen et al., 2013; Jambert et al., 1994; Kolbjørn Jensen et al., 2017; Korom, 1992). In strictly forest sites, denitrification in groundwater is usually limited by the scarcity of NO₃, whereas in strictly crop sites denitrification is often limited by organic C availability (Tab. 2; Jambert et al., 1994; Starr and Gillham, 1993). Thus, N fertilizer application associated with different groundwater denitrification rates in the different plots creates the observed spatial pattern of groundwater NO₃ concentration in crop, riparian and forest sites (Tab. 2; Fig. 4c). In local maize croplands, Jambert et al (1997) found that 13% of the N fertilizers inputs were converted to N₂ gas, demonstrating that denitrification could occur in these oxic crop soils and groundwater. Although oxic conditions are not favorable for groundwater denitrification, some studies in agricultural catchments do describe this process at relatively high O₂ (150 µmol L⁻¹)

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levels (McAleer et al., 2017; Otero et al., 2009). In crop soils, Rubol et al. (2016) investigated the spatiotemporal dynamics in oxidative microbial activity and the development of anoxic micro zones (i.e., anoxic hot-spots) at the microscopic level (µm to cm). They found that labile C addition resulted in maximum rates of local metabolic activity within a few minutes and led to the subsequent formation of anoxic hotspots and thus, both oxic and anoxic conditions coexisted closely within a small volume of crop soils. Consequently, denitrification probably occurs in anoxic microsites in waterlogged soil during irrigation as higher soil moisture results in lower soil oxygen concentration, lower redox potential and higher leaching of soil DOC (Hagedorn et al., 2000; Jambert et al., 1997; Rubol et al., 2012; Silver et al., 1999). N fertilizer load in local croplands is 25 g N m⁻² yr⁻¹ (Jambert et al., 1997), whereas export (using drainage of 637 mm yr⁻¹ and the average NO₃ concentration in crop groundwater) of NO₃ through crop groundwater was estimated at 10 g N m⁻² yr⁻¹ (40% of the annual N fertilizer load), and export (using same drainage and the average NO₃ concentration in riparian groundwater) of NO₃ through riparian groundwater was estimated at 2.8 g N m⁻² yr⁻¹ (8% of the annual N fertilizer load). This shows the importance of riparian groundwater to attenuate N inputs from adjacent croplands to streams, otherwise a large portion of the annual N fertilizer load would have been leached into adjacent streams rather than being denitrified or used by plants. In riparian groundwater adjacent to a farm in the New York state (USA), Anderson et al. (2014) found that total groundwater denitrification was equivalent to 32% of manure N spread on the adjacent upland field. (Mekala et al., 2017) simulated the transport and dynamics of N in an agricultural soil under flooded conditions and concluded that relatively shallow aguifers with sandy soil are vulnerable to NO₃ contamination at around 10 days if continuous irrigation is practiced. They also stated that NO₃ had higher leaching potential than NH₄ or DOC. At our study site, irrigation and associated desorption of DOC and NO₃ could explain their slight increase in crop groundwater during base flow (Tab. 3, Figs. 1, S2c). In a storm infiltration basin in Florida (USA), O'Reilly et al. (2012) found that concomitant peaks in groundwater O₂ and NO₃ concentrations after storm rainfall were a consequence of organic N leaching, indicating that there were short periods of ammonification and

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nitrification. In crop groundwater of Wallonia (Belgium), when groundwater O_2 levels are higher than 125 μ mol L^{-1} (as at the study site), nitrification rather than denitrification promotes the accumulation of N_2O in groundwater (Jurado et al., 2017).

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4.4. Stream biogeochemical functioning: mostly a function of groundwater composition

NO₃ inputs to streams cause stream eutrophication (Carpenter et al., 1998; Jordan and Weller, 1996; Smith, 2003; Zhou et al., 2017). This is consistent with our field observations where we observed that crop streams were highly vegetated with macrophytes during base flow stages. Compared to high flow conditions, crop stream eutrophication was accompanied by higher pH and δ^{13} C-DIC, and lower pCO₂caused by preferential ¹²CO₂ uptake during the macrophyte plant photosynthesis (Tab. 4; Figs. S3a, h, j; De Carvalho et al., 2009; Raven et al., 2002). The development of macrophytes in crop streams modifies flow and can cause a significant drop in water velocity, which in turn, gives rise to extensive deposition and retention of sediment beneath the macrophytes (Cotton et al., 2006; Sand-Jensen and Pedersen, 1999). This leads to seasonal accumulation of organic matter, a predominance of anoxic conditions in stream sediments, and thus the occurrence of methanogenesis as evidenced by peaks in dissolved CH₄ during base flow (Tab. 4; Figs. S3g; Borges et al., 2018; Crawford et al., 2016; Sanders et al., 2007). Crop stream CH₄ concentration was 390 nmol L⁻¹ during base flow (Tab. 2), a concentration significantly lower (1,430 nmol L⁻¹) than chalk streams impacted by macrophyte vegetation in England (Sanders et al., 2007). This discrepancy probably resulted from the increased in silt and clay fraction during summer of the underlying sediment in chalk streams (Sanders et al., 2007). This would suggest that the permeability of chalk stream sediment became lower than that of sandy stream sediment and created stronger reducing conditions in chalk stream sediments, which likely increased the potential for methanogenesis (Baker et al., 1999; Findlay, 1995; Kankaala et al., 2005; Morrice et al., 1997). Sanders et al. (2007) also showed that the chalk streams' emissions of CH₄ to the atmosphere were approximately 50 times lower than the CH₄ production in stream sediments, illustrating the high potential for CH₄ oxidation

in the water column of crop stream. During base flow, a second explanation for higher CH₄ (and NH₄⁺) concentrations in crop streams relative to forest streams could be differential hydrology. Drainage plot is a function of the water table height (hydraulic gradient, Darcy's law). During base flow, the water table in cropland was deeper than in forest (e.g., 4 m deeper in Sep. 2014; Fig. 2), and so during this period, potentially more forest groundwater was drained into crop streams. However, in forest streams we usually did not observe CH₄ (or NH₄⁺) concentrations as high as in crop streams (Tab. 2; Fig. 7d) indicating that CH₄ (or NH₄⁺) in crop streams primarily originated from crop stream sediments rather than from higher discharge of forest groundwater. In crop streams, CH₄ was correlated with NH₄⁺ but not correlated with NO₃ or DOC (Figs. 6d, 7c, d). Such relationships were also observed in the Meuse river basin (Belgium) (Borges et al., 2018) and in a global meta-analysis of riverine CH₄ (Stanley et al., 2016). In contrast, this does not fit the conceptual model of Schade et al (2016) developed from data in New Hampshire streams, whereby the CH₄ was positively correlated with DOC, while negatively related to NO₃. Sandy sediments of low order stream beds impacted by eutrophication are significant areas of NO₃ reduction over the spring and summer, lowering DOC and NO₃ concentrations in stream water (Tab. 4; Figs. S3c, 1; Böhlke et al., 2009; Mulholland et al., 2008; Sanders et al., 2007). Additionally, the decreased of stream velocity during base flow increased residence times of NO₃ in the hyporheic zone and the time for denitrification (Bardini et al., 2012). In a small stream dominated by maize cropland in the USA, Böhlke et al. (2009) demonstrated that denitrification mainly occurred in sediments and not in the water column since integrated rates of pore-water denitrification derived from ¹⁵N tracer profiles within the hyporheic zone were similar to the reach-scale rates derived from measurements in the stream. In crop streams, a portion of the NO₃ variability between the two hydrological periods could also result from higher drainage of forest groundwater during base flow, which would dilute the NO₃ signal from crop groundwater. Considering the flat catchment topography, a minor portion of TSM and POC in streams originates from soil erosion and surface runoff. The most frequent effects of dredging on aquatic ecosystems are changes

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in the concentration of suspended solids, turbidity and light penetration (Lewis et al., 2001; Newell et al., 1998). Higher concentrations of POC (and TSM) observed in crop streams were also caused by macrophyte biomass developed in summer became a sediment trap. When stream discharge was sufficiently energetic, it re-suspended all the accumulated sediment and removed this litter. Moreover, we observed peaks of CH₄ and NH₄⁺ in crop streams during high flow (Tab. 4; Figs. S3d, g), suggesting that dredging or streambed erosion of crop streams also release CH₄ and NH₄⁺ from the sediment. In forest streams, we observed significantly lower concentrations of Fe²⁺, NH₄⁺, and CH₄ than in forest groundwater and significant negative linear relationships between O₂ on the one side and Fe²⁺, NH₄⁺, or CH₄ (Tab. 2; Figs. 4d, e, g, 7e, f). This suggests there were low O₂ concentration groundwater inputs with high concentrations of reduced compounds and that the stream water was gradually oxygenated, which induced Fe²⁺ and CH₄ oxidations and nitrification. Mulholland et al. (2000) studied N cycling by adding ¹⁵N-labeled NH₄⁺ into a forest stream in eastern Tennessee (USA). They concluded that the residence time of NH₄⁺ in the water column was low (5 min) and that nitrification was an important sink for NH₄⁺, accounting for 19% of total ammonium uptake. In forest streams, the NH₄⁺ concentration was approximately 3 µmol L⁻¹ lower than in forest groundwater and thus did not explained the NO₃ increase of 50 µmol L⁻¹ (Tab. 2; Figs. 3c, d). Up to 76% of N exports from local forest are in organic forms but these N exports are very low (< 0.2 g N m⁻² yr⁻¹; De Wit et al., 2005; Rimmelin, 1998; Vernier et al., 2003), so in-stream mineralization of organic N coupled to nitrification could not explain NO₃ concentrations in forest streams. Since the sampled forest streams are not strictly forested, NO₃ concentration are explained by simple hydrological mixing between crop and forest groundwater (Tab. 2). In streams, pCO₂ was lower and O₂ was higher than in groundwater (Tab. 2; Figs. 3f, h, 5a). This shows that gas exchange between stream water and the atmosphere occurs quickly, favored by low stream depth 614 and strong concentration gradients between the two compartments. Some authors (e.g., Bodmer et al. 2016; Borges et al., 2018) found elevated pCO₂ in crop streams rather than in forest streams, due to higher levels of dissolved and particulate organic matter in crop dominated systems compared to the forested

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ones that facilitated the in-stream degradation of organic matter. Moreover, land uses are expected to change the composition of terrestrial soil organic matter leached to streams, shifting from vegetation- to microbe-derived organic matter with greater agricultural land use and potentially higher emissions in crop streams (Fuss et al., 2017; Graeber et al., 2015; Wilson and Xenopoulos, 2009). Those results contrasted with ours because we found no difference in pCO₂ between crop and forest streams. Forest groundwater did have higher pCO₂ than crop groundwater, indicating a more intense degassing in forest streams. The similar δ^{13} C-DIC signatures in forest and crop streams despite the strong difference between crop and forest groundwater suggests faster isotopic equilibration of DIC resulting from degassing. The greater gas transfer velocity in forest streams is a consequence of the abundance of coarse woody debris which generates higher levels of water turbulence (e.g., Bodmer et al. 2016), and is consistent with our field observations. A lower gas transfer velocity lower in crop streams results from stream calibration reducing turbulent flow, and macrophyte vegetation that protects the water surface from wind shear.

