

# Lake sediment mercury biogeochemistry controlled by sulphate input from drainage basin

Axel Canredon, Pierre Anschutz, Damien Buquet, Celine Charbonnier, David Amouroux, Emmanuel Tessier, Dominique Poirier, Stéphane Bujan, Ludovic Devaux, Benoit Gouillieux, et al.

# ▶ To cite this version:

Axel Canredon, Pierre Anschutz, Damien Buquet, Celine Charbonnier, David Amouroux, et al.. Lake sediment mercury biogeochemistry controlled by sulphate input from drainage basin. Applied Geochemistry, 2019, 104, pp.135-145. 10.1016/j.apgeochem.2019.03.023 . hal-02282515

# HAL Id: hal-02282515 https://univ-pau.hal.science/hal-02282515

Submitted on 22 Oct 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial | 4.0 International License Version of Record: https://www.sciencedirect.com/science/article/pii/S0883292719300861 Manuscript\_3a818fc0f532d7c4e24ef03ba19153f5

- 1 Lake sediment mercury biogeochemistry controlled by sulphate input from drainage basin
- 2 Axel CANREDON<sup>1</sup>, Pierre ANSCHUTZ<sup>1</sup>\*, Damien BUQUET<sup>1</sup>, Céline CHARBONNIER<sup>1</sup>, David
- 3 AMOUROUX<sup>4</sup>, Emmanuel TESSIER<sup>4</sup>, Dominique POIRIER<sup>1</sup>, Stéphane BUJAN<sup>1</sup>, Ludovic
- 4 DEVAUX<sup>1,3</sup>, Benoît GOUILLIEUX<sup>2</sup>, Sophie GENTES<sup>2</sup>, Alexia LEGEAY<sup>2</sup>, Agnès FEURTET-
- 5 MAZEL<sup>2</sup>, Serge GALAUP<sup>5</sup>, Régine MAURY-BRACHET<sup>2</sup>.
- 6 (1) Université de Bordeaux CNRS, Environnements et Paléoenvironnements Océaniques et
- 7 Continentaux EPOC, UMR5805, 33615 Pessac, France
- 8 (2) Université de Bordeaux CNRS, Environnements et Paléoenvironnements Océaniques et
- 9 Continentaux EPOC, UMR5805, Station Marine d'Arcachon, 2 Rue du Professeur Jolyet, 33120
- 10 Arcachon, France
- (3) EPHE Dynamique des Environnements Naturels et Anthropisés (DENA), UMR5805, 33615
   Pessac, France
- 13 (4) CNRS Université de Pau & Pays Adour, Institut des Sciences Analytiques et de Physico-chimie
- 14 pour l'Environnement et les Matériaux IPREM, UMR5254, 64000, PAU, France.
- (5) ENSEGID Bordeaux INP, EA 4592 Géoressources & Environnement, 1 allée Daguin 33607
   Pessac, France
- 17 (\*) corresponding author, ORCID identifier is 0000-0001-5331-7974
- 18
- 19 Abstract

Mercury (Hg) and methyl mercury (MeHg) concentrations have never been measured in sediments of coastal lakes of Aquitaine, although high concentrations of Hg have been measured in fish. Our objective was to characterize benthic biogeochemical processes and the distribution of Hg in lake sediments and to connect these results with fish contamination. For this, we mapped and characterized sediments. We measured sediment Hg and MeHg content, and biogeochemical parameters. We identified organic deposits in deep areas, and sandy sediments in shallow areas. Sediments were anoxic below the sediment–water interface. The average Hg concentration in organic

sediment was 213 µg kg<sup>-1</sup> dry weight. Sandy sediments had an average Hg concentration of 4 µg kg<sup>-1</sup> 27 dw. We measured concentrations below 6 µg kg<sup>-1</sup> dw in sediments from streams that drain the 28 29 catchment. Similar concentrations in the four lakes suggest that the source of total Hg was not a point 30 source in a given lake. The highest MeHg concentrations were found in the upper centimetres of 31 organic sediments, where sulphate reduction occurred. MeHg represented 2.53% of total Hg for Lake 32 Carcans-Hourtin, less in other lakes. The proportion of MeHg in sediment followed lake water 33 sulphate concentrations. High sulphate concentrations resulted from agricultural activity in the Lake 34 Carcans-Hourtin catchment. Our results corroborate the hypothesis that Hg methylation is linked to 35 sulphate-reducing activity. High fluxes of sulphate from a drainage basin may induce large proportions of potentially bioavailable MeHg in lake sediment, even in non-polluted areas. The Hg methylation 36 37 activity in the sediment reflected the exposure of predatory fish to MeHg.

38

39 Keywords : Mercury methylation, fish contamination, aquatic sediment, sulphate, freshwater lake

40

# 41 *1. Introduction*

Technological, industrial, and agricultural progress triggered both the exploitation of natural resources and the dispersion of anthropogenic contaminants. Since the 19<sup>th</sup> century, mercury (Hg) fluxes in the environment have increased at a global scale. Typical concentrations of Hg range from 50 to 150  $\mu$ g kg<sup>-1</sup> in the continental crust (Wedepohl, 1995), from 0.1 to 2 ng L<sup>-1</sup> in continental and ocean waters (Mason et al., 1994), and from 1 to 3 ng m<sup>-3</sup> in the atmosphere (Schroeder and Munthe, 1998).

47 Mercury dynamics is characterizes by properties, such as volatility at ambient temperature, 48 stability of its bonds with carbon and sulphur, and very high bioconcentration and toxicity (Cossa and 49 Ficht, 1999). Methyl mercury (MeHg) is the most toxic chemical form of Hg. It is a neurotoxin for 50 vertebrates, which leads to damage in central nervous system (Allen et al., 2002). Methyl mercury 51 passes easily through biological barriers and can bioaccumulate and biomagnify along the food chain. 52 Although it is present at very low concentrations in water, it can concentrate up to 10 million times in 53 aquatic organisms such as predators (Cossa and Ficht, 1999). Human exposure to MeHg compounds occurs often through the consumption of fish and fish products (Driscoll et al., 2013). High trophic level species of fish can present very high MeHg concentrations. The maximum tolerable level for the sale of carnivorous fish is 1 mg kg<sup>-1</sup> wet weight according to the European Commission directive 93/351/EEC.

58 Mercury is naturally emitted in the environment through degassing of the Earth's crust, volcanoes, and erosion of soils and rocks. Human-related Hg emissions dominate the Hg cycle such 59 60 that most of the Hg in the atmosphere is anthropogenic (Mason et al., 1994). Natural Hg emission and re-emission mainly occurs as elemental mercury vapours (Hg<sup>0</sup>). The main mercury source in aquatic 61 62 environments comes from wet and dry atmospheric deposition, and via runoff from the drainage basin 63 (Mason and Fitzgerald, 1996). Globally, the main anthropogenic Hg emissions into the atmosphere 64 originate from artisanal gold production, the combustion of coal, and solid wastes (Pirrone et al., 2010; 65 UNEP, 2013; Streets et al. 2017). These emissions, coupled with long-distance transport of elemental Hg, result in a worldwide distribution of Hg deposition with a global average of 5.6  $\mu$ g m<sup>-2</sup> yr<sup>-1</sup> 66 67 (Lamborg et al., 2002).

68 In 2015, a large-scale sampling of piscivorous fish was conducted in order to study fish 69 contamination by mercury in the four main lakes in Aquitaine, specifically from the north to the south, 70 lakes Carcans-Hourtin, Lacanau, Cazaux-Sanguinet, and Parentis-Biscarrosse (Fig. 1). The results 71 showed an increasing Hg contamination gradient from Lake Parentis-Biscarrosse to Lake Carcans-72 Hourtin (south-north gradient), with values in the top predator zander (Sander lucioperca) from 2000 µg kg<sup>-1</sup> dw in the southernmost lake to 7400 µg kg<sup>-1</sup> dw in the Lake Carcans-Hourtin (Gentès et al., 73 submitted). This latter study showed that trophic transfer efficiency does not explain this Hg latitudinal 74 75 gradient in biota. The mercury concentration in fish suggested that the lakes underwent variable 76 contamination pressure. However, the four lakes are located in the same geological context, they are 77 subject to identical climate, and they do not have known local Hg sources, suggesting that different 78 fish contamination in each lake results from distinct methylation efficiency of a background Hg flux. 79 Therefore, these lakes represent a natural laboratory, in which the study of biogeochemical parameters 80 allows us to determine the impact of the main factors that control methylation of Hg. Methylation 81 occurs through the action of sulphate-reducing bacteria (Gilmour et al., 1992) and, to a lesser extent, 82 iron-reducing and methanogenic microorganisms (Alpers et al., 2013; Hamelin et al. 2011; Kerin et 83 al., 2006; Parks et al., 2013), which are common in organic-rich sediments. Consequently, methylation 84 is localized in the anoxic zone of aquatic environments such as biofilms, sediment, or water column. As Aquitaine lakes are shallow, between 7 and 23 m maximum depth, the water column is generally 85 well mixed and oxygenated. Therefore, it was expected that anoxic conditions favourable to Hg 86 87 methylation occurred in anoxic sediments instead of the water column. Indeed, it has been observed recently that the bottom of the deepest parts of Aquitaine lakes are blanketed with anoxic organic 88 89 muds (Buquet et al., 2017).