5. Conclusion

The present study demonstrates that C and IN concentrations in shallow groundwater and in first-order streams are strongly sensitive to land use. In sandy lowland catchments, simultaneous measurements of biogeochemical parameters in groundwater and streams are crucial for identifying and quantifying biogeochemical processes involved at the groundwater-stream interface. We also show that a statistical clustering analysis based on NO₃⁻ dataset enables partitioning of groundwater and streams into cropaffected or forest-dominated waters. Such a classification could be useful to river managers and policy makers. The water table had greater depth in croplands and was a crucial parameter necessary for understanding groundwater biogeochemical variability in relation to land use. Higher water table stages in forests created anoxic conditions and increased soil leaching. Conversely, in croplands, the deeper water table prevented anoxic conditions, creating different groundwater compositions from forest groundwater and inhibiting the denitrification of the N fertilizers, which led to groundwater NO₃⁻ accumulation. Despite

642 the occurrence of groundwater denitrification in riparian and forest sites, N fertilizers inputs in crop 643 streams were still high enough to generate eutrophic conditions in these streams. Eutrophication resulted 644 in a biogeochemical cascading effect, which sustained high CH₄ concentration and lowered NO₃. High 645 CO₂ and CH₄ production occurs in forest soils and groundwater, but these two gases exhibit lower 646 concentrations in forest streams, indicating intense degassing or oxidation. 647 The groundwater-stream interface is a biogeochemical hotspot and hot moment for C emissions and N 648 removal processes (McClain et al., 2003). Future studies focusing on the groundwater-stream interface in 649 relation to land use are needed to better understand C and N dynamics in aquatic systems in order to 650 correctly close C and N budgets at regional and global scales. 651 652 Acknowledgments This research is part of the CNP-Leyre project funded by the Cluster of Excellence COTE at the 653 654 Université de Bordeaux (ANR-10-LABX-45). 655 656 **Bibliography** Abril, G., Borges, A.V., 2018. Carbon leaks from flooded land: do we need to re-plumb the inland water 657 658 active pipe? Biogeosciences Discuss. 2018, 1-46. https://doi.org/10.5194/bg-2018-239 659 Abril, G., Bouillon, S., Darchambeau, F., Teodoru, C.R., Marwick, T.R., Tamooh, F., Ochieng Omengo, F., Geeraert, N., Deirmendjian, L., Polsenaere, P., Borges, A.V., 2015. Technical Note: Large 660 661 overestimation of pCO2 calculated from pH and alkalinity in acidic, organic-rich freshwaters. Biogeosciences 12, 67-78. https://doi.org/10.5194/bg-12-67-2015 662 Abril, G., Martinez, J.-M., Artigas, L.F., Moreira-Turcq, P., Benedetti, M.F., Vidal, L., Meziane, T., Kim, J.-H., 663 Bernardes, M.C., Savoye, N., Deborde, J., Souza, E.L., Albéric, P., Landim de Souza, M.F., Roland, 664 665 F., 2014. Amazon River carbon dioxide outgassing fuelled by wetlands. Nature 505, 395–398. 666 https://doi.org/10.1038/nature12797 667 Aitkenhead, J.A., Hope, D., Billett, M.F., 1999. The relationship between dissolved organic carbon in stream water and soil organic carbon pools at different spatial scales. Hydrol. Process. 13, 1289-668

Anderson, L., 1979. Simultaneous spectrophotometric determination of nitrite and nitrate by flow

injection analysis. Anal. Chim. Acta 110, 123-128.

669

670

671

1302.

- Anderson, T.R., Groffman, P.M., Kaushal, S.S., Walter, M.T., 2014. Shallow groundwater denitrification in riparian zones of a headwater agricultural landscape. J. Environ. Qual. 43, 732–744.
- Asner, G.P., Elmore, A.J., Olander, L.P., Martin, R.E., Harris, A.T., 2004. Grazing systems, ecosystem responses, and global change. Annu Rev Env. Resour 29, 261–299.

- Augusto, L., Bakker, M.R., Morel, C., Meredieu, C., Trichet, P., Badeau, V., Arrouays, D., Plassard, C.,
 Achat, D.L., Gallet-Budynek, A., Merzeau, D., Canteloup, D., Najar, M., Ranger, J., 2010. Is 'grey
 literature' a reliable source of data to characterize soils at the scale of a region? A case study in a
 maritime pine forest in southwestern France. Eur. J. Soil Sci. 61, 807–822.
 https://doi.org/10.1111/j.1365-2389.2010.01286.x
 - Baker, M.A., Dahm, C.N., Valett, H.M., 1999. Acetate retention and metabolism in the hyporheic zone of a mountain stream. Limnol. Oceanogr. 44, 1530–1539.
 - Bakker, M.R., Augusto, L., Achat, D.L., 2006. Fine root distribution of trees and understory in mature stands of maritime pine (Pinus pinaster) on dry and humid sites. Plant Soil 286, 37–51.
 - Bakker, M.R., Jolicoeur, E., Trichet, P., Augusto, L., Plassard, C., Guinberteau, J., Loustau, D., 2009. Adaptation of fine roots to annual fertilization and irrigation in a 13-year-old Pinus pinaster stand. Tree Physiol. 29, 229–238.
 - Balestrini, R., Sacchi, E., Tidili, D., Delconte, C.A., Buffagni, A., 2016. Factors affecting agricultural nitrogen removal in riparian strips: examples from groundwater-dependent ecosystems of the Po Valley (Northern Italy). Agric. Ecosyst. Environ. 221, 132–144.
 - Barnes, R.T., Raymond, P.A., 2010. Land-use controls on sources and processing of nitrate in small watersheds: insights from dual isotopic analysis. Ecol. Appl. 20, 1961–1978.
 - Barnes, R.T., Raymond, P.A., 2009. The contribution of agricultural and urban activities to inorganic carbon fluxes within temperate watersheds. Chem. Geol. 266, 318–327.
 - Bass, A.M., Munksgaard, N.C., Leblanc, M., Tweed, S., Bird, M.I., 2014. Contrasting carbon export dynamics of human impacted and pristine tropical catchments in response to a short-lived discharge event. Hydrol. Process. 28, 1835–1843.
 - Bastviken, D., Tranvik, L.J., Downing, J.A., Crill, P.M., Enrich-Prast, A., 2011. Freshwater methane emissions offset the continental carbon sink. Science 331, 50–50.
 - Bell, R.A., Darling, W.G., Ward, R.S., Basava-Reddi, L., Halwa, L., Manamsa, K., Dochartaigh, B.Ó., 2017. A baseline survey of dissolved methane in aquifers of Great Britain. Sci. Total Environ. 601, 1803–1813.
 - Bernot, M.J., Sobota, D.J., Hall, R.O., Mulholland, P.J., Dodds, W.K., Webster, J.R., Tank, J.L., Ashkenas, L.R., Cooper, L.W., Dahm, C.N., 2010. Inter-regional comparison of land-use effects on stream metabolism. Freshw. Biol. 55, 1874–1890.
 - Bertran, P., Allenet, G., Gé, T., Naughton, F., Poirier, P., Goñi, M.F.S., 2009. Coversand and Pleistocene palaeosols in the Landes region, southwestern France. J. Quat. Sci. 24, 259–269.
 - Bertran, P., Bateman, M.D., Hernandez, M., Mercier, N., Millet, D., Sitzia, L., Tastet, J.-P., 2011. Inland aeolian deposits of south-west France: facies, stratigraphy and chronology. J. Quat. Sci. 26, 374–388
- Bodmer, P., Heinz, M., Pusch, M., Singer, G., Premke, K., 2016. Carbon dynamics and their link to
 dissolved organic matter quality across contrasting stream ecosystems. Sci. Total Environ. 553,
 574–586.
- Böhlke, J.K., Antweiler, R.C., Harvey, J.W., Laursen, A.E., Smith, L.K., Smith, R.L., Voytek, M.A., 2009.
 Multi-scale measurements and modeling of denitrification in streams with varying flow and
 nitrate concentration in the upper Mississippi River basin, USA. Biogeochemistry 93, 117–141.
- Borges, A.V., Darchambeau, F., Lambert, T., Bouillon, S., Morana, C., Brouyère, S., Hakoun, V., Jurado, A.,
 Tseng, H.-C., Descy, J.-P., 2018. Effects of agricultural land use on fluvial carbon dioxide, methane

- and nitrous oxide concentrations in a large European river, the Meuse (Belgium). Sci. Total Environ. 610, 342–355.
- Borges, A.V., Darchambeau, F., Teodoru, C.R., Marwick, T.R., Tamooh, F., Geeraert, N., Omengo, F.O.,
 Guérin, F., Lambert, T., Morana, C., Okuku, E., Bouillon, S., 2015. Globally significant greenhouse gas emissions from African inland waters. Nat. Geosci. 8, 637–642.
 https://doi.org/10.1038/ngeo2486
- Camino-Serrano, M., Gielen, B., Luyssaert, S., Ciais, P., Vicca, S., Guenet, B., Vos, B.D., Cools, N., Ahrens,
 B., Altaf Arain, M., Borken, W., Clarke, N., Clarkson, B., Cummins, T., Don, A., Graf Pannatier, E.,
 Laudon, H., Moore, T., Nieminen, T.., Nilsson, M.B., Peichi, M., Schwendenmann, L., Siemens, J.,
 Janssens, I.A., 2014. Linking variability in soil solution dissolved organic carbon to climate, soil
 type, and vegetation type. Glob. Biogeochem. Cycles 28, 497–509.
 https://doi.org/10.1002/2013GB004726.
- 731 Canton, M., Anschutz, P., Coynel, A., Polsenaere, P., Auby, I., Poirier, D., 2012. Nutrient export to an 732 Eastern Atlantic coastal zone: first modeling and nitrogen mass balance. Biogeochemistry 107, 733 361–377.