The present study focuses on the relationship between the biogeochemistry of lakes and the distribution of mercury and MeHg in sediments. The aim was to (i) perform a mapping of lake sediments in order to link Hg concentrations and the nature of the substrate, (ii) measure concentrations of total mercury and methyl mercury in several representative sediment samples, (iii) analyse the redox compounds influencing Hg speciation in sediments, (iv) attempt to explain the differences in sediment mercury speciation between the four lakes studied, and finally (v) determine the relationship of Hg in sediments and Hg in fish.

- 97
- 98
- 99 **2.** Materials and Methods

# 100 **2.1.Study site**

Large lakes in Aquitaine are located close to the French Atlantic coast in the south-western part of France. They are separated from the ocean by a 5-km-wide forested coastal dune system (Fig. 1), which represents a physical barrier to streams draining the lowland of Landes de Gascogne. Most of the drainage basin of the lakes is located on the eastern side. The surface areas of the catchments are similar, between 200 and 310 km<sup>2</sup> (Table 1), which is relatively small. The relief of the drainage basins is very flat with an westward slope lower than 0.2%. The geology is homogeneous and consists of podzols established on Quaternary siliceous aeolian deposits (Jolivet et al., 2007). Soil occupation
has been determined by remote sensing using maps of the French national institute of geography
(IGN), and aerial photographs analysed with the Geographic Information System (GIS) ArcGIS
(Canton et al., 2012). Pine (*Pinus pinaster*) forests dominate the land use and occupy more than 80%
of the total surface (Fig. 1). Intensive farming covers between 5% and 20% of lake drainage basins.
Housing is confined to the coastal zone.

113 Lakes are fed by direct rainfall on the lake surface and by low-discharge rivers. River systems 114 are artificial ditches dug to dry up marshes. Water level of lakes is controlled by locks in channels that 115 connect lakes. Locks also determine the water residence time (Table 1). Lake Lacanau is located 116 downstream from Lake Carcans-Hourtin. The lakes are connected by the 7.5-km-long Etang channel. 117 Waters of Lake Lacanau flow to the Arcachon Lagoon through the 18.5-km-long Porge channel. Lake 118 Parentis-Biscarrosse is downstream from Lake Cazaux-Sanguinet. All of these lakes are shallow (less 119 than 23 m) (Table 1). The deepest part is located on the western side of the lakes, at the base of the 120 dunes. Only the southern lakes are deep enough to have a marked summer thermocline at about 12 m 121 depth. The hypolimion does not become totally anoxic in the oligotrophic Lake Cazaux-Sanguinet. 122 Cyanobacteria blooms occur frequently in Lake Parentis-Biscarrosse due to eutrophication caused by 123 the legacy of large amounts of phosphorus stored in lake sediment. This lake experiences short periods 124 of bottom-water anoxia for a few weeks during summer water stratification. Less than 10% of the 125 water volume becomes anoxic during these episodes. Dense stands of invasive aquatic plants (Egeria 126 densa and Lagarosiphon major) are found in Lake Parentis-Biscarrosse and Lake Lacanau. They are 127 developed in shallow-sheltered or deep-exposed areas and represent about 10% of lake surface area 128 (Bertrin et al., 2017). Only restricted vegetated areas occurs in the other two lakes. The highest 129 biomass is present in Lake Parentis-Biscarrosse, with an estimated amount of 2800 tons (Bertrin et al., 130 2017).

# 131 **2.2.Sampling**

Fish were collected from the four lakes by fishing associations and federations and by aprofessional fisherman during autumn 2015. Three well-represented piscivorous species in the four

134 lakes were selected: zander (Sander lucioperca), pike (Esox Lucius) and perch (Perca fluviatilis). They 135 were placed in zip-locked bags and immediately cooled at 4°C. Sampling of lake sediment was carried 136 out using cores and grabs. The sediment mapping of lakes was obtained from sediment grabs that were 137 deployed along high spatial resolution N-S and E-W transects (Fig. 2). This strategy allowed us to collect samples from the different bathymetric levels. We collected the top 10 cm of sediment. We 138 139 obtained 70, 55, 59, and 60 homogenized sediment samples from Lakes Carcans-Hourtin, Lacanau, 140 Cazaux-Sanguinet, and Parentis-Biscarrosse, respectively. For Lake Cazaux-Sanguinet the sampling 141 location was constrained by the security perimeter of the Cazaux military base. Maps of the 142 distribution of sediment properties, such as organic carbon or Hg content, were obtained from 143 interpolation between grab and core samples, and bathymetry was used for extrapolation to map whole 144 lakes. Additional grabs have been collected to validate the sediment mapping based on bathymetry.

145 In addition, sediment cores allowed us to define the vertical distribution of reactive 146 compounds at the water-sediment interface. Sediment cores were collected at three to nine stations for 147 each lake. Stations were selected according to the sedimentary map established with sediment grabs. 148 Cores were collected in organic deposits and sandy sediments from each lake, and also in the 149 vegetated sediment of Lake Parentis-Biscarrosse. Some stations were sampled twice in two different 150 seasons; in April 2014 and in January 2015 for the two northern lakes, and in spring and summer 2016 151 for the two southern lakes. Bottom water in contact with sediment cores was always oxic. Core 152 sampling was carried out by scuba divers using 10-cm-diameter polycarbonate tubes. Sealed cores 153 were immediately brought to the shore and cut horizontally into several subsamples for pore water 154 extraction. Vertical subsampling and pore water extraction from lake sediment required specific 155 approaches. We distinguished broadly two kinds of sediment: sandy permeable sediments and very 156 fluid organic-rich muds. This latter sediment type was subsampled with a vertical resolution of 2 cm 157 using Tygon tubing connected to a 100-mL syringe. Sandy permeable sediment was collected using a 158 PVC tube pre-cut in the vertical dimension. The core was laid horizontally on its side and immediately 159 halved into two similar parts. The sediment was cut rapidly into 1- to 2-cm-thick slices. This technique 160 prevented the percolation of pore waters in the vertical dimension during subsampling. For each level 161 of sandy and muddy sediment, a subsample was immediately sealed in a pre-weighed clean vial and 162 frozen under an N<sub>2</sub>-atmosphere for further analysis of the solid fraction and determination of water
 163 content. Another subsample was put into a centrifuge vial.

164 Pore waters were extracted by centrifugation at 2100 g for 20 min under an inert  $N_2$ -165 atmosphere. For muddy sediments, the supernatant was immediately filtered (0.2-µm cellulose acetate 166 syringe filter). For sandy sediments, we used 0.2-µm VivaSpin20 centrifuge vials (Anschutz and 167 Deborde, 2016). The possibility that traces of oxygen had affected the pore water concentrations of 168 reduced elements during the slicing and filtration could not be excluded. Our strategy to prevent 169 oxidation of reduced compounds was to handle the samples in less than 30 min from slicing to 170 conditioning. One part of filtered interstitial water was frozen at  $-18^{\circ}$ C, and another part was acidified 171 with HCl at a pH close to 2 to measure dissolved manganese and iron. Soils of the Landes de 172 Gascogne forest were sampled to estimate the geochemical mercury background levels of the area. 173 Soils were collected every 5 cm from the surface to a depth of 1 m at three stations in young to mature 174 forests of the Lake Cazaux-Sanguinet watershed.

175 Lake water and stream waters of the drainage area were collected to measure dissolved 176 sulphate, Fe, and Mn (Buquet et al., 2017). A total of 13 streams were sampled every month in 2014 177 and 2015. Sampling locations were along roads parallel to the lakes at about 1 km from the east shore. 178 Samples were also collected in headwaters close to agricultural areas. Sediment from the beds of rivers 179 feeding the northernmost lakes was collected manually using 50-mL centrifuge tubes in order to 180 measure total mercury concentrations. Surface lake waters were sampled every month at several 181 stations using a 2-L Niskin bottle. Bottom waters were also sampled at deep stations, where cores were 182 collected on the western side of the lakes. More than 50 samples were collected for each lake. For 183 determination of dissolved sulphate, Fe, and Mn concentration, water was filtered in situ with 0.20-um 184 cellulose acetate syringe filters. Filtered samples were stored in polypropylene tubes in a fridge after 185 acidification with HCl. Solutes were analysed within 3 weeks.

- 186 **2.3. Analyses**
- 187 2.3.1. Fish

Fish dissection was performed within 48 h in laboratory conditions. Dorsal muscle tissue free of skin was dissected and frozen then freeze-dried and crushed. Based on the fact that size is correlated with the age of the individuals, only zander between 50 and 70 cm (standard length), pike between 50 and 70 cm and perch between 18 and 23 cm were considered to compare Hg concentrations between lakes.

### 193 *2.3.2. Solid fraction*

194 The grain-size distribution was measured using a Malvern laser diffraction particle size 195 analyser. For organic sediments, granulometry was carried out after organic matter removal with  $H_2O_2$ 196 to recover only the mineral particulate fraction. The bulk mineralogy was determined for this fraction 197 by X-ray diffraction using a Siemens D500 X-ray diffractometer with CuK $\alpha$  radiation (1.5405-Å 198 wavelength). Scans were done from 5° to 80° 2 $\theta$  at 0.02° s<sup>-1</sup>, using a 40-kV accelerating voltage and 199 30-mA current. Peak identification and relative abundance estimates of minerals were determined 200 using EVA interpretation software and the DIFFRAC software package.