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741

742

743

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746

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748

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750

751

752

753

754

755

- Carpenter, S.R., Caraco, N.F., Correll, D.L., Howarth, R.W., Sharpley, A.N., Smith, V.H., 1998. Nonpoint pollution of surface waters with phosphorus and nitrogen. Ecol. Appl. 8, 559–568.
- Cey, E.E., Rudolph, D.L., Aravena, R., Parkin, G., 1999. Role of the riparian zone in controlling the distribution and fate of agricultural nitrogen near a small stream in southern Ontario. J. Contam. Hydrol. 37, 45–67.
- Christensen, J.R., Nash, M.S., Neale, A., 2013. Identifying riparian buffer effects on stream nitrogen in southeastern coastal plain watersheds. Environ. Manage. 52, 1161–1176.
- Christensen, T.R., Ekberg, A., Ström, L., Mastepanov, M., Panikov, N., Öquist, M., Svensson, B.H., Nykänen, H., Martikainen, P.J., Oskarsson, H., 2003. Factors controlling large scale variations in methane emissions from wetlands. Geophys. Res. Lett. 30.
- Ciais, P., Wattenbach, M., Vuichard, N., Smith, P., Piao, S.L., Don, A., Luyssaert, S., Janssens, I.A., Bondeau, A., Dechow, R., others, 2010. The European carbon balance. Part 2: croplands. Glob. Change Biol. 16, 1409–1428.
- Clague, J.C., Stenger, R., Clough, T.J., 2015. Denitrification in the shallow groundwater system of a lowland catchment: a laboratory study. Catena 131, 109–118.
 - Clark, I., Fritz, P., 1997. Environmental isotopes in hydrology. Lewis Publishers, Boca Raton, Fla.
- Cole, J.J., Prairie, Y.T., Caraco, N.F., McDowell, W.H., Tranvik, L.J., Striegl, R.G., Duarte, C.M., Kortelainen, P., Downing, J.A., Middelburg, J.J., Melack, J., 2007. Plumbing the Global Carbon Cycle: Integrating Inland Waters into the Terrestrial Carbon Budget. Ecosystems 10, 171–184. https://doi.org/10.1007/s10021-006-9013-8
 - Corbier, P., Karnay, G., Bourgine, B., Saltel, M., 2010. Gestion des eaux souterraines en région Aquitaine Reconnaissance des potentialités aquifères du Mio-Plio-Quaternaire des kandes de Gascogne et du Médoc en relation avec les SAGE, Module 7 (No. 57813). BRGM, Orléans, France.
- Cotton, J.A., Wharton, G., Bass, J.A.B., Heppell, C.M., Wotton, R.S., 2006. The effects of seasonal changes
 to in-stream vegetation cover on patterns of flow and accumulation of sediment.
 Geomorphology 77, 320–334.
- Crawford, J.T., Loken, L.C., Stanley, E.H., Stets, E.G., Dornblaser, M.M., Striegl, R.G., 2016. Basin scale
 controls on CO2 and CH4 emissions from the Upper Mississippi River. Geophys. Res. Lett. 43,
 1973–1979.
- Das, A., Krishnaswami, S., Bhattacharya, S.K., 2005. Carbon isotope ratio of dissolved inorganic carbon (DIC) in rivers draining the Deccan Traps, India: sources of DIC and their magnitudes. Earth Planet. Sci. Lett. 236, 419–429.

Datry, T., Malard, F., Gibert, J., 2004. Dynamics of solutes and dissolved oxygen in shallow urban groundwater below a stormwater infiltration basin. Sci. Total Environ. 329, 215–229.

- De Carvalho, M.C., Hayashizaki, K.-I., Ogawa, H., 2009. SHORT-TERM MEASUREMENT OF CARBON STABLE
 ISOTOPE DISCRIMINATION IN PHOTOSYNTHESIS AND RESPIRATION BY AQUATIC MACROPHYTES,
 WITH MARINE MACROALGAL EXAMPLES 1. J. Phycol. 45, 761–770.
 - De Wit, R., Leibreich, J., Vernier, F., Delmas, F., Beuffe, H., Maison, P., Chossat, J.-C., Laplace-Treyture, C., Laplana, R., Clave, V., 2005. Relationship between land-use in the agro-forestry system of les Landes, nitrogen loading to and risk of macro-algal blooming in the Bassin d'Arcachon coastal lagoon (SW France). Estuar. Coast. Shelf Sci. 62, 453–465.
 - Deirmendjian, L., Abril, G., 2018. Carbon dioxide degassing at the groundwater-stream-atmosphere interface: isotopic equilibration and hydrological mass balance in a sandy watershed. J. Hydrol. 558, 129–143. https://doi.org/10.1016/j.jhydrol.2018.01.003
 - Deirmendjian, L., Loustau, D., Augusto, L., Lafont, S., Chipeaux, C., Poirier, D., Abril, G., 2018. Hydroecological controls on dissolved carbon dynamics in groundwater and export to streams in a temperate pine forest. Biogeosciences 15, 669–691. https://doi.org/10.5194/bg-15-669-2018
 - Downing, J.A., Cole, J.J., Duarte, C.M., Middelburg, J.J., Melack, J.M., Prairie, Y.T., Kortelainen, P., Striegl, R.G., McDowell, W.H., Tranvik, L.J., 2012. Global abundance and size distribution of streams and rivers. Inland Waters 2, 229–236.
 - EEA, 2014. Corine Land Cover 2006 raster data. Eur. Environ. Agency EEA Available Httpwww Eea Eur. Eudata--Mapsdatadsresolveuida645109f7a11d43f5d7e275d81f35c61 3.
 - Etcheber, H., Taillez, A., Abril, G., Garnier, J., Servais, P., Moatar, F., Commarieu, M.-V., 2007. Particulate organic carbon in the estuarine turbidity maxima of the Gironde, Loire and Seine estuaries: origin and lability. Hydrobiologia 588, 245–259.
 - Evans, S.D., Lindstrom, M.J., Voorhees, W.B., Moncrief, J.F., Nelson, G.A., 1996. Effect of subsoiling and subsequent tillage on soil bulk density, soil moisture, and corn yield. Soil Tillage Res. 38, 35–46.
 - Findlay, S., 1995. Importance of surface-subsurface exchange in stream ecosystems: The hyporheic zone. Limnol. Oceanogr. 40, 159–164.
 - Findlay, S., Quinn, J.M., Hickey, C.W., Burrell, G., Downes, M., 2001. Effects of land use and riparian flowpath on delivery of dissolved organic carbon to streams. Limnol. Oceanogr. 46, 345–355.
 - Foley, J.A., DeFries, R., Asner, G.P., Barford, C., Bonan, G., Carpenter, S.R., Chapin, F.S., Coe, M.T., Daily, G.C., Gibbs, H.K., 2005. Global consequences of land use. science 309, 570–574.
 - Foulquier, A., Malard, F., Mermillod-Blondin, F., Datry, T., Simon, L., Montuelle, B., Gibert, J., 2010. Vertical change in dissolved organic carbon and oxygen at the water table region of an aquifer recharged with stormwater: biological uptake or mixing? Biogeochemistry 99, 31–47.
 - Frankignoulle, M., Borges, A.V., 2001. Direct and Indirect pCO2 Measurements in a Wide Range of pCO2 and Salinity Values (The Scheldt Estuary). Aquat. Geochem. 7, 267–273. https://doi.org/10.1023/A:1015251010481
 - Fuss, T., Behounek, B., Ulseth, A.J., Singer, G.A., 2017. Land use controls stream ecosystem metabolism by shifting dissolved organic matter and nutrient regimes. Freshw. Biol. 62, 582–599.
 - Gillikin, D.P., Bouillon, S., 2007. Determination of δ 18O of water and δ 13C of dissolved inorganic carbon using a simple modification of an elemental analyser-isotope ratio mass spectrometer: an evaluation. Rapid Commun. Mass Spectrom. 21, 1475–1478.
- 808 Gleick, P.H., 2003. Water use. Annu. Rev. Environ. Resour. 28, 275–314.
- Goldscheider, N., Hunkeler, D., Rossi, P., 2006. Microbial biocenoses in pristine aquifers and an assessment of investigative methods. Hydrogeol. J. 14, 926–941.
- Govind, A., Bonnefond, J.-M., Kumari, J., Moisy, C., Loustau, D., Wigneron, J.-P., 2012. Modeling the ecohydrological processes in the Landes de Gascogne, SW France, in: Plant Growth Modeling,

- Simulation, Visualization and Applications (PMA), 2012 IEEE Fourth International Symposium On. IEEE, pp. 133–140.
- Graeber, D., Boëchat, I.G., Encina-Montoya, F., Esse, C., Gelbrecht, J., Goyenola, G., Gücker, B., Heinz, M.,
 Kronvang, B., Meerhoff, M., 2015. Global effects of agriculture on fluvial dissolved organic
 matter. Sci. Rep. 5.
- Gran, G., 1952. Determination of the equivalence point in potentiometric titrations of seawater with hydrochloric acid. Ocean. Acta 5, 209–218.

821

822

828

829

830

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832

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834

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836

837

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839

840

841

842

843

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845

846

847

848

849

850

851

- Hagedorn, F., Kaiser, K., Feyen, H., Schleppi, P., 2000. Effects of redox conditions and flow processes on the mobility of dissolved organic carbon and nitrogen in a forest soil. J. Environ. Qual. 29, 288–297.
- Hagerdon, F., Schleppi, P., Waldner, P., Fluhler, H., 2000. Export of dissolved organic carbon and nitrogen from Gleysol dominated catchments—the significance of water flow paths. Biogeochemistry 50, 137–161.
- 826 Harwood, J.E., Kühn, A.L., 1970. A colorimetric method for ammonia in natural waters. Water Res. 4, 827 805–811. https://doi.org/10.1016/0043-1354(70)90037-0
 - Hiscock, K.M., Lloyd, J.W., Lerner, D.N., 1991. Review of natural and artificial denitrification of groundwater. Water Res. 25, 1099–1111.
 - Hotchkiss, E.R., Hall Jr, R.O., Sponseller, R.A., Butman, D., Klaminder, J., Laudon, H., Rosvall, M., Karlsson, J., 2015. Sources of and processes controlling CO2 emissions change with the size of streams and rivers. Nat. Geosci. 8, 696–699.
 - Hu, Y., Lu, Y., Edmonds, J.W., Liu, C., Wang, S., Das, O., Liu, J., Zheng, C., 2016. Hydrological and land use control of watershed exports of dissolved organic matter in a large arid river basin in northwestern China. J. Geophys. Res. Biogeosciences 121, 466–478.
 - Jackson, R.B., Carpenter, S.R., Dahm, C.N., McKnight, D.M., Naiman, R.J., Postel, S.L., Running, S.W., 2001. Water in a changing world. Ecol. Appl. 11, 1027–1045.
 - Jambert, C., Delmas, R.A., Labroue, L., Chassin, P., 1994. Nitrogen compound emissions from fertilized soils in a maize field pine tree forest agrosystem in the southwest of France. J. Geophys. Res. Atmospheres 99, 16523–16530.
 - Jambert, C., Serca, D., Delmas, R., 1997. Quantification of N-losses as NH 3, NO, and N 2 O and N 2 from fertilized maize fields in southwestern France. Nutr. Cycl. Agroecosystems 48, 91–104.
 - Jeong, C.H., 2001. Effect of land use and urbanization on hydrochemistry and contamination of groundwater from Taejon area, Korea. J. Hydrol. 253, 194–210.
 - Johnson, M.S., Lehmann, J., Couto, E.G., Novaes Filho, J.P., Riha, S.J., 2006. DOC and DIC in flowpaths of Amazonian headwater catchments with hydrologically contrasting soils. Biogeochemistry 81, 45–57.
 - Johnson, M.S., Lehmann, J., Riha, S.J., Krusche, A.V., Richey, J.E., Ometto, J.P.H., Couto, E.G., 2008. CO2 efflux from Amazonian headwater streams represents a significant fate for deep soil respiration. Geophys. Res. Lett. 35, L17401. https://doi.org/10.1029/2008GL034619.
 - Jolivet, C., Arrouays, D., Andreux, F., Lévèque, J., 1997. Soil organic carbon dynamics in cleared temperate forest spodosols converted to maize cropping. Plant Soil 191, 225–231.
- Jolivet, C., Arrouays, D., Leveque, J., Andreux, F., Chenu, C., 2003. Organic carbon dynamics in soil particle-size separates of sandy Spodosols when forest is cleared for maize cropping. Eur. J. Soil Sci. 54, 257–268.
- Jolivet, C., Augusto, L., Trichet, P., Arrouays, D., 2007. Forest soils in the Gascony Landes Region: formation, history, properties and spatial variability. Rev. For. Fr. 59, 7–30. https://doi.org/10.4267/2042/8480
- Jones, J.B., Mulholland, P.J., 1998. Carbon dioxide variation in a hardwood forest stream: an integrative measure of whole catchment soil respiration. Ecosystems 1, 183–196.