Particulate carbon and total sulphur were measured on freeze-dried samples by infrared spectroscopy (LECO 200 C-S analyser). Organic carbon was measured after removal of carbonates with 2 M HCl from 50 mg of powdered sample (Etcheber et al., 1999). The detection limit was 0.2%for carbon and 0.1% for sulphur. The precision of measurement was better than 5%. Ascorbate solution was used to obtain the most reactive particulate Fe(III) and Mn(III,IV) fractions (Fe-asc, Mnasc) (Kostka and Luther, 1994). Iron and manganese content was analysed colorimetrically according to standardized techniques (Anschutz and Deborde 2016) with ± 5% precision.

Total mercury concentration was determined in 100 mg dry and crushed sediment and in fish muscle by cold vapour atomic absorption spectrometry after incineration and amalgamation, using an Advanced Mercury Analyser (Altec AMA 254). The Teflon grinding bowls for fish and the mortar for sediments were washed with HCl 3% and rinsed with milli-Q water between each sample. The analytical results were quality-checked by analysing international certified reference materials (IAEA 407 for fish and PACS-2 and MESS-3 for sediment) after each set of 10 samples. For this method, precision was  $\pm$  5% and the detection limit was 0.1 ng Hg. 215 Methyl mercury content in solid sediment was determined in samples of eight chosen cores 216 collected in winter or early spring, one organic sediment and one sandy sediment for each lake. 217 Measurements were performed by gas chromatography coupled to inductively coupled plasma mass 218 spectrometry (Trace Ultra GC-XII Series ICPMS, Thermofisher). Sample preparation and analytical 219 protocol were adapted from Bouchet et al. (2013) and Renedo et al. (2017). Mercury species were 220 extracted from 0.25 g of dry and crushed sediment in 5 mL of 6N ultrapure grade HNO<sub>3</sub> solution using 221 a microwave system (Discover SP-D, CEM). Digestion was achieved in CEM Pyrex vessels by 1 min 222 of warming up to 75 °C and 3 min at 75 °C with magnetic agitation to homogenize the samples. Prior 223 to GC injection, sample extracts were ethylated at pH 4 (NaBEt<sub>4</sub>, 5% v:v, Merseburger Spezialchemikalien), in order to produce volatile ethylated forms of Hg, then extracted in isooctane by 224 225 mechanical shaking (elliptic table, 20 min) and readily separated by gas chromatography. 226 Quantification was performed by species-specific isotope dilution by spiking known amounts and concentrations of isotopically enriched standard solutions (MM<sup>201</sup>Hg and <sup>199</sup>Hg(II), ISC) (Rodriguez 227 Martin-Doimeadios et al., 2004). The detection limit of the method was 0.03 ng Hg g<sup>-1</sup>. The analytical 228 229 results were continuously quality-checked by analysing reference materials certified for Hg speciation 230 (IAEA-405 Estuarine Sediment). Relative standard deviation of MeHg concentration has been 231 evaluated to 1.7%, from triplicate preparations and analyses of the reference material.

232 2.3.3. Dissolved compounds

The sediment water content was determined from the weight difference between wet and dry sediments. Dissolved iron (Fe<sup>2+</sup>) was analysed using the colorimetric method by adding a ferrozine solution in an aliquot (Stookey, 1970). Dissolved manganese (Mn<sup>2+</sup> and Mn<sup>3+</sup>) was determined colorimetrically using a Cd–TCPP complex according to Madison et al. (2011). Dissolved Fe and Mn were determined with  $\pm$  5% precision. Dissolved sulphate was analysed by a nephelometric method based on BaSO<sub>4</sub> precipitation using BaCl<sub>2</sub> (Rodier, 1976). The detection limit was 10 µM, and the precision of replicate samples was better than 8%.

240 **3.** *Results* 

**3.1. Mercury in fish** 

We observed the highest average total Hg concentrations in carnivorous fish from Lake Carcans-Hourtin (Table 2), with a maximum value of 6.238 mg kg<sup>-1</sup> dw. Total Hg concentrations were two to five times lower in fish collected from Lake Parentis-Biscarrosse. We measured intermediate values in fish from Lake Lacanau and Lake Cazaux-Sanguinet. Therefore, the data revealed a decreasing gradient from north to south for total Hg in piscivorous species.

247

# **3.2. Sedimentary cover of lakes**

# 248 *3.2.1.* Sediment mapping

249 Sediment type was strongly influenced by bathymetry. Therefore, we used bathymetry to draw 250 the sediment map from the grab and core observations (Fig. 2). Deep areas in the western parts of the 251 lakes presented muddy deposits. These deposits had a homogeneous appearance and a very soft 252 consistency. The water content of this substrate was greater than 95%, making the sediment texture 253 very fluffy. These sediments were highly organic deposits. Organic deposits were found below 5 m 254 depth in lakes Carcans-Hourtin and Lacanau, below 7 m in Lake Cazaux-Sanguinet, and below 10 m 255 in Lake Parentis-Biscarrosse. For these two last lakes, deep areas also presented a mixture of these 256 organic deposits with sand, called sandy organic deposits or organic sands, depending on the 257 proportion of sand and organic carbon (Fig. 2). Organic sands were present in sub-lacustrine channels. 258 These channels are the paleo-valley of the rivers that flowed to the ocean before the formation of lakes 259 through the development of coastal dunes in the last thousand years (Buquet et al., 2017). Shallow 260 zones presented sands with shades of green, yellow, black, or grey depending on the content of iron 261 oxides or organic matter. The water content of sands was 20% in weight.

Vegetated areas of lakes Lacanau and Parentis-Biscarrosse consisted of sandy sediments. In sheltered
areas, 2 to 10 cm of muddy sediment covered the sand.

264 *3.2.2. Grain size* 

Sand grain size after organic matter removal with  $H_2O_2$  was very homogeneous, with a median size of 362  $\mu$ m, a standard deviation (SD) of 49  $\mu$ m, and a standard error of the mean (s.e.) of 7  $\mu$ m. The sand was the same as the soils of the lake catchment (Fig. 3). The grain size of the mineral fraction of organic deposits was centred between 10 and 20 µm (Fig. 3). X-ray diffraction indicated that the mineral fraction consisted of quartz, clay minerals (illite and kaolinite), and feldspars.
Carbonate minerals were not detected.

271 *3.2.3. Total carbon and total sulphur* 

Particulate organic carbon (POC) and total carbon (TC) measurements on grab and core sediments gave the same concentration, indicating that TC consisted exclusively of POC and that inorganic carbonate was absent or very scarce. TC concentration ranged from 10% to 30% in organic deposits. Averaged TC concentrations ( $\pm$  SD) in organic sediments were 15.8% ( $\pm$  3.1), 17% ( $\pm$  1.4), 15.5% ( $\pm$  1.7), and 15.2% ( $\pm$  0.9) for lakes Carcans-Hourtin, Lacanau, Cazaux-Sanguinet, and Parentis-Biscarrosse, respectively (Fig. 4). Concentrations ranged from 3% to 10% in sandy organic deposits, and from 1% to 3% in organic sands. TC was below 1% in sands.

Total sulphur (TS) content was also dependent on the type of sediment. TS ranged from 1% to 4% in organic deposits (Fig. 4). The mean concentrations were close to 2.5% ( $\pm$  0.8% SD) except for organic sediments of Lake Parentis-Biscarrosse, in which the value was 1.2%. TS ranged from 0.1% to 1% in sandy organic deposits and organic sands. TS was below 0.1% in sands.

283

## *3.2.4. Particulate total mercury (HgTp)*

284 HgTp concentrations were closely related to the nature of sediment and the organic carbon 285 concentrations. Classes of mercury concentration fitted classes of TC content (Fig. 2). HgTp concentrations ranged from 100 to 420  $\mu$ g kg<sup>-1</sup> in organic deposits (means of 230 (± 82 SD), 203 (± 60 286 SD) 206 (± 68 SD), and 142 µg kg<sup>-1</sup> (± 53 SD) for lakes Carcans-Hourtin, Lacanau, Cazaux-287 Sanguinet, and Parentis-Biscarrosse, respectively). HgTp concentrations in Lake Parentis-Biscarrosse 288 289 were significantly lower (ANOVA test) than in other lakes (Fig. 4). HgTp concentrations ranged from 20 to 100  $\mu$ g kg<sup>-1</sup> in sandy organic deposits. HgTp concentrations ranged from 4 to 20  $\mu$ g kg<sup>-1</sup> in 290 organic sands. Finally, HgTp concentrations ranged from below the detection limit to 10 µg kg<sup>-1</sup> in 291 sands (Fig. 2). 292

*3.2.5. Geochemical background levels* 

Sand samples from forest soil east of Lake Cazaux-Sanguinet had the same particle size distribution as the sandy sediments from lakes (Fig. 3). HgTp concentrations ranged from 0.5 to 21  $\mu$ g kg<sup>-1</sup>. Highest values were associated with soil samples enriched in organic carbon. However, HgTp values were not significantly different (ANOVA test) in lake sands and in drainage basin soils (Fig. 2). Fine sediments collected from the beds of streams that feed Lake Carcans-Hourtin and Lake Lacanau had a mean HgTp concentration of  $2.3 \pm 1.8 \,\mu$ g kg<sup>-1</sup>, which was much lower than values measured in lake organic sediments.