- Jordan, T.E., Weller, D.E., 1996. Human contributions to terrestrial nitrogen flux. BioScience 46, 655–664.
- Jurado, A., Borges, A., Pujades, E., Hakoun, V., Knöller, K., Brouyère, S., 2017. Occurrence of greenhouse gases (CO2, N2O and CH4) in groundwater of the Walloon Region (Belgium).
- Juste, C., Tauzin, J., Dureau, P., Courpron, C., 1982. Exportation des éléments fertilisants par lessivage en sol sableux des Landes de Gascogne. Résultats de 8 années d'observations en cases lysimétriques. Agronomie 2, 91–98.
 - Kankaala, P., Käki, T., Mäkelä, S., Ojala, A., Pajunen, H., Arvola, L., 2005. Methane efflux in relation to plant biomass and sediment characteristics in stands of three common emergent macrophytes in boreal mesoeutrophic lakes. Glob. Change Biol. 11, 145–153.
 - Kassambra, A., Mundt, F., 2017. factoextra: Extract and Visualize the Results of Multivariate Data Analyses. R package version 1.0.5.

- Klüber, H.D., Conrad, R., 1998. Effects of nitrate, nitrite, NO and N2O on methanogenesis and other redox processes in anoxic rice field soil. FEMS Microbiol. Ecol. 25, 301–318.
- Kokic, J., Wallin, M.B., Chmiel, H.E., Denfeld, B.A., Sobek, S., 2015. Carbon dioxide evasion from headwater systems strongly contributes to the total export of carbon from a small boreal lake catchment. J. Geophys. Res. Biogeosciences 120, 13–28. https://doi.org/10.1002/2014JG002706
- Kolbjørn Jensen, J., Engesgaard, P., Johnsen, A.R., Marti, V., Nilsson, B., 2017. Hydrological mediated denitrification in groundwater below a seasonal flooded restored riparian zone. Water Resour. Res. 53, 2074–2094.
- Korom, S.F., 1992. Natural denitrification in the saturated zone: a review. Water Resour. Res. 28, 1657–881 1668.
 - Lamba, J., Thompson, A.M., Karthikeyan, Kg., Fitzpatrick, F.A., 2015. Sources of fine sediment stored in agricultural lowland streams, Midwest, USA. Geomorphology 236, 44–53.
 - Lauerwald, R., Laruelle, G.G., Hartmann, J., Ciais, P., Regnier, P.A., 2015. Spatial patterns in CO2 evasion from the global river network. Glob. Biogeochem. Cycles 29, 534–554.
 - Lê, S., Josse, J., Husson, F., 2008. FactoMineR: an R package for multivariate analysis. J. Stat. Softw. 25, 1–18.
 - Legigan, P., 1979. L'élaboration de la formation du sable des Landes, dépôt résiduel de l'environnement sédimentaire pliocène-pléïstocène centre aquitain (Thèse de Doctorat d'Etat n°642). Université de Bordeaux I, Bordeaux.
 - Lehrter, J.C., 2006. Effects of land use and land cover, stream discharge, and interannual climate on the magnitude and timing of nitrogen, phosphorus, and organic carbon concentrations in three coastal plain watersheds. Water Environ. Res. 78, 2356–2368.
 - Lewis, E., Wallace, D., Allison, L.J., 1998. Program developed for CO2 system calculations. Carbon Dioxide Information Analysis Center, managed by Lockheed Martin Energy Research Corporation for the US Department of Energy Tennessee, Oak Ridge, Tennessee.
 - Lewis, M.A., Weber, D.E., Stanley, R.S., Moore, J.C., 2001. Dredging impact on an urbanized Florida bayou: effects on benthos and algal-periphyton. Environ. Pollut. 115, 161–171.
 - Ludwig, W., Amiotte-Suchet, P., Probst, J.-L., 1996a. River discharges of carbon to the world's oceans: determining local inputs of alkalinity and of dissolved and particulate organic carbon. Sci. Terre Planètes Comptes Rendus Académie Sci. 323, 1007–1014.
 - Ludwig, W., Probst, J.-L., Kempe, S., 1996b. Predicting the oceanic input of organic carbon by continental erosion. Glob. Biogeochem. Cycles 10, 23–41.
- 904 Lundström, U.S., van Breemen, N., Bain, D., 2000. The podzolization process. A review. Geoderma 94,
 905 91–107.
- 906 MacQueen, J., 1967. Some methods for classification and analysis of multivariate observations, in: 907 Proceedings of the Fifth Berkeley Symposium on Mathematical Statistics and Probability. 908 Oakland, CA, USA, pp. 281–297.

- 909 Malard, F., Hervant, F., 1999. Oxygen supply and the adaptations of animals in groundwater. Freshw. 910 Biol. 41, 1–30.
- 911 Marx, A., Dusek, J., Jankovec, J., Sanda, M., Vogel, T., Geldern, R., Hartmann, J., Barth, J.A.C., 2017. A 912 review of CO2 and associated carbon dynamics in headwater streams: a global perspective. Rev. 913 Geophys.
- 914 Masese, F.O., Salcedo-Borda, J.S., Gettel, G.M., Irvine, K., McClain, M.E., 2017. Influence of catchment
 915 land use and seasonality on dissolved organic matter composition and ecosystem metabolism in
 916 headwater streams of a Kenyan river. Biogeochemistry 132, 1–22.
- 917 McAleer, E.B., Coxon, C.E., Richards, K.G., Jahangir, M.M.R., Grant, J., Mellander, P.E., 2017. 918 Groundwater nitrate reduction versus dissolved gas production: a tale of two catchments. Sci. 919 Total Environ. 586, 372–389.
- 920 McClain, M.E., Boyer, E.W., Dent, C.L., Gergel, S.E., Grimm, N.B., Groffman, P.M., Hart, S.C., Harvey, J.W., 921 Johnston, C.A., Mayorga, E., McDowell, W.H., Pinay, G., 2003. Biogeochemical hot spots and hot 922 moments at the interface of terrestrial and aquatic ecosystems. Ecosystems 6, 301–312.
- 923 McMahon, P.B., Chapelle, F.H., 2008. Redox processes and water quality of selected principal aquifer 924 systems. Groundwater 46, 259–271.
- 925 Mekala, C., Gaonkar, O., Nambi, I.M., 2017. Understanding nitrogen and carbon biogeotransformations 926 and transport dynamics in saturated soil columns. Geoderma 285, 185–194.
- 927 Meybeck, M., 1987. Global chemical weathering of surficial rocks estimated from river dissolved loads. 928 Am. J. Sci. 401–428.
- Meybeck, M., 1982. Carbon, nitrogen, and phosphorus transport by world rivers. Am J Sci 282, 401–450.
- 930 Millero, F.J., 1979. The thermodynamics of the carbonate system in seawater. Geochim. Cosmochim. 931 Acta 43, 1651–1661.
- 932 Molofsky, L.J., Connor, J.A., McHugh, T.E., Richardson, S.D., Woroszylo, C., Alvarez, P.J., 2016. 933 Environmental factors associated with natural methane occurrence in the Appalachian Basin. 934 Groundwater 54, 656–668.
- 935 Montgomery, D.R., 2007. Soil erosion and agricultural sustainability. Proc. Natl. Acad. Sci. 104, 13268– 936 13272.
- 937 Moore, T.R., Knowles, R., 1989. The influence of water table levels on methane and carbon dioxide 938 emissions from peatland soils. Can. J. Soil Sci. 69, 33–38.

940

941

942

943

944

- Moreaux, V., Lamaud, É., Bosc, A., Bonnefond, J.-M., Medlyn, B.E., Loustau, D., 2011. Paired comparison of water, energy and carbon exchanges over two young maritime pine stands (Pinus pinaster Ait.): effects of thinning and weeding in the early stage of tree growth. Tree Physiol. 903–921. https://doi.org/10.1093/treephys/tpr048
- Morrice, J.A., Valett, H.M., Dahm, C.N., Campana, M.E., 1997. Alluvial characteristics, groundwater—surface water exchange and hydrological retention in headwater streams. Hydrol. Process. 11, 253–267.
- 946 Mulholland, P.J., Helton, A.M., Poole, G.C., Hall, R.O., Hamilton, S.K., Peterson, B.J., Tank, J.L., Ashkenas, 947 L.R., Cooper, L.W., Dahm, C.N., 2008. Stream denitrification across biomes and its response to 948 anthropogenic nitrate loading. Nature 452, 202.
- 949 Mulholland, P.J., Tank, J.L., Sanzone, D.M., Wollheim, W.M., Peterson, B.J., Webster, J.R., Meyer, J.L., 950 2000. Nitrogen cycling in a forest stream determined by a 15N tracer addition. Ecol. Monogr. 70, 951 471–493.
- Naumburg, E., Mata-Gonzalez, R., Hunter, R.G., Mclendon, T., Martin, D.W., 2005. Phreatophytic vegetation and groundwater fluctuations: a review of current research and application of ecosystem response modeling with an emphasis on Great Basin vegetation. Environ. Manage. 35, 726–740.

- Newell, R.C., Seiderer, L.J., Hitchcock, D.R., 1998. The impact of dredging works in coastal waters: a review of the sensitivity to disturbance and subsequent recovery of biological resources on the sea bed. Oceanogr. Mar. Biol. Annu. Rev. 36, 127–178.
- 959 O'Leary, M.H., 1988. Carbon isotopes in photosynthesis. Bioscience 328–336.

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991

- 960 Onderka, M., Pekarova, P., Miklanek, P., Halmova, D., Pekar, J., 2010. Examination of the dissolved 961 inorganic nitrogen budget in three experimental microbasins with contrasting land cover—a 962 mass balance approach. Water. Air. Soil Pollut. 210, 221–230.
 - O'Reilly, A.M., Chang, N.-B., Wanielista, M.P., 2012. Cyclic biogeochemical processes and nitrogen fate beneath a subtropical stormwater infiltration basin. J. Contam. Hydrol. 133, 53–75.
 - Otero, N., Torrentó, C., Soler, A., Menció, A., Mas-Pla, J., 2009. Monitoring groundwater nitrate attenuation in a regional system coupling hydrogeology with multi-isotopic methods: the case of Plana de Vic (Osona, Spain). Agric. Ecosyst. Environ. 133, 103–113.
 - Pabich, W.J., Valiela, I., Hemond, H.F., 2001. Relationship between DOC concentration and vadose zone thickness and depth below water table in groundwater of Cape Cod, USA. Biogeochemistry 55, 247–268.
- Polsenaere, P., Abril, G., 2012. Modelling CO2 degassing from small acidic rivers using water pCO2, DIC
 and δ13C-DIC data. Geochim. Cosmochim. Acta 91, 220–239.
 https://doi.org/10.1016/j.gca.2012.05.030
 - Polsenaere, P., Savoye, N., Etcheber, H., Canton, M., Poirier, D., Bouillon, S., Abril, G., 2013. Export and degassing of terrestrial carbon through watercourses draining a temperate podzolized catchment. Aquat. Sci. 75, 299–319.
 - Postel, S., 1999. Pillar of sand: can the irrigation miracle last? WW Norton & Company.
 - Quénéa, K., Derenne, S., Largeau, C., Rumpel, C., Mariotti, A., 2006. Influence of change in land use on the refractory organic macromolecular fraction of a sandy spodosol (Landes de Gascogne, France). Geoderma 136, 136–151.
 - Quinton, J.N., Govers, G., Van Oost, K., Bardgett, R.D., 2010. The impact of agricultural soil erosion on biogeochemical cycling. Nat. Geosci. 3, 311–314.
 - R Core Team, 2018. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
 - Ramankutty, N., Foley, J.A., 1999. Estimating historical changes in global land cover: Croplands from 1700 to 1992. Glob. Biogeochem. Cycles 13, 997–1027.
 - Ramos, T.B., Rodrigues, S., Branco, M.A., Prazeres, A., Brito, D., Gonçalves, M.C., Martins, J.C., Fernandes, M.L., Pires, F.P., 2015. Temporal variability of soil organic carbon transport in the Enxoé agricultural watershed. Environ. Earth Sci. 73, 6663–6676.
 - Raven, J.A., Johnston, A.M., Kübler, J.E., Korb, R., McInroy, S.G., Handley, L.L., Scrimgeour, C.M., Walker, D.I., Beardall, J., Vanderklift, M., 2002. Mechanistic interpretation of carbon isotope discrimination by marine macroalgae and seagrasses. Funct. Plant Biol. 29, 355–378.
- Ravishankara, A.R., Daniel, J.S., Portmann, R.W., 2009. Nitrous oxide (N2O): the dominant ozonedepleting substance emitted in the 21st century. science 326, 123–125.
- 995 Raymond, P.A., Cole, J.J., 2003. Increase in the export of alkalinity from North America's largest river. 996 Science 301, 88–91.
- 997 Raymond, P.A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover, M., Butman, D., Striegl, R.,
 998 Mayorga, E., Humborg, C., Kortelainen, P., Dürr, H., Meybeck, M., Ciais, P., Guth, P., 2013. Global
 999 carbon dioxide emissions from inland waters. Nature 503, 355–359.
 1000 https://doi.org/10.1038/nature12760
- Rimmelin, P., 1998. Etude des apports allochtones d'azote inorganique dissous parvenant à un système lagunaire: le Bassin d'Arcachon.

- Robinson, M., Ryder, E.L., Ward, R.C., 1985. Influence on streamflow of field drainage in a small agricultural catchment. Agric. Water Manag. 10, 145–158.
- 1005 Rodrigues, V., Estrany, J., Ranzini, M., de Cicco, V., Martín-Benito, J.M.T., Hedo, J., Lucas-Borja, M.E.,
 1006 2018. Effects of land use and seasonality on stream water quality in a small tropical catchment:
 1007 The headwater of Córrego Água Limpa, São Paulo (Brazil). Sci. Total Environ. 622, 1553–1561.
- Rosegrant, M.W., Cai, X., Cline, S.A., 2002. World water and food to 2025: dealing with scarcity. Intl Food Policy Res Inst.
- Rubol, S., Dutta, T., Rocchini, D., 2016. 2D visualization captures the local heterogeneity of oxidative metabolism across soils from diverse land-use. Sci. Total Environ. 572, 713–723.
- Rubol, S., Silver, W.L., Bellin, A., 2012. Hydrologic control on redox and nitrogen dynamics in a peatland soil. Sci. Total Environ. 432, 37–46.
- Salomons, W., Mook, W.G., 1986. Isotope geochemistry of carbonates in the weathering zone. Handb. Environ. Isot. Geochem. 2, 239–269.
 - Salvia-Castellví, M., Iffly, J.F., Vander Borght, P., Hoffmann, L., 2005. Dissolved and particulate nutrient export from rural catchments: a case study from Luxembourg. Sci. Total Environ. 344, 51–65.
 - Sanders, I.A., Heppell, C.M., Cotton, J.A., Wharton, G., Hildrew, A.G., Flowers, E.J., Trimmer, M., 2007. Emission of methane from chalk streams has potential implications for agricultural practices. Freshw. Biol. 52, 1176–1186.
- Sand-Jensen, K., Pedersen, O., 1999. Velocity gradients and turbulence around macrophyte stands in streams. Freshw. Biol. 42, 315–328.
- Schade, J.D., Bailio, J., McDowell, W.H., 2016. Greenhouse gas flux from headwater streams in New Hampshire, USA: Patterns and drivers. Limnol. Oceanogr. 61.
- 1025 Sharp, J.H., 1993. The dissolved organic carbon controversy: an update. Oceanography 6.

1017

1018

1019

- Silver, W.L., Lugo, A.E., Keller, M., 1999. Soil oxygen availability and biogeochemistry along rainfall and topographic gradients in upland wet tropical forest soils. Biogeochemistry 44, 301–328.
- Smith, V.H., 2003. Eutrophication of freshwater and coastal marine ecosystems a global problem. Environ. Sci. Pollut. Res. 10, 126–139.
- Stanley, E.H., Casson, N.J., Christel, S.T., Crawford, J.T., Loken, L.C., Oliver, S.K., 2016. The ecology of methane in streams and rivers: patterns, controls, and global significance. Ecol. Monogr. 86, 146–171.
- Starr, R.C., Gillham, R.W., 1993. Denitrification and organic carbon availability in two aquifers.

 Groundwater 31, 934–947.
- Stella, P., Lamaud, E., Brunet, Y., Bonnefond, J.-M., Loustau, D., Irvine, M., 2009. Simultaneous measurements of CO 2 and water exchanges over three agroecosystems in South-West France. Biogeosciences 6, 2957–2971.
- 1038 Stookey, L.L., 1970. Ferrozine—a new spectrophotometric reagent for iron. Anal. Chem. 42, 779–781.
- Stott, T., 2005. Natural recovery from accelerated forest ditch and stream bank erosion five years after
 harvesting of plantation forest on Plynlimon, mid-Wales. Earth Surf. Process. Landf. J. Br.
 Geomorphol. Res. Group 30, 349–357.
- Thivolle-Cazat, A., Najar, M., 2001. Évolution de la productivité et de la récolte du pin maritime dans le massif Landais. Evaluation de la disponibilité future en Gironde. Rev. For. Fr. 53, 351–355.
- Trichet, P., Bakker, M.R., Augusto, L., Alazard, P., Merzeau, D., Saur, E., 2009. Fifty years of fertilization experiments on Pinus pinaster in Southwest France: the importance of phosphorus as a fertilizer. For. Sci. 55, 390–402.
- Tsypin, M., Macpherson, G.L., 2012. The effect of precipitation events on inorganic carbon in soil and shallow groundwater, Konza Prairie LTER Site, NE Kansas, USA. Appl. Geochem. 27, 2356–2369.

- 1049 Ulrich, E., Coddeville, P., Lanier, M., 2002. Retombées atmosphériques humides en France entre 1993 et
 1050 1998:[données et références-coordination technique de la surveillance de la qualité de l'air].
 1051 Ademe.
- Vernier, F., Beuffe, H., Chossat, J.-C., 2003. Forêt et ressource en eau: étude de deux bassins versants en sol sableux (Landes de Gascogne). Rev. For. Fr. 55, 523–542.
 - Vernier, F., Castro, A., 2010. Critère Préservation de l'environnement Sous-critère Eau (Rapport d'expertise: Critère "Préservation de l'environnement"). GIP-ECOFOR, Bordeaux, France.

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1057

10631064

1065

1083

1084

- Vidon, P., Wagner, L.E., Soyeux, E., 2008. Changes in the character of DOC in streams during storms in two Midwestern watersheds with contrasting land uses. Biogeochemistry 88, 257–270.
- Vogel, J.C., Ehleringer, J.R., Hall, A.E., Farquhar, G.D., 1993. Variability of carbon isotope fractionation
 during photosynthesis., in: Stable Isotopes and Plant Carbon-Water Relations. Academic Press
 Inc., pp. 29–46.
- Wachniew, P., 2006. Isotopic composition of dissolved inorganic carbon in a large polluted river: The Vistula, Poland. Chem. Geol. 233, 293–308.
 - Wallin, M.B., Grabs, T., Buffam, I., Laudon, H., Ågren, A., Öquist, M.G., Bishop, K., 2013. Evasion of CO2 from streams The dominant component of the carbon export through the aquatic conduit in a boreal landscape. Glob. Change Biol. 19, 785–797. https://doi.org/10.1111/gcb.12083
- Weiss, R., 1974. Carbon dioxide in water and seawater: the solubility of a non-ideal gas. Mar. Chem. 2, 203–215.
- Widdel, F., Schnell, S., Heising, S., Ehrenreich, A., Assmus, B., Schink, B., 1993. Ferrous iron oxidation by anoxygenic phototrophic bacteria. Nature 362, 834.
- Wilson, H.F., Xenopoulos, M.A., 2009. Effects of agricultural land use on the composition of fluvial dissolved organic matter. Nat. Geosci. 2, 37–41.
- Wynn, T.M., Mostaghimi, S., 2006. Effects of riparian vegetation on stream bank subaerial processes in
 southwestern Virginia, USA. Earth Surf. Process. Landf. J. Br. Geomorphol. Res. Group 31, 399–
 413.
- Young, R.G., Huryn, A.D., 1999. Effects of land use on stream metabolism and organic matter turnover. Ecol. Appl. 9, 1359–1376.
- Zhang, F., Wang, J., Wang, X., 2018. Recognizing the Relationship between Spatial Patterns in Water
 Quality and Land-Use/Cover Types: A Case Study of the Jinghe Oasis in Xinjiang, China. Water 10,
 646.
- Zhou, Y., Xu, J.F., Yin, W., Ai, L., Fang, N.F., Tan, W.F., Yan, F.L., Shi, Z.H., 2017. Hydrological and
 environmental controls of the stream nitrate concentration and flux in a small agricultural
 watershed. J. Hydrol. 545, 355–366.