301

#### 3.3. Lake sedimentary column

Cores of organic deposits had a TC concentration between 10% and 30% (Fig. 1S, 302 303 Supplementary Material), like sediment grabs. TC decreased slightly with depth, except for Lake Carcans-Hourtin, which showed a sharp decrease in TC below 10 cm depth from 30% to 15%. TS 304 305 concentrations were similar to values measured in grabs. Concentrations increased with depth, except 306 in organic muds of Lake Carcans-Hourtin, where sediments below 25 cm depth were less enriched in 307 sulphur than those above. Iron and manganese oxide concentrations extracted by leaching with 308 ascorbate decreased with depth. Ascorbate-extracted iron was two- to three-fold higher in muddy 309 sediments of Lake Parentis-Biscarrosse than in other lakes. HgTp concentrations were between 100 and 300 µg kg<sup>-1</sup> in organic sediments. Shallow vegetated sediment from Lake Parentis-Biscarrosse 310 contained up to 80 µg kg<sup>-1</sup> HgTp in the muddy layer of the superficial sediment. This muddy layer of 311 312 vegetated sediment contained about 10% TC.

Methylmercury concentrations of organic sediments were maximal in the first centimetres below the water-sediment interface (Fig. 1S, Supplementary Material). There was an increasing gradient in MeHg from the southern lakes to the northern lakes: average vertical concentrations in the top 10 cm below the sediment-water interface were 4.61 ( $\pm$  2.6 SD) µg kg<sup>-1</sup> for Lake Carcans-Hourtin, 5.14 ( $\pm$  2.8 SD) µg kg<sup>-1</sup> for Lake Lacanau, 1.56 ( $\pm$  0.14 SD) µg kg<sup>-1</sup> for Lake Cazaux-Sanguinet, and 1.1 ( $\pm$  0.32 SD) µg kg<sup>-1</sup> for Lake Parentis-Biscarrosse, which represented 2.53% ( $\pm$  1.0), 1.69% ( $\pm$ 0.7), 0.55% ( $\pm$  0.1), and 0.53% ( $\pm$  0.1) of HgTp, respectively. 320 Concentrations of TC, mercury, Fe and Mn oxyhydroxides, and sulphides were lower by a 321 factor of 10 to 100 in sandy sediment cores compared to muddy sediments. Contrary to observations 322 for organic sediments, mercury decreased with depth. MeHg concentrations were often below the 323 detection limit of the GC-ICPMS method (i.e.  $0.03 \mu g Hg kg^{-1}$ ).

324

## **3.4. Lake water and pore waters**

325 Sandy sediment depleted in organic carbon (e.g, Fig. 1S-A and 1S-G, Supplementary Material) did not show a pronounced gradient of dissolved redox compounds below the sediment 326 327 surface, except a small sulphate decrease with depth. Pore water dissolved Fe and Mn remained at concentrations similar to those measured in lake waters. Sandy sediments with an organic carbon 328 329 content above 0.5% showed more pronounced redox gradients, with appearance of reduced dissolved 330 species and rapid loss of sulphate in the first centimetres below the interface (Fig. 1S-H and 1S-J, 331 Supplementary Material). Dissolved iron and manganese concentrations were present from the first 332 centimetre of the sedimentary pore water column in cores that were collected in muddy sediments. 333 Dissolved sulphate concentrations decreased below the sediment surface in organic sediment pore 334 water to reach the detection limit a few centimetres below the interface. Dissolved sulphate 335 concentrations of overlying water differed depending on the lake. Mean water sulphate concentrations 336 were 328 (± 28 SD), 221 (± 26 SD), 100 (± 19 SD), and 82 (± 17 SD) µM for lakes Carcans-Hourtin, 337 Lacanau, Cazaux-Sanguinet, and Parentis-Biscarrosse, respectively (N > 50 for each lake). Therefore, 338 there was a decreasing north-south gradient of sulphate concentrations. Mean dissolved sulphate 339 concentration of the stream waters that feed the lakes was lower than 150  $\mu$ M, except for the Lake 340 Carcans-Hourtin catchment, where streams contained up to 600 µM sulphate (Fig. 5). For this lake, 341 sulphate concentrations were the highest in head waters close to agricultural areas. Values above 1000 342 µM were measured in upstream waters of the Caillava river, which drain large maize fields (Fig. 5). Head waters that drain maize fields of the Cazaux-Sanguinet catchment were also enriched in 343 dissolved sulphate, with mean concentrations of 300 µM. 344

345 **4.** Discussion

#### 346 **4.1. Mercury distribution, diagenetic processes, and mercury mobility in sediments**

347

### 4.1.1. Mercury distribution and background concentrations

The concentration of HgTp was close to 200  $\mu$ g kg<sup>-1</sup> in organic muds and below 20  $\mu$ g kg<sup>-1</sup> in 348 sandy sediments. Concentrations were below 2 µg kg<sup>-1</sup> in sands poor in organic carbon. Sandy 349 350 sediments had concentrations identical to those measured in forest sandy soil from the lake drainage 351 basin. Sands were deposited before the formation of Aquitaine lakes, whereas the fine particles present 352 in organic deposits were autochthonous sediments. Higher HgTp values in organic muds may result 353 from a grain-size effect: deep parts of lakes accumulate fine-grained particles that consist of fine 354 quartz and clay minerals, which are probably enriched in Hg relative to sandy sediments (Bloom and 355 Crecelius, 1987). However, grain size alone cannot explain the difference, because mercury has a 356 strong affinity with organic carbon, sulphur, and Fe/Mn oxides (Feyte et al., 2010), which are highly 357 enriched in muddy sediment. The difference in HgTp content between sands, sands enriched in organic matter, and muds is most likely explained by the organic matter content (Fig. 2), suggesting 358 359 that organic matter is the initial carrier phase of Hg that accumulates in the sediment.

#### 360

### 4.1.2. Diagenetic processes

361 Mercury has affinity with organic matter, iron/manganese oxyhydroxides, and sulphides. 362 Therefore, it is strongly influenced by diagenetic processes (Matty and Long, 1995). These processes are directly linked to benthic organic matter mineralization. Mineralization of organic matter is the 363 result of its oxidation, carried out in presence of microorganisms, and is accompanied by reduction of 364 electron acceptors according to a sequence of reactions where O<sub>2</sub> is used preferentially, then nitrate 365 when the sediment becomes depleted in dissolved oxygen. In anoxic sediments, mineralization 366 consumes oxides and oxyhydroxides of manganese (MnO<sub>2</sub>, MnO(OH)) and iron (Fe(OH)<sub>3</sub>) present in 367 the solid fraction of sediments, and dissolved sulphate  $(SO_4^{2-})$  and  $CO_2$ , which generally represent 368 369 both the main electron acceptors in anoxic environments (Stumm and Morgan, 1996). Anoxic 370 sediment is generally enriched in particulate sulphur because of the precipitation of sulphide minerals 371 as a result of sulphate reduction.

372 Redox species profiles in the cores (Fig. 1S, Supplementary Material) agreed with a typical 373 diagenetic sequence. Iron and manganese oxides extracted with an ascorbate solution have the highest 374 concentrations at the sediment surface and decrease with depth. Dissolved reduced iron and 375 manganese appear in the top core samples. This indicates that sediments are anoxic from the first 376 centimetre of the sediment column. Sulphate concentration decreases rapidly and reaches a value 377 below the detection limit a few centimetres below the sediment surface, which means that the sulphate 378 reduction process occurs effectively. Sulphide precipitate is produced as FeS, as suggested by the high 379 particulate S content in organic deposits and the decreasing pore water iron concentration with depth. 380 It must be noted that the concentration gradients in organic sediments are stronger than gradients in 381 sandy sediments, suggesting that diagenetic processes are more intense in muddy sediments. However, 382 the decrease and disappearance of sulphate in sandy cores suggest that the small amount of organic 383 matter present in sands is labile.

384

### 4.1.3. Mercury diagenesis

385 Mercury transformation in sediments depends on many biogeochemical processes, which may 386 affect the ability of mercury to be methylated, or to migrate from sediment to the water column 387 (Schäfer et al., 2010). Mercury mobility depends on its speciation between solid and dissolved phases. 388 This distribution is intimately coupled to organic matter mineralization as Hg distribution is influenced 389 by the presence of Mn/Fe oxyhydroxides and sulphide minerals. In oxic sediment layers, Hg mobility 390 is generally low because it is essentially associated with the solid phase by adsorption or precipitation 391 with Fe/Mn oxyhydroxides (Feyte et al., 2010). During the organic matter mineralization process, 392 Fe/Mn oxyhydroxides may be reduced, and associated Hg is released (Matty and Long, 1995). In the 393 sediment cores studied, the decrease in Fe/Mn oxyhydroxide concentrations with depth in lakes 394 Carcans-Hourtin, Lacanau, and Cazaux-Sanguinet can be interpreted as metal oxide reduction. In Lake 395 Parentis-Biscarrosse, the oxide concentration remains high in all organic cores, suggesting that Hg mobility is limited in this lake. In anoxic sediments, sulphate reduction leads to sulphide production, 396 397 for which Hg has a strong affinity. Mercury precipitates as HgS or it is absorbed on FeS and included 398 in pyrite (Fitzgerald and Lamborg, 2003). Grab samples show that the Hg concentration increases 399 when total S concentration increases as well (Fig. 6). This trend also occurs in organic sediment cores. 400 Sulphur is associated with organic matter either as an initial compound or as a result of the 401 sulphurization by sulphide of organic matter during diagenesis (Urban et al., 1999). Therefore, high 402 organic matter concentrations correspond to high sulphur concentrations. Urban et al. (1999) found an 403 averaged C:S ratio of organic matter between 167 and 198 based on sediment cores from 50 lakes. For 404 lake sediments with the highest concentrations of organic carbon they found a maximum organic sulphur concentration of 260  $\mu$ mol g<sup>-1</sup> (0.83%), which is below the concentration of sulphur measured 405 406 in muddy sediments (Fig. 6). This indicates that most of the sedimentary sulphur occurs as particulate 407 sulphide, and that particulate sulphides are major carrier phases of Hg in organic sediments. Organic 408 matter itself presents many adsorption sites for Hg, which plays a role in Hg mobility (Benoit et al., 409 1998). This probably explains why organic sediments are enriched in Hg relative to organic-poor 410 sandy sediments (Fig. 2). Organic matter is mineralized in sediments through early diagenesis 411 processes. In anoxic lake sediments, Hg initially trapped in the fraction of organic matter that is 412 consumed is most likely released into the interstitial waters and then quickly trapped by sulphides 413 when sulphate reduction occurs (Benoit et al., 1999).