Stream Order	Description	Catchment area (km²) ^a	Crop (%) ^b	Forest (%) ^b	Urban (%) ^b	During High Flow ^c	During Base Flow ^c
1	Ditch	1.0	86.5	13.5	0.0	C	С
1	Ditch	1.3	53.8	46.2	0.0	C	C
1	Ditch	11.3	44.2	55.8	0.0	C	C
1	Ditch	13.4	42.5	57.5	0.0	C	C
1	Stream	57.0	30.7	69.3	0.0	C	C
1	Stream	16.8	7.8	92.2	0.0	F	F
1	Ditch	7.9	5.8	94.2	0.0	F	F
1	Ditch	2.3	5.2	94.8	0.0	F	C
1	Stream	16.0	4.6	93.8	1.6	C	F
1	Stream	34.0	3.8	96.2	0.0	F	F
1	Stream	31.0	2.3	97.7	0.0	F	F
1	Headwater	0.3	0.0	100.0	0.0	F	F
0	Groundwater in a riparian forest but very near (5m) a maize cropland (P1)					R	R
0	Groundwater in maize cropland (P2)					C	С
0	Groundwater in maize cropland (P3)					C	C
0	Groundwater in pine forest (P4)					F	F
0	Groundwater in pine forest (P5)					F	F

Table 1. Characteristics of groundwater and stream sampling stations, ranked in decreasing order of cropland percentage in their respective subcatchments. ^a delimited with a geographic information system software (ArcGIS 10.5®) using an hydrological database in a polyline form (BD CARTHAGE®) and a digital elevation model (BD ALTI®, resolution of 25m), which both have been made available by the national geographic institute of France (http://www.ign.fr/). ^b retrieved with the CORINE land cover 2006 database (EEA, 2014) using a geographic information system software (ArcGIS 10.5®). ^c C, F, R corresponding to crop, forest and riparian waters, respectively, either during high or base flow. Piezometer 1 (P1) is located in a riparian mixed pine and oak forest near a first-order stream and near a maize cropland, which where P2 is located. P2 and P3 are located in the middle of two different maize croplands of 0.6 km² and 6 km², respectively. P5 is located in an 11-years old pine plot of 0.6 km² and is part of the ICOS (name is FR-Bil) research infrastructure (http://icos-ri.eu), whereas P4 is located in another pine forest (approximately same age as P5 pine forest). The depth of piezometers (from the soil surface to the bottom of the piezometer) is 5.3m for P1, 4.9m for P2, 9.1m for P3, 5m for P4 and P5.

	Crop continuum		Forest cont	Riparian groundwater	
	Groundwaters (22)	Streams (59)	Groundwaters (22)	Streams (78)	Groundwaters (11)
рН	4.5±0.2	6.0±0.3	4.5±0.3	5.8±0.5	4.7±0.1
	4.3-5.0	5.4-7.0	3.7-4.8	4.2-6.9	4.4-4.8
Temperature (°C)	14.5±1.8	13.6 ± 4.2	12.8±1.8	12.9±3.9	14.9±2.4
•	10.7-17.5	6.4-25.8	8.5-15.1	4.8-22.1	11.8-17.9
EC (µS cm ⁻¹)	360±70	220±55	90±10	115±30	160±50
•	220-465	75-370	65-115	70-200	95-270
NO_3^- (µmol L^{-1})	$1,140\pm485$	340±200	25±40	75±70	310±260
	260-1,785	10-950	0-120	0-275	40-860
NH_4^+ (µmol L^{-1})	0.4 ± 0.8	6.1±7.0	4.5±7.0	1.8±1.7	0.4 ± 0.4
,	0-3.5	0-40	0.3-30	0-7.8	0-1.5
Fe^{2+} (µmol L ⁻¹)	0.9 ± 0.4	5.9 ± 4.4	15±15	7.9 ± 12.0	0.6 ± 0.5
•	0.1-1.9	0.1-22	0.9 ± 56	0.6 ± 58	0.2-1.5
O_2 (µmol L ⁻¹)	220±65	290+45	20±30	280±50	100±70
- ,	100-315	160-400	0-110	110-370	0-170
CH_4 (nmol L^{-1})	40±25	460±950	$1,770\pm1,830$	240±300	1,470±1,490
. ,	15-130	20-4,900	50-6,700	20-2,370	30-4,150
pCO ₂ (ppmv)	$30,650\pm11,590$	$4,480\pm2,680$	50,630±26,070	$4,900\pm4,500$	42,950±28,560
	19,000-60,550	1,040-14,080	7,680-116,380	1,000-27,200	17,300-103,300
$TA (\mu mol L^{-1})$	90±25	100±50	70±30	90±50	70±15
•	35-130	30-300	30-135	30-280	45-85
δ^{13} C-DIC (‰)	-19.8±1.3	-18.2±3.5	-26.7±1.0	-19.8 ± 2.8	-25.2±1.1
, ,	-22 to -17.6	-27.6 to -11.3	-28.8 to -24	-27.6 to -14	-27.9 to -23.4
DIC (µmol L ⁻¹)	$1,450\pm480$	315±135	$2,460\pm1,130$	320±210	1,960±1,150
,	820-2,590	90-650	570-5,370	120-1,280	940-4,480
DOC (µmol L ⁻¹)	510±150	605±320	930±930	470±250	400±100
, ,	275-880	220-2,290	310-3,670	190-1,725	280-620
$TSM (mg L^{-1})$		5.6±8.6		2.3±1.7	
\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \		0.1-50.5		0.4-8.2	
POC (%)		28±10		28±10	
` '		0-50		10-80	
POC (µmol L ⁻¹)		120±180		50±35	
4 /		0-1,100		0-170	

Tab. 2. Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in crop and forest continuums and in riparian groundwater. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed average±standard deviations and the range.

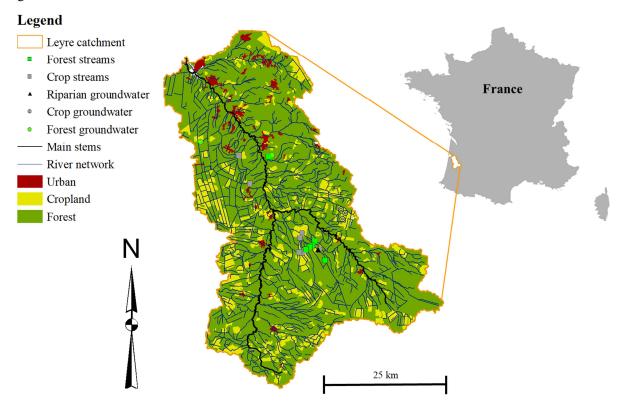
pH Temperature (°C) EC (μS cm ⁻¹)	Cropland_HF (4) 4.6±0.3 4.3-4.9 12.8±1.7 10.7-14.5	Cropland_BF (18) 4.5±0.2 4.3-5.0 14.9±1.6	Forest_HF (6) 4.4±0.3 4.0-4.8	Forest_BF (16) 4.5±0.3	Riparian_HF (2) 4.7±0.1	Riparian_BF (9)
Temperature (°C) EC (μS cm ⁻¹)	4.3-4.9 12.8±1.7	4.3-5.0		4.5 ± 0.3	47+01	1.6.0
EC (μS cm ⁻¹)	12.8±1.7		4.0-4.8		1.7 =0.1	4.6±0
EC (μS cm ⁻¹)		1/1 0+1 6		3.7-4.8	4.6-4.8	4.4-4.
	10.7-14.5	14.7.1.0	10.8 ± 1.4	13.5±1.4	12.2±0.6	15.6±2
		11.6-17.5	8.5-12.2	10.7-15.1	11.8-12.6	12.1-17
. 1	370±60	360±70	90±15	90±10	200±20	150±5
. 1	320-460	220-470	70-115	70-115	185-215	95-27
NO_3^- (µmol L ⁻¹)	$1,040\pm300$	$1,160\pm420$	30±50	20±40	510±20	260±27
	760-1,320	260-1785	0-120	0-120	500-520	40-86
NH_4^+ (µmol L ⁻¹)	0.5 ± 0.4	0.4 ± 0.9	3.3 ± 2.2	5.0 ± 8.0	0.3 ± 0.1	0.4 ± 0
•	0.1-1	0-3.5	1.1-7	0-3-30	0.2-0.3	0-1
Fe^{2+} (µmol L ⁻¹)	0.8 ± 0.2	0.9 ± 0.4	10.0 ± 8.2	15±15	0.4 ± 0.2	0.7 ± 0
·	0.7-1.1	0.1-1.9	2.7-25.5	0.9-56.6	0.2-0.5	0.2-1
$O_2 (\mu \text{mol } L^{-1})$	250±90	220±70	20±20	20 ± 30	170±0	100±8
•	180-310	100-320	0-40	0-110	170-170	0-20
CH ₄ (nmol L ⁻¹)	30±3	50±25	480±630	$2,260\pm1,900$	$1,460\pm2,010$	$1,470\pm1,50$
	25-30	16-130	50-1,700	50-6,700	40-2,880	30-4,13
pCO ₂ (ppmv)	$22,050\pm2,000$	32,560±12,000	$28,100\pm11,580$	$59,080\pm25,060$	21,530±5,950	47,700±29,59
	19,800-24,270	19,000-60,550	7,680-39,000	29,685-116,400	17,320-25,740	20,600-103,30
TA (μmol L ⁻¹)	85±2	92±30	95±40	65±30	83±2	60-
•	82-86	35-130	60-135	30-100	82-85	45-
δ^{13} C-DIC (‰)	-20.7±1.1	-19.6±1.3	-26.6±1.3	-26.8 ± 1.0	-26.9 ± 1.4	-24.9±0
,	-22 to -19.7	-21.9 to -17.6	-27.6 to -24.0	-28.8 to -25.3	-27.9 to -25.9	-25.7 to -23
DIC (µmol L ⁻¹)	$1,100\pm180$	1,520±490	1,500±550	$2,830\pm1,080$	1,160±315	$2,140\pm1,2$
4 /	930-1,300	820-2,590	570-2,040	1,650-5,370	940-1,380	1,020-4,4
DOC (µmol L ⁻¹)	420±120	550±140	2,230±1,440	740±380	310±50	420±1
4 /	320-590	340-880	575-3,670	310-1,720	275-350	310-6

Tab. 3. Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in different types of groundwater across hydrological seasons. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed the average±standard deviations and the range. We defined six groups that are Cropland_HF/Cropland_BF, Forest_HF/Forest_BF and Riparian_HF/Riparian_BF corresponding to groundwaters during high flow (HF) or base flow (BF); in either cropland, forest or riparian forest.