414

# 4.2. Methylation

# 415 *4.2.1.* Vertical distribution

Organic sediments of Lake Carcans-Hourtin and Lake Lacanau show the highest 416 417 concentrations of MeHg at the depth where the decreasing gradient of dissolved sulphate occurs. 418 Inorganic Hg methylation takes place during Hg(II) absorption by some anaerobic microorganisms 419 which methylate Hg in their cytoplasm (Benoit et al., 1999; Gilmour et al., 1998). Microbial sulphate 420 reduction (MSR) is recognized to be the most common pathway leading to mercury methylation in 421 sediments (Barkay and Wagner-Döbler, 2005; Compeau and Bartha, 1985). Other anaerobic 422 microorganisms such as iron-reducing bacteria are able to methylate Hg (Podar et al., 2015). 423 Diagenetic profiles of organic cores show that anaerobic processes occur at the depth of the maximum 424 MeHg concentrations (Fig. 1S, Supplementary Material). This suggests that the predominant process 425 of mercury methylation is carried out by a consortium of anaerobic microorganisms, including those 426 involved in MSR. Communities of Fe- and sulphate-reducing bacteria are also involved in MeHg 427 demethylation processes (Oremland et al., 1991; Bouchet et al., 2013). It is the balance between 428 methylation and demethylation in sediment that dictates the standing pool of MeHg (Rodríguez 429 Martín-Doimeadios et al., 2004). The progressive decrease in MeHg with depth of the sedimentary 430 column could be related to the dominance of demethylation processes in the deep sediment, when 431 sulphate becomes depleted (Pak and Bartha 1998).

432

## 4.2.2. Spatial distribution

Organic sediments of the four lakes have different MeHg concentrations. The fraction of total Hg present as MeHg (%MeHg) describes a positive south–north gradient from 0.5% in Lake Parentis-Biscarrosse to 2.5% in Lake Carcans-Hourtin (Fig. 7). The proportions between 0.5% and 0.6% obtained in the southernmost lakes are consistent with those found in the literature for lacustrine or coastal systems (Table 3). Generally, MeHg concentration in sediment is less than 1% of HgTp (Morel et al., 1998). MeHg proportions in northern lake sediments are high (especially Lake Carcans-Hourtin), which means that the methylation potential is high in these lake sediments.

440 Main factors that are known to influence the methylation rate are Hg(II) availability and 441 bacteria activity within the sedimentary column. Sulphate inputs stimulate MSR activity. As sulphate 442 concentrations differ between lakes, the high proportion of MeHg in northern lakes, more particularly 443 in Lake Carcans-Hourtin, can be related to high methylation potential as sulphate concentrations are 444 the highest (Fig. 7). Previous studies have found that sulphate addition stimulates net methylation rates 445 in fresh water sediments (e.g, Gilmour et al. 1992; Åkerblom et al. 2013). Sulphide from sulphate 446 reduction is known to have two effects on the availability of Hg(II). At low concentrations, sulphide 447 promotes methylation. By contrast, for higher concentrations, as may be the case in marine sediments, 448 sulphide limits the bioavailability of Hg(II) to be methylated (Benoit et al., 1998; Hsu-Kim et al., 449 2013; Johnson et al., 2016). This suggests that there is a range of sulphate concentration which is 450 optimal for Hg methylation, above which methylation is inhibited. A broad range in the sulphate 451 concentration associated with maximum efficiency of Hg methylation has been observed. For 452 example, Orem et al. (2014) observed maximum MeHg concentrations at sulphate concentrations of 2

and 10–15 mg L<sup>-1</sup>in the Everglades Protection Area. Peak MeHg was observed in wetland mesocosms 453 with a sulphate concentration of 59 and 93 mg L<sup>-1</sup> (Myrbo et al., 2017). In the case of Aquitaine lake 454 455 muddy sediments, MeHg concentrations increase with sulphate, suggesting that MSR is not limited, 456 and that produced sulphide does not inhibit Hg availability. The availability of dissolved sulphide 457 produced by MSR will depend on iron oxide availability. The reactive fraction of iron oxides 458 determined by the ascorbate reagent is above 100  $\mu$ mol g<sup>-1</sup> in the surface sediments of the four lakes, which represents at least, when considering sediment water content, 4000 µmol L<sup>-1</sup>, suggesting that 459 460 the sulphide produced is efficiently trapped as Fe sulphide phases with this abundant available reactive 461 Fe.

462 Temperature is an important control factor for methylation process. Low temperatures will slow down methylation(Hintelmann and Wilken, 1995; Ramlal et al., 1993). The cores studied were 463 sampled in different seasons, but MeHg was measured only in cores collected in winter and early 464 spring. Higher concentrations of MeHg in summer may be expected, although the effect of 465 466 temperature on biotic demethylation is not well known. Gradients of dissolved sulphate between the water column and the sediment were sharper when the water temperature was the highest at a given 467 station sampled in two seasons, suggesting that the sulphate reduction rate, and the related 468 469 methylation, was more efficient in summer and autumn.

470 The gradient of mercury contamination in carnivorous fish muscles between the four lakes 471 (Table 2) fits well with sediment MeHg concentrations, since the highest levels were measured in fish 472 from northern lakes. Therefore, the muddy sediment compartment may be one of the main sources of 473 MeHg which accumulates in the trophic chain (e.g. Hammerschmidt et al., 2004). Periphyton 474 microbial communities of aquatic macrophytes of Lake Cazaux-Sanguinet have also been recognized 475 to potentially methylate Hg (Gentès et al., 2013). This study was based on experimental batches placed 476 under high environmental Hg concentrations and anoxic conditions. Hence, through dietary 477 bioaccumulation, the MeHg in higher trophic levels within Aquitaine lakes may be attributed to MeHg 478 produced in anoxic muds, as observed in the present study and in the periphyton (Gentès et al., 2013), 479 both being limited by sulphate.

480 Sulphate from lake waters may originate from several sources: atmospheric inputs from rain 481 and ocean spray, rocks from the drainage area, and anthropogenic inputs. The distance from lake to 482 ocean is the same for the four lakes studied, suggesting that the atmospheric sources by spray are 483 similar. The lake catchment geology is also the same for the four lakes. The major difference between 484 lake catchments is the land use. The drainage area of Lake Carcans-Hourtin has the highest proportion 485 of agriculture areas (Fig. 1). Tributaries of this lake are enriched in dissolved sulphate, with a 486 concentration as high as 1200 µM in ditches located upstream, in the cultivated areas (Fig. 6). Head 487 waters collected in ditches close to agriculture surfaces of the Lake Cazaux-Sanguinet catchment are 488 enriched in dissolved sulphate compared to waters collected downstream, but the concentrations 489 remain below 300  $\mu$ M. Thus, agricultural activity is the main source of sulphate. Discussions with 490 farmers informed us of the fact that sulphate was supplied indirectly to croplands through liming, 491 which allows control of soil pH. Lime comes from limestone or dolomite quarries, and in some cases 492 in the recent past from neighbouring paper mill plants that produce lime sludge as a by-product. 493 Depending on lime origin, the sulphate content can be high, and lime can become a source of sulphate 494 for lakes.

495

### 4.2.3. MeHg diffusion and bioavailability

496 Our study points out that MeHg concentration is related to sulphate concentration. Methyl 497 mercury produced in the sediment can be transferred to the water column through molecular diffusion 498 and resuspension of surface sediments. Hence, the amount of MeHg diffusing into the water column 499 and thus the amount bioavailable to pelagic organisms depends, among other things, on the proportion 500 of lake floor carpeted with organic sediments. Organic deposits account for 21%, 20%, 19%, and 18% 501 of the surface area of lakes Carcans-Hourtin, Lacanau, Cazaux-Sanguinet, and Parentis-Biscarrosse, 502 respectively. These proportions are close and suggest that the proportion of organic sediment is not the 503 variable that explains the gradient of fish contamination. Bioturbation and bioirrigation may affect the 504 transport of Hg from sediment to water (Benoit et al., 2006). We observed the presence of chironomid 505 larvae in some grabs from the four lakes. The density of these organisms was low, with zero to three 506 individuals per grab. Even if the density of benthic organisms was low, burrowing organisms may 507 assimilate MeHg directly from underlying anoxic sediments and become vectors of MeHg to pelagic 508 predators (Gagnon et al., 1996). Finally, despite the occurrence of episodic bottom-water anoxia 509 during the summer in Lake Parentis-Biscarrosse, this lake remains the one in which the fish are less 510 contaminated with Hg. The methylation of Hg in a polymictic lake can be shifted from the sediment to 511 the anoxic water column, because sulphate reduction is shifted in the same way to the water column 512 (Ramlal et al., 1993; Wartras et al., 1995). This suggests that sulphate availability remains a key 513 environmental parameter for Hg methylation, wherever sulphate reduction occurs, in the water column 514 or at the sediment surface.