		First-order streams						
	Cropland_HF (22)	Cropland_BF (37)	Forest_HF (23)	Forest_BF (55)				
pH	5.9±0.3	6.1±0.4	5.7±0.6	6.1±0.4				
	5.4-6.6	5.5-7.0	4.2-6.8	5.0-6.9				
Temperature (°C)	10.2±1.6	15.7±3.9	9.0±1.9	14.6±3.3				
•	6.4-12.1	9.1-25.8	4.8-12	8.1-22.1				
EC (μS cm ⁻¹)	230±50	220±60	110±20	120±30				
•	145-340	75-370	80-150	70-200				
NO_3^- (µmol L^{-1})	420±220	290±170	95±70	65±70				
	180-950	8.5-705	0-275	0-275				
NH_4^+ (µmol L^{-1})	7.0 ± 8.4	5.5±6.0	1.7±1.7	1.7±1.7				
•	0.5-38.7	0-25.3	0.3-7.8	0-6.9				
Fe^{2+} (µmol L ⁻¹)	6.7 ± 3.8	5.4±4.7	5.7±3.0	8.8±14.0				
,	1.6-15.7	0.1-22	2.6-13.6	0.6-57.1				
O_2 (µmol L^{-1})	290±50	290±40	300±40	270±60				
•	190-400	160-370	210-370	110-360				
CH ₄ (nmol L ⁻¹)	580±1,080	390±880	185±190	270±340				
, ,	30-4,380	20-4,900	40-980	20-2,370				
pCO ₂ (ppmv)	5,200±2,370	$4,040\pm2,790$	4,200±2,430	5,200±5,100				
- 41	1,040-10,740	1,220-14,080	$1,240\pm11,690$	1,010-27,200				
TA (μmol L ⁻¹)	105±50	100±50	70±40	95±55				
,	40-300	30-255	35-195	30-280				
δ^{13} C-DIC (‰)	-20.6±3.9	-16.8±2.4	-22.1±2.5	-18.9±2.3				
` ,	-27.6 to -11.3	-22.3 to -12.4	-27.6 to -16.8	-23.1 to -14.0				
DIC (μmol L ⁻¹)	380±130	280±120	300±150	330±230				
,	1,000-650	90-600	150-750	120-1,280				
DOC (µmol L ⁻¹)	750±400	520±230	540±305	450±220				
4	300-2,290	220-1,520	260-1725	190-1540				
TSM (mg L ⁻¹)	9.3±11.5	3.1±4.9	2.8±1.7	2.1±1.7				
<i>\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ </i>	0.9-51	0.1-27	0.5-6.6	0.4-8.2				
POC (%)	26±10	30±10	29±8	29±10				
` '	15-48	16-48	20-50	12-80				
POC (µmol L ⁻¹)	190±250	65±100	65±40	40±30				
,	0-1100	0.3-540	0-170	0.5-140				

Tab. 4. Values of carbon and ancillary parameters throughout sampling period (Jan. 2014-Jul. 2015) in different types of streams across hydrological seasons. Numbers between brackets are corresponding to the sampling size. For each parameter, the table showed the average±standard deviations and the range. We defined four groups that are Cropland_HF/Cropland_BF, Forest_HF/Forest_BF corresponding to streams during high flow (HF) or base flow (BF); in either cropland-affected or forest-dominated land use.

Figure 1: Land use map of the Leyre catchment showing river network and the sampling locations of groundwaters and streams.



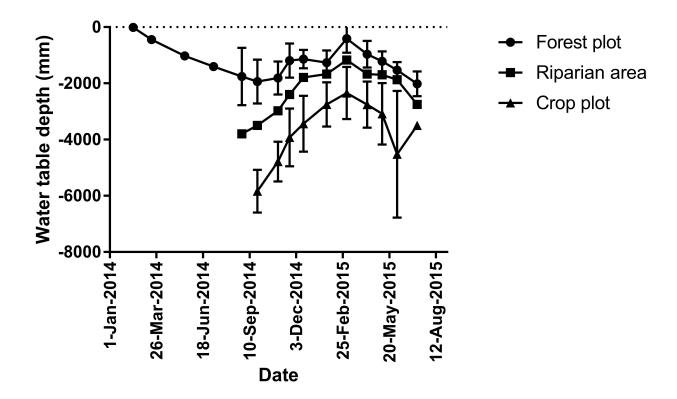


Figure 2: Water table depth during the sampling period (Jan. 2014-Jul. 2015) across land use in the Leyre catchment. The water table in riparian area is the water table at P1 (Tab. 1). The water table in crop plot is the average±standard deviations of water tables at P2 and P3 (Tab. 1). The water table in forest plot is the average±standard deviations of water tables at P4 and P5 (Tab. 1).

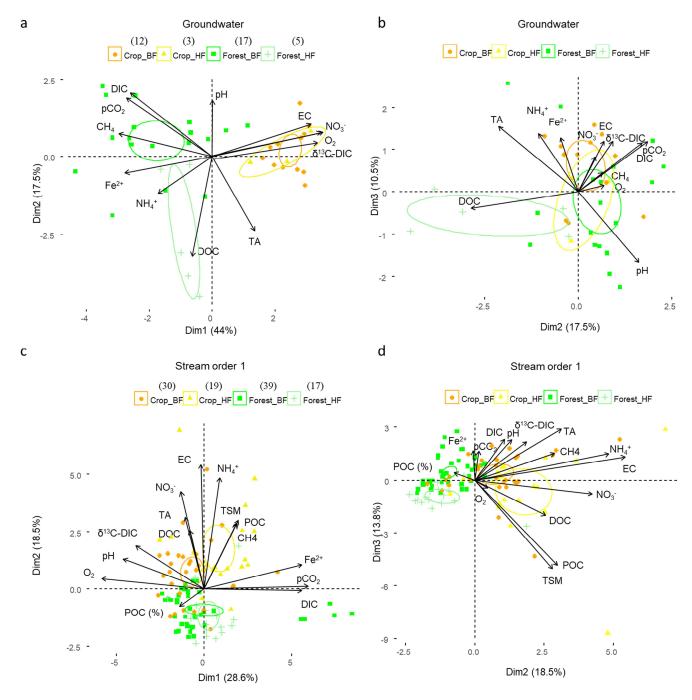


Figure 3: Principal component analysis (PCA) of shallow groundwater dataset (a-b) and first-order streams dataset (c-d). We represented only the first three dimensions. Numbers between brackets are corresponding to the sampling size. The sampling size in the PCAs did not correspond exactly to the sampling size in Tables 3 and 4 because R software deletes stations from the analysis with a missing value for one parameter. In these PCAs, we used all the quantitative variables measured in this study. In each PCA, we plotted as well the individuals separated in four groups. The first group corresponds to cropland-affected samples during high flow (Crop_HF), the second group corresponds to cropland-affected samples during base flow (Crop_BF), the third group corresponds to forest-dominated samples during base flow (Forest_BF).

The mean value of each qualitative group has 95% chance to be within the corresponding confidence ellipse.

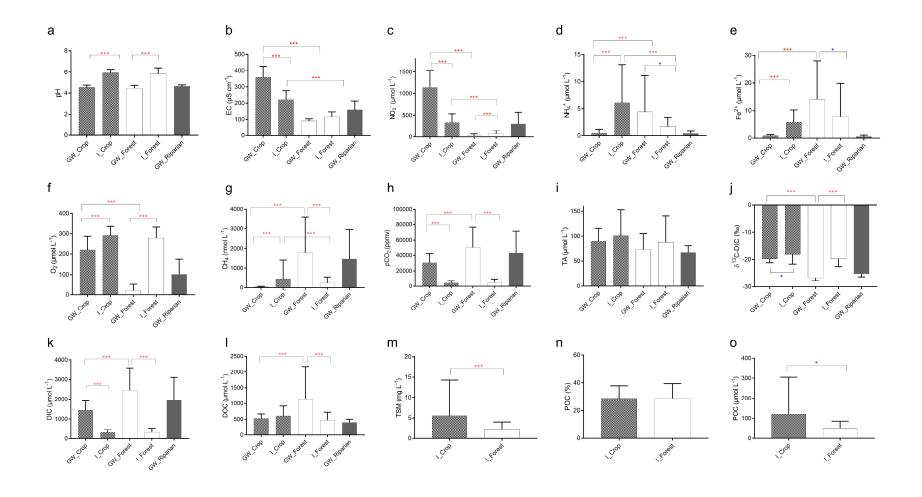


Figure 4: Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in groundwater and streams across land use. Histograms represent the mean with standard deviations of a given parameter. We defined four groups that are GW_Forest/GW_Crop and I_Forest/I_crop corresponding to groundwaters and streams order 1 either dominated by forests or croplands. A fifth group is GW_Riparian and corresponding to riparian groundwater. Then, based on Mann-Whitney statistical analysis, we compared GW_Crop VS I_Crop, GW_Forest VS I_Forest, GW_Crop VS GW_Forest, I_Crop VS I_Forest. Three red stars (***) indicate that data were significantly different with p<0.001. One blue star (*) indicates that data were significantly different with p<0.05).

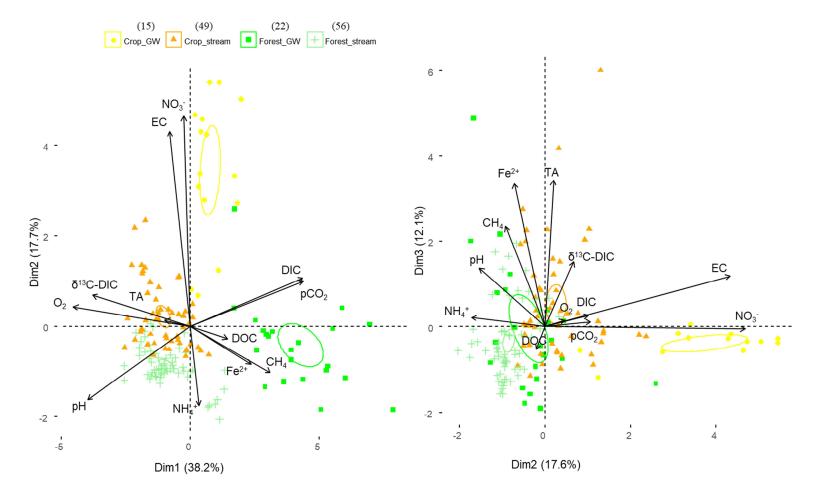


Figure 5: Principal component analysis (PCA) of shallow groundwater and stream datasets. We represented only the first three dimensions. Numbers between brackets are corresponding to the sampling size. The sampling size in the PCA did not correspond exactly to the sampling size in Table 2 because R software deletes stations from the analysis with a missing value for one parameter. In these multivariate statistical analyses, we used all the quantitative variables measured in this study. We defined four groups that are Crop_GW/Forest_GW and Crop_stream/Forest_stream, which are corresponding to groundwater and streams order 1, either dominated by forests or croplands. The mean value of each qualitative group has 95% chance to be within the corresponding confidence ellipse

Figure 6: Scatter plots of (a) CO_2 (ppmv) vs. O_2 (μ mol L^{-1}), (b) CO_2 (ppmv) vs. DOC (μ mol L^{-1}), (c) CH_4 (nmol L^{-1}) vs. O_2 (μ mol L^{-1}), (d) CH_4 vs. DOC (μ mol L^{-1}), (e) CH_4 (nmol L^{-1}) vs. CO_2 (ppmv), and (f) O_2 (μ mol L^{-1}) vs. DOC (μ mol L^{-1}), in all sampled groundwater and streams