## 515 **4.3. Sources of Hg**

516 Land use is not impacted by activities that may represent point sources of Hg. HgTp 517 concentrations are similar in lake sandy sediments and in watershed sandy soils, with values below 20  $\mu$ g kg<sup>-1</sup>, suggesting that the geological background level is low. Moreover, we measured HgTp 518 519 concentrations of between 1 and 7 µg kg<sup>-1</sup> in fine sediments of river beds, which is low compared to 520 the 200 µg kg<sup>-1</sup> measured in lakes muds, suggesting that the contribution of Hg from lake catchment is 521 negligible. Hg distribution in lakes depends exclusively on sediment type, not on geography: there is no zone that presents a peak of HgTp concentration. This suggests the absence of a point source in 522 523 lakes. Several lines of evidence suggest that Hg that accumulates in muddy sediment is mainly of 524 atmospheric origin. Considering that the superposition principle applies to organic sediment deposits, the decrease in Hg concentrations when approaching the water-sediment interface (Fig. 1S, 525 526 Supplementary Material) may be explained by the decrease in atmospheric emission in European 527 countries in recent decades (UNEP, 2013).

South-western France is not a hot spot for atmospheric Hg emissions (Colette et al., 2016). In the light of the concentrations measured in lake sediments, Hg fluxes are not alarming. Average atmospheric Hg flux on the Aquitaine coast is estimated by modelling as  $10 \ \mu g \ m^{-2} \ yr^{-1}$  for the year 2001 (Roustan et al., 2006). Such a flux represents 620, 200, 580, and 350 g yr<sup>-1</sup> for lakes Carcans-Hourtin, Lacanau, Cazaux-Sanguinet, and Parentis-Biscarrosse, respectively. Considering that all Hg that reaches the lake surface is trapped in organic matter that accumulates in organic sediments,

considering also the mean HgTp of each lake and the particle content of organic sediment (40 g  $L^{-1}$ ), 534 535 we found that 1 cm of deposit contains between 1 and 1.8 years of Hg deposition. This suggests that the mean sedimentation rate is between 0.6 and 1.0 cm yr<sup>-1</sup>. A very similar value was found for lakes 536 537 Lacanau and Carcans-Hourtin using a mass balance based on dissolved inorganic nitrogen retention (Buquet et al., 2017). The higher sedimentation rate found here (1 cm yr<sup>-1</sup>) is for Lake Parentis-538 539 Biscarrosse. This lake has a higher phosphorus content than the other lakes (Cellamare et al., 2011), 540 which results in higher primary production. Eutrophication of the lake may explain a higher 541 sedimentation rate of organic matter. As a consequence, the concentration of HgTp is lower in the 542 muds of Lake Parentis-Biscarrosse than that of other lakes, even if the atmospheric flux is the same, 543 due to a dilution effect.

544 **5.** Conclusion

545 Our study presents new results from Aquitaine lake sediments. We have established the first 546 map of sediment distribution and we have determined several physicochemical properties of these sediments. More particularly, we showed that the bottom of the four lakes consists of sands. Sandy 547 548 sediments are covered with organic muds, which accumulate only in the deepest parts of lakes and in 549 shallow densely vegetated areas. The highest Hg concentrations are encountered in these organic deposits. However, they do not exceed reference thresholds for sediments (1100 µg kg<sup>-1</sup> for the 550 551 probable effect concentration; MacDonald et al., 2000). Organic sediment traps Hg in the four lakes, 552 which is most likely of background diffusive atmospheric origin. These sediments are hot spots for Hg methylation, due to sulphate reduction processes. 553

The mercury concentration gradient in the muscles of carnivorous fish is the same as the methyl mercury concentration gradient of lake sediment. This suggests that the sedimentary compartment could be a MeHg source for the water column, resulting in its accumulation along the trophic chain. Methylation rate depends on sulphate reduction activity. This activity is limited by sulphate availability. Lake Carcans-Hourtin is the most concentrated in sulphate. Lake Lacanau, whose waters come partly from Lake Carcans-Hourtin, is enriched in sulphate by this very fact. Agricultural activity is the main source of sulphate in the Lake Carcans-Hourtin drainage basin. This excess 561 sulphate that is leached from soils instead of being absorbed and exported by plants originates likely 562 from liming. Therefore, the use of sulphate-rich lime in the headwater drainage area has an indirect 563 consequence on carnivorous fish Hg content in lakes located at the outlet of the catchment, because sulphate favours methylation of the background flux of atmospheric Hg in anoxic environments of 564 lakes, such as autochthonous organic sediments, as seen in this study, or in other compartments, such 565 as aquatic plant periphyton (e.g. Gentès et al., 2013). One question that remains unresolved focuses on 566 567 the pathway that transmits MeHg produced from the sediment to fish. For that the distribution of Hg in 568 fish and in the food web is presently conducted in Aquitaine lakes.

569

### 570 Acknowledgements

We thank Cécile Bossy, Lionel Dutruch, Rémy Sinays, and Thierry Corrège for their assistance 571 during field and laboratory work. We acknowledge the assistance given by Frank Quenault. This 572 573 research was founded by the project CLAQH (Agence de l'eau Adour-Garonne, Région Nouvelle 574 Aquitaine), the Syndicat Intercommunal d'Aménagement des Eaux du Bassin Versant des Etangs du Littoral Girondin (SIAEBVELG), the LITTOLAC project supported by the French national program 575 INSU-EC2CO-BIOHEFECT, and the project PSDR AQUAVIT (Région Nouvelle Aquitaine and 576 577 INRA). This study has been carried out in the frame of the Investments for the future Program, within the Cluster of Excellence COTE (ANR-10-LABEX-45). 578

579

# 580 **References**

- Åkerblom, S., Bishop, K., Björn, E., Lambertsson, L., Eriksson, T., Nilsson, M.B., 2013., Significant interaction
   effects from sulfate deposition and climate on sulfur concentrations constitute major controls on
   methylmercury production in peatlands. Geochim. Cosmochim. Acta 102: 1–11.
- Allen, J.W., Shanker, G., Tan, K.H., Aschner, M., 2002. The consequences of methylmercury exposure on
   interactive functions between astrocytes and neurons. NeuroToxicology 23: 755–759.
- Alpers, C.N., Fleck, J.A., Marvin-DiPasquale, M., Stricker, C.A., Stephenson, M., Taylor, H.E., 2013. Mercury
  cycling in agricultural and managed wetlands, Yolo Bypass, California: spatial and seasonal variations
  in water quality. Sci. Total Environ. 484: 276–287.

- Anschutz, P., Deborde, J., 2016. Spectrophotometric determination of phosphate in matrices from sequential
  leaching of sediments. Limnol. Oceanogr. Methods 14: 245–256.
- Avramescu, M.L., Yumvihoze, E., Hintelmann. H., Ridal, J., Fortin, D.R.S., Lean, D., 2011. Biogeochemical
   factors influencing net mercury methylation in contaminated freshwater sediments from the St.
   Lawrence River in Cornwall, Ontario, Canada. Sci. Total Environ. 409: 968–978.
- Barkay, T., Wagner Döbler, I., 2005. Microbial transformations of mercury: potentials, challenges, and
   achievements in controlling mercury toxicity in the environment. Adv. Appl. Microbiol. 57:1–52
- Benoit, J.M., Gilmour, C.C., Mason, R.P., Heyes, A., 1999. Sulfide controls on mercury speciation and
  bioavailability to methylating bacteria in sediment pore waters. Environ. Sci. Technol. 33: 951–957.
- Benoit, J.M., Gilmour, C.C., Mason, R.P., Riedel, G.S., Riedel, G.F., 1998. Behavior of mercury in the Patuxent
   River estuary. Biogeochemistry 40: 249–265.
- Benoit, J.M., Shull, D.H., Robinson, P., Ucran, L.R., 2006. Infaunal burrow densities and sediment
  monomethylmercury distributions in Boston Harbor, Massachusetts. Mar. Chem. 102:124–133.
- Bertrin, V., Boutry, S., Jan, G., Ducasse, G., Grigoletto, F., Ribaudo, C., 2017. Effects of wind-induced sediment
  resuspension on distribution and morphological traits of aquatic weeds in shallow lakes. J. Limnol.
  76(s1) https://doi.org/10.4081/jlimnol.2017.1678
- Bloom, N.S., Crecelius, E.A., 1987. Distribution of silver, mercury, lead, copper and cadmium in central puget
  sound sediments. Mar. Chem. 21: 377–390.
- Bouchet, S., Amouroux, D., Rodriguez-Gonzalez, P., Tessier, E., Monperrus, M., Thouzeau, G., Clavier, J.,
  Amice, E., Deborde, J., Bujan, S., Grall, J., Anschutz, P., 2013. MeHg production and export from
  intertidal sediments to the water column of a tidal lagoon (Arcachon Bay, France). Biogeochemistry
  114: 341–358.
- Bowles, K.C., Apte, S.C., Maher, W.A., Kawei, M., Smith, R., 2001. Bioaccumulation and biomagnification of
  mercury in Lake Murray, Papua New Guinea. Can. J. Fish. Aquat. Sci. 58: 888–897.
- Buquet, D., Anschutz, P., Charbonnier, C., Rapin, A., Sinays, R., Canredon, A., Bujan, S., Poirier, D., 2017.
  Nutrient sequestration in Aquitaine lakes (S France) limits nutrient flux to the coastal zone. J. Sea Res.
  130: 24–35
- Canton, M., Anschutz, P., Coynel, A., Polsenaere, P., Auby, I., Poirier, D., 2012. Nutrient export to an eastern
  Atlantic coastal zone: First modeling and nitrogen mass balance. Biogeochemistry 107: 361–377.
- 618 Castelle, S., 2008. Spéciation et réactivité du mercure dans le système fluvio-estuarien Girondin. Thesis,
  619 University Bordeaux 1.
- Cellamare, M., Morin, S., Coste, M., Haury. J., 2011. Ecological assessment of French Atlantic lakes based on
   phytoplankton, phytobenthos and macrophytes. Environ. Monit. Assess. 184: 4685–4708.
- Colette, A., Aas, W., Banin, L., et al., 2016. Air pollution trends in the EMEP region between 1990 and 2012.
  Joint Report of the EMEP Task Force on Measurements and Modelling (TFMM), Chemical Coordinating Centre (CCC), Meteorological Synthesizing Centre-East (MSC-E), Meteorological
  Synthesizing Centre-West (MSC-W). Kjeller, NILU (EMEP: TFMM/CCC/MSC-E/MSC-W Trend
  Report) (EMEP/CCC, 01/2016).
- 627 Compeau, G.C., Bartha, R., 1985. Sulfate-reducing bacteria: principal methylators of mercury in anoxic
  628 estuarine sediment. Appl. Environ. Microbiol. 50 : 498–502.