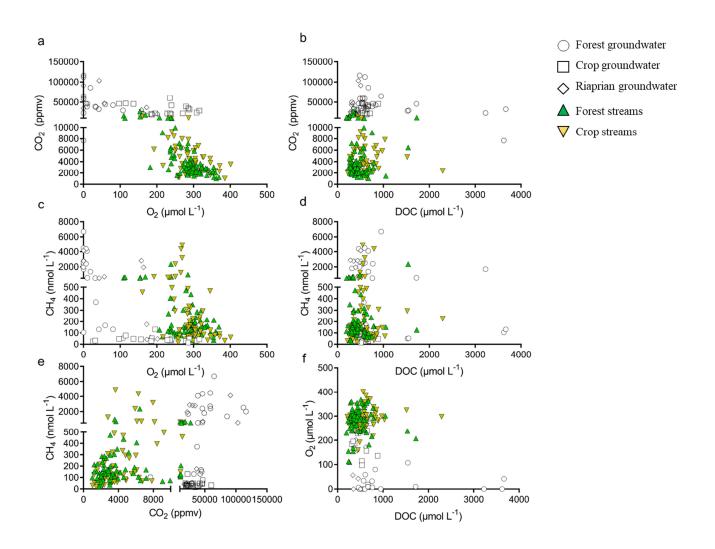
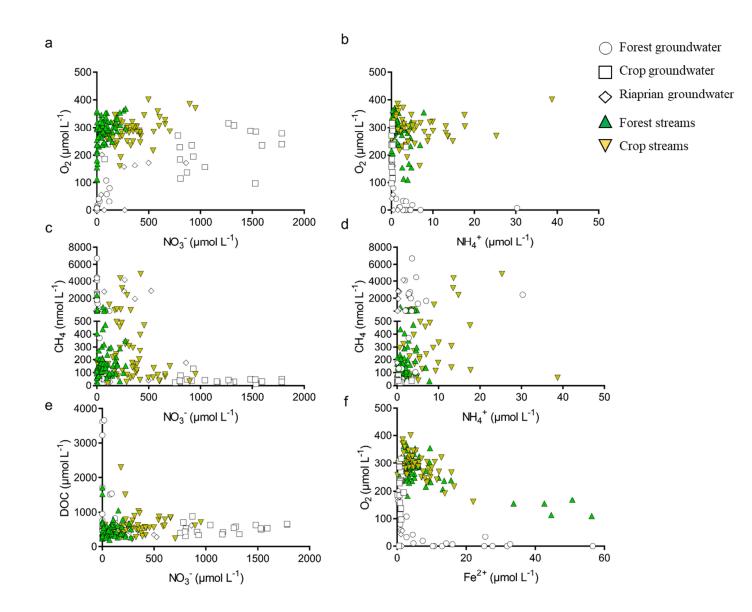


Figure 7: Scatter plots of (a) $O_2(\mu mol\ L^{-1})$ vs. $NO_3^-(\mu mol\ L^{-1})$, (b) $O_2(\mu mol\ L^{-1})$ vs. $NH_4^+(\mu mol\ L^{-1})$, (c) $CH_4(\mu mol\ L^{-1})$ vs. $NO_3^-(\mu mol\ L^{-1})$, (d) $CH_4(\mu mol\ L^{-1})$ vs. $DOC(\mu mol\ L^{-1})$, (e) $DOC(\mu mol\ L^{-1})$ vs. $NO_3^-(\mu mol\ L^{-1})$, and (f) $O_2(\mu mol\ L^{-1})$ vs. $Fe^{2+}(\mu mol\ L^{-1})$, in all sampled groundwater and streams.



Supplementary Material

Seasons	Date	Groundwaters			Streams
		P5	P1, P2, P3	P4	
HF	29/01/2014				X
HF	12/02/2014	X			
HF	07/03/2014				X
HF	17/03/2014	X			
BF	24/04/2014				X
BF	16/05/2014	X			
BF	21/05/2014				X
BF	25/06/2014				X
BF	17/072014	X			X
BF	27/08/2014	X	X		X
BF	24/09/2014	X	X		X
BF	31/10/2014	X	X		X
BF	21/11/2014	X	X		X
BF	16/12/2014	X	X		X
HF	27/01/2015	X	X	X	X
HF	04/03/2015	X	X	X	X
BF	10/04/2015	X	X	X	X
BF	07/05/2015	X	X	X	X
BF	03/06/2015	X	X	X	X
BF	09/07/2015	X	X	X	X

Table S1: Sampling dates of groundwaters and streams. X corresponds to a sampling. HF and BF are corresponding to high flow and base flow periods, respectively.

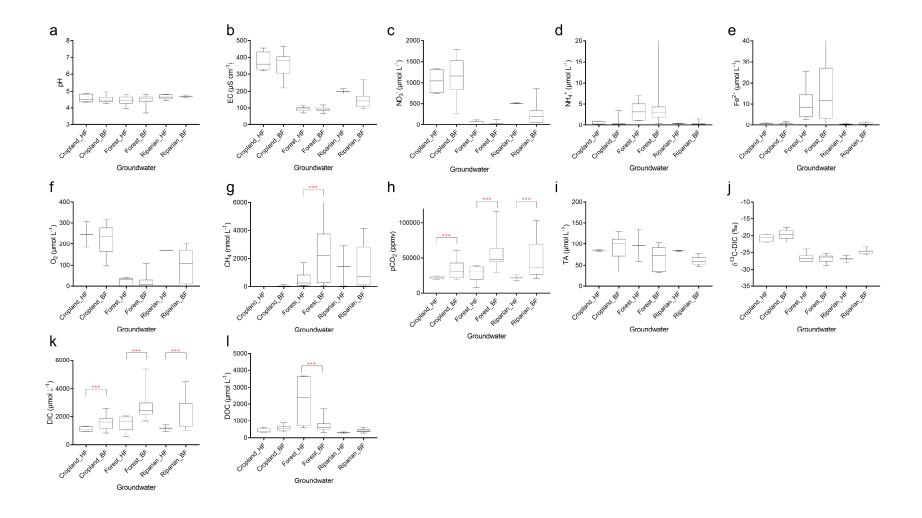


Fig. S2. Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in groundwaters across hydrological seasons and land use. Box-plots represent the median (black bar) and the extreme (min-max) values. We defined six groups that are Cropland_HF/Cropland_BF, Forest_HF/Forest_BF and Riparian_HF/Riparian_BF corresponding to groundwater during high or base flow; in either cropland, forest or riparian forest. Then, based on Mann-Whitney statistical analysis, we compared Cropland_HF vs. Cropland_BF,

Forest_HF vs. Forest_BF and Riparian_HF vs. Riparian_BF. Three red stars (***) indicate that data were significantly different with p<0.001. One blue star (*) indicates that data were significantly different with p<0.05. No stars indicate that data were not significantly different (p>0.05)

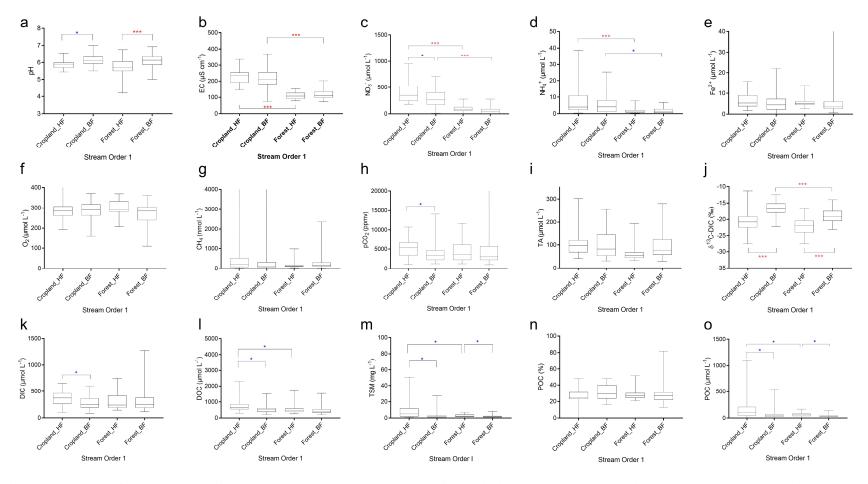


Figure S3. Values of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in streams across hydrological seasons and land use Box-plots represent the median (black bar) and the extreme (min-max) values. We defined four groups that are Cropland_HF/Cropland_BF, Forest_HF/Forest_BF corresponding to streams during high or base flow periods; in either cropland-affected or forest-dominated land use. Then, based on Mann-Whitney statistical analysis, we compared Cropland_HF vs. Cropland_BF, Forest_HF vs. Forest_BF, Cropland_BF vs. Forest_HF and Cropland_BF vs. Forest_BF. Three red stars (***) indicate that data were significantly different with p<0.001. One blue star (*) indicates that data were significantly different with p<0.05)

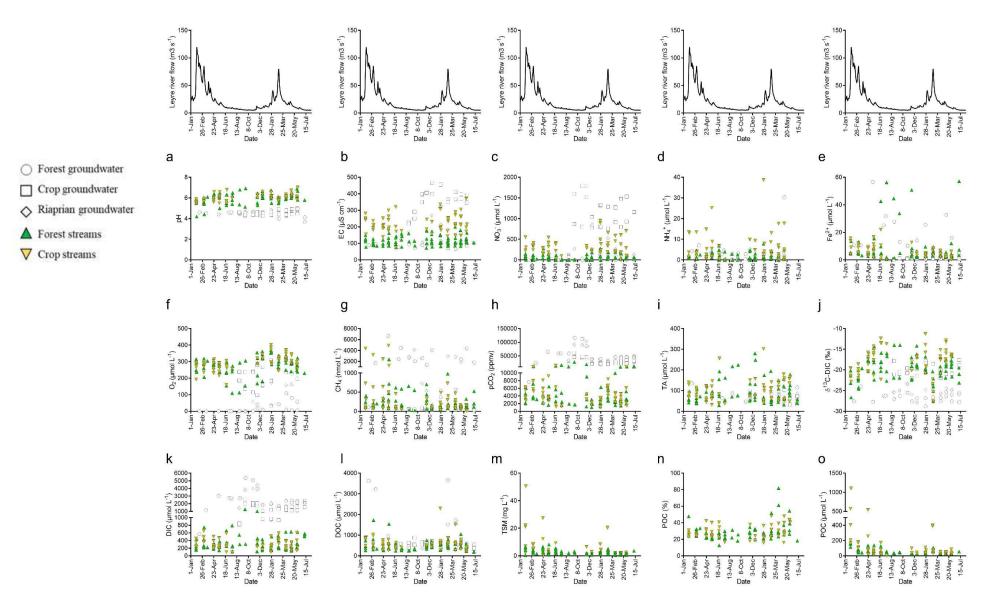


Figure S4: Temporal variations of carbon and ancillary parameters throughout the sampling period (Jan. 2014-Jul. 2015) in all sampled

groundwater and streams. Top panels represent the Leyre River flow (main stem).