- 629 Cossa, D., Ficht, A., 1999. La dynamique du mercure. Editions Quae, Versailles.
- Driscoll, C.T., Mason, R.P., Chan, H.M., Jacob, D.J., Pirrone. N., 2013. Mercury as a global pollutant: Sources,
  pathways, and effects. Environ. Sci. Technol. 47: 4967–4983.
- Etcheber, H., Relexans, J.C., Beliard, M., Weber, O., Buscail, R., Heussner, S., 1999. Distribution and quality of
  sedimentary organic matter on the Aquitanian margin (Bay of Biscay). Deep Sea Res. Part II Top. Stud.
  Oceanogr. 46: 2249–2288.
- Feyte, S., Tessier, A., Gobeil, C., Cossa, D., 2010. In situ adsorption of mercury, methylmercury and other
  elements by iron oxyhydroxides and organic matter in lake sediments. Appl. Geochem. 25: 984–995.
- Fitzgerald, W.F., Lamborg, C.H., 2003. Geochemistry of mercury in the environment. Treatise on Geochemistry
  9, Elsevier, pp 107–148.
- Fleck, J..A, Marvin-DiPasquale, M., Eagles-Smith, C.A., Ackerman, J.T., Lutz, M.A., Tate, M., Alpers, C.N.,
  Hall, B.D., Krabbenhoft, D.P., Eckley, C.S., 2016. Mercury and methylmercury in aquatic sediment
  across western North America. Sci. Tot. Environ. 568 : 727–738.
- Gagnon, C., Pelletier, E., Mucci, A., Fitzgerald, W.F., 1996. Diagenetic behavior of methylmercury inorganicrich coastal sediments. Limnol. Oceanogr. 41, 428–434.
- Gentes, S., Monperrus, M., Legeay, A., Maury-Brachet, R., Davail, S., Andre, J.M., Guyoneaud, R., 2013.
  Incidence of invasive macrophytes on methylmercury budget in temperate lakes: Central role of bacterial periphytic communities. Environ. Pollut. 172: 116–123.
- 647 Gilmour, C.C., Henry, E,A, Mitchell, R., 1992. Sulfate Stimulation of mercury methylation in freshwater
  648 sediments. Environ. Sci. Technol. 26: 2281–2287.
- 649 Gilmour, C.C., Riedel, G.S., Ederington, M.C., Bell, J.T., Gill, G.A., Stordal, M.C., 1998. Methylmercury
  650 concentrations and production rates across a trophic gradient in the northern Everglades.
  651 Biogeochemistry 40: 327–345.
- Guédron, S., Point, D., Acha, D., Bouchet, S., Baya, P.A., Tessier, E., Monperrus, M., Molina, C.I., Groleau, A.,
  Chauvaud, L., Thebault, J., Amice, E., Alanoca, L., Duwig, C., Uzu, G., Lazarro, X., Bertrand, A.,
  Bertrand, S., Barbraud, C., Delord, K., Gibon, F.M., Ibanez, C., Flores, M., Fernandez Saavedra, P.,
  Ezpinoza, M.E., Heredia, C., Rocha, F., Zepita, C., Amouroux, D., 2017. Mercury contamination level
  and speciation inventory in Lakes Titicaca & Uru-Uru (Bolivia): Current status and future trends.
  Environ. Pollut. 231: 262-270.
- Hamelin, S., Amyot, M., Barkay, T., Wang, Y., Planas, D., 2011. Methanogens: principal methylators of
   mercury in lake periphyton. Environ Sci Technol 45: 7693–7700.
- Hammerschmidt, C.R., Fitzgerald, W.F., Lamborg, C.H., Balcom, P.H., Visscher, P.T., 2004. Biogeochemistry
  of methylmercury in sediments of long island sound. Mar. Chem. 90: 31–52.
- Hintelmann, H., Wilken, R.D., 1995. Levels of total mercury and methylmercury compounds in sediments of the
   polluted Elbe River: influence of seasonally and spatially varying environmental factors. Sci. Total
   Environ. 166 : 1–10.
- Hsu-Kim, H., Kucharzyk, K.H., Zhang, T., Deshusses, M.A., 2013. Mechanisms regulating mercury
  bioavailability for methylating microorganisms in the aquatic environment: A critical review. Environ.
  Sci. Technol. 47 : 2441–2-456.

- Johnson, N.W., Mitchell, C.P., Engstrom, D.R., Bailey, L.T., Coleman Wasik, J.K., Berndt, M.E., 2016.
  Methylmercury production in a chronically sulfate-impacted sub-boreal wetland. Environ. Sci.:
  Processes & Impacts 18: 725–734.
- Jolivet, C., Augusto, L., Trichet, P., Arrouays, D., 2007. Forest soils in the Gascony Landes Region: formation,
  history, properties and spatial varaibility [WWW Document]. URL http://hdl.handle.net/2042/8480
- Kerin, E.J., Gilmour, C.C., Roden, E., Suzuki, M.T., Coates, J.D., Mason, R.P., 2006. Mercury methylation by
  dissimilatory iron-reducing bacteria. Appl. Environ. Microbiol. 72, 7919–7921.
- Kostka, J.E., Luther, G.W., 1994. Partitioning and speciation of solid phase iron in saltmarsh sediments.
  Geochim. Cosmochim. Acta 58: 1701–1710.
- Lamborg, C.H., Fitzgerald, W.F., Damman, A.W.H., Benoit, J.M., Balcom, P.H., Engstrom, D.R., 2002. Modern
  and historic atmospheric mercury fluxes in both hemispheres: Global and regional mercury cycling
  implications. Global. Biogeochem. Cycles 16: 51-1–51-11
- MacDonald, D.D., Ingersoll C.G., Berger T.A., 2000. Development and evaluation of consensus-based
   sediment quality guidelines for freshwater ecosystems. Arch. Environ. Contam. Toxicol. 39:
   20–31.
- Madison, A.S., Tebo, B.M., Luther, III G.W., 2011. Simultaneous determination of soluble manganese(III),
   manganese(II) and total manganese in natural (pore)waters. Talanta 84: 374–381.
- Mason, R.P., Fitzgerald, W.F., 1996. Sources, Sinks and Biogeochemical Cycling of Mercury in the Ocean. In:
   Baeyens W, Ebinghaus R, Vasiliev O. (eds) Global and Regional Mercury Cycles: Sources, Fluxes and
   Mass Balances, NATO ASI Series. Springer Netherlands, pp 249–272.
- Mason, R.P., Fitzgerald, W.F., Morel, F.M.M., 1994. The biogeochemical cycling of elemental mercury:
  Anthropogenic influences. Geochim. Cosmochim. Acta 58: 3191–3198.
- Matty, J.M., Long, D.T., 1995. Early Diagenesis of Mercury in the Laurentian Great Lakes. J. Great Lakes Res.
  21: 574–586.
- Morel, F.M.M., Kraepiel, A.M.L., Amyot, M., 1998. The Chemical Cycle and Bioaccumulation of Mercury.
  Annu. Rev. Ecol. Syst. 29: 543–566.
- Myrbo, A., Swain, E.B., Johnson, N.W., Engstrom, D.R., Pastor, J., Dewey, B., Monson, P., Brenner, J.,
  Dykhuizen Shore, M., Peters, E.B., 2017. Increase in nutrients, mercury, and methylmercury as a
  consequence of elevated sulfate reduction to sulfide in experimental wetland mesocosms. J. Geophys.
  Res.: Biogeosciences 122: 2769–2785.
- Orem, W., Fitz, H.C., Krabbenhoft, D., Tate, M., Gilmour, C., Shafer, M., 2014. Modeling sulfate transport and
   distribution and methylmercury production associated with Aquifer Storage and Recovery
   implementation in the Everglades Protection Area. Sustainability of Water Quality and Ecology 3-4:
   33–46.
- Oremland, R.S., Culbertson, C.W., Winfrey, M.R., 1991. Methylmercury decomposition in sediments and
   bacterial cultures: involvement of methanogens and sulfate reducers in oxidative demethylation
   (English). Appl. Environ. Microbiol. 57:130–137.
- Pak, K.R., Bartha, R., 1998. Mercury methylation and demethylation in anoxic lake sediments and by strictly
  anaerobic bacteria. Appl. Environ. Microbiol. 64(3): 1013–1017.
- 707 Parks, J.M., Johs, A., Podar, M., Bridou, R., Hurt, R.A., Smith, S.D., Tomanicek, S.J., Qian, Y., Brown, S.D.,

- Brandt, C.C., Palumbo, A.V., Smith, J.C., Wall, J.D., Elias, D.A., Liang, L., 2013. The genetic basis for
  bacterial mercury methylation. Science 339, 1332–1335.
- Pirrone, N., Cinnirella, S., Feng, X., Finkelman, R., Friedli, H., Leaner, J., Mason, R., Mukherjee, A., Stracher,
  G., Streets, D., Telmer, K., 2010. Global mercury emissions to the atmosphere from anthropogenic and
  natural sources. Atmos. Chem. Phys. 10 : 5951–5964.
- Podar, M., Gilmour, C.C., Brandt, C.C., Soren, A., Brown, S.D., Crable, B.R., Palumbo, A.V., Somenahally,
  A.C., Elias, D.A., 2015. Global prevalence and distribution of genes and microorganisms involved in
  mercury methylation. Sci. Adv. 1(9):e1500675.
- Ramlal, P.S., Kelly, C.A., Rudd, J.W.M., Furutani, A., 1993. Sites of Methyl Mercury Production in Remote
  Canadian Shield. Can. J. Fish. Aquat. Sci. 50 : 972–979.
- Renedo, R., Bustamante, P., Tessier, E., Pedrero, Z., Cherel, Y., Amouroux, D., 2017. Assessment of mercury
   speciation in feathers using species-specific isotope dilution analysis. Talanta 174 : 100–110.
- 720 Rodier, J., 1976. L'analyse de l'eau, eaux naturelles, eaux résiduaires, eau de mer. Dunod, Paris.
- Rodríguez Martín-Doimeadios, R.C., Tessier, E., Amouroux, D., Guyoneaud, R., Duran, R., Caumette, P.,
   Donard, O.F.X., 2004. Mercury methylation/demethylation and volatilization pathways in estuarine
   sediment slurries using species specific enriched stable isotopes. Mar. Chem. 90: 107-123.
- Rolfhus, K.R., Sakamoto, H.E., Cleckner, L.B., Stoor, R.W., Babiarz, C.L., Back, R.C., Manolopoulos, H.,
  Hurley, J.P., 2003. Distribution and fluxes of total and methylmercury in Lake Superior. Environ. Sci.
  Technol. 37, 865e872.
- Roustan, Y., Bocquet, M., Musson-Genon, L., Sportisse, B., 2006. Modélisation du mercure, du plomb et du
   cadmium à l'échelle européenne. Pollution Atmosphérique 191: 317–326.
- Schäfer, J., Castelle, S., Blanc, G., Dabrin, A., Masson, M., Lanceleur, L., Bossy, C., 2010. Mercury methylation
  in the sediments of a macrotidal estuary (Gironde Estuary, south-west France). Estuar. Coast. Shelf Sci.
  90: 80–92.
- Schroeder, W.H., Munthe, J., 1998. Atmospheric Transport, Chemistry and Deposition of MercuryAtmospheric
   mercury-An overview. Atmos. Environ. 32: 809–822.
- 734 Stookey, L.L., 1970. Ferrozine-a new spectrophotometric reagent for iron. Anal Chem 42: 779–781.
- Streets, D.G., Horowitz, H.M., Jacob, D.J., Lu, Z., Levin, L., Ter Schure, A.F.H., Sunderland, E.M. 2017. Total
   Mercury Released to the Environment by Human Activities, Environ. Sci. Technol. 51: 5969-5977.
- 737 Stumm, W., Morgan, J.J., 1996. Aquatic chemistry, 3rd ed. John Wiley & Sons.
- United Nations Environment Programme (UNEP), 2013. Global Mercury Assessment 2013: Sources, Emissions,
   Releases and Environmental Transport. Geneva.
- 740 Urban, N.R., Ernst, K., Bernasconi, S., 1999. Addition of sulfur to organic matter during early diagenesis of lake
  741 sediments. Geochim. Cosmochim. Acta 63: 837–853.
- Watras, C.J., Bloom, N.S., Claas, S.A., Morrison, K.A., Gilmour, C.C., Craig, S.R., 1995. Methylmercury
  production in the anoxic hypolimnion of a dimictic seepage lake. Water, Air, & Soil Pollut. 80(1-4):
  735-745.
- 745 Wedepohl, K.H., 1995. The composition of the continental crust. Geochim. Cosmochim. Acta 59: 1217–1232.
- 746
- 747

# 748 Figure captions

Fig 1 Location map of Aquitaine coastal lakes and drainage basin land use. Small rings: Location ofriver sampling for sulphate measurements; black star: location of soil sampling.

Fig 2 Sediment mapping of Aquitaine lakes. Black dots and stars show the location of grab and core samples in which Hg has been measured, respectively. Red dots are additional grabs that have been collected to validate the sediment mapping based on bathymetry. The range of total carbon (TC) concentrations and particulate total mercury (HgTp) levels for each sediment class is indicated in the legend.

Fig 3 Average grain size distribution of lake sandy sediment and catchment soils (right panel), and of lake organic deposits after organic matter removal with  $H_2O_2$  (left panel).

Fig 4 Mean concentration of total carbon (TC), total sulphur (TS) and total Hg (HgTp) in organic
deposits (white bars) and in sandy sediments and catchment soils (grey bars) from the four Aquitaine
lakes. Error bars are standard deviations. Number of samples is indicated.

Fig 5 Mean sulphate concentrations in streams that feed the lakes. Error bars are standard deviations.
 Number of samples indicated in brackets. See Figure 1 for sampling location.

Fig 6 Total Hg (HgTp) concentrations vs total sulphur (TS) concentrations in sediment grabs collectedin Aquitaine lakes.

Fig 7 Relationship between the proportion (%) of MeHg relative to HgTp in organic sediments and
 average sulphate concentrations in lake water column. Error bars are standard deviations (N=5 for
 %MeHg, and N=15 for sulphate)

- 768
- 769
- 770
- 771
- 772

773

774

775

- 776
- 777
- 778
- 779
- 780
- 781
- 782
- 783
- 784

# 

Lake	Area	Maximum depth	Volume	Water residence time	Elevation	Catchment area
	(km <sup>2</sup> )	( <b>m</b> )	$(10^6 \text{ m}^3)$	(year)	( <b>m</b> )	(km <sup>2</sup> )
Carcans-Hourtin	62	9	210	1.8	15	302
Lacanau	20	7	53	0.4	14	310
Cazaux-Sanguinet	58	23	498	4.3	21	200
Parentis-Biscarrosse	35	20.5	252	1	20	303

# **Table 1** Morpho-dynamics characteristics of Aquitaine lake.

**Table 2** Average mercury concentrations (mean total Hg,  $\mu$ g kg<sup>-1</sup> dry weight) in muscle of adult Perch794(length 18-23 cm), Pike (>50 cm), and Zander (>50 cm) in the four lakes. Value ± standard deviation;795N: number of samples for different analysis.

	Carcans-Hourtin	Lacanau	Cazaux-Sanguinet	Parentis-Biscarrosse
Perch (Perca fluviatilis)	1811±281 (N=6)	876±217 (N=7)	379±75 (N=16)	337±209 (N=4)
Pike (Esox spp.)	3151±1207 (N=8)	1608±752 (N=12)	1725±544 (N=7)	1459±333 (N=5)
Zander (Sander lucioperca)	6238 (N=1)	4809 (N=1)	3563±913 (N=6)	1642±485 (N=3)

801	Table 3 Total mercury (HgTp) and methyl mercury concentrations and methyl mercury fraction
802	relative to HgTp of lake sediments and of SW France coastal sediments, and values reported in the
803	literature.

Sediments	$[HgT_p] (\mu g kg^{-1})$	[MMHg <sub>p</sub> ] (µg kg <sup>-1</sup> )	MMHg/HgTp(%)	References
Aquitaine lakes	100-420	1.1-5.1	0.5-2.5	This study
Arcachon Lagoon (SW France)	~ 200	~ 1	0.5	Bouchet et al., 2013
Gironde estuary (SW France)	~ 216	~ 0.7	0.3	Castelle, 2008
Everglades lakes (Florida)	50-400	<0.1-5	<0.2-2	Gilmour et al., 1998
Murray Lake (Papoua-New	~ 111	~ 0.9	0.8	Bowles et al., 2001
Guinea)				
St Lawrence River	~ 1500	~ 2.5	0.2	Avramescu et al., 2011
Lake Superior	83 ±12	$0.21 \pm 0.03$	<1	Rolfhus et al., 2003
Lake Titicaca	45-114	0.51-1.36	1.8±1.5	Guedron et al., 2017
Western North America lakes	29.3±6.5	0.55±0.05	0.9±0.1	Fleck et al., 2016
data base				















