

Environmental fate of chlordecone in coastal habitats: recent studies conducted in Guadeloupe and Martinique (Lesser Antilles)

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Environmental fate of chlordecone in coastal habitats: recent studies conducted in Guadeloupe and Martinique (Lesser Antilles).

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Abstract

- The organochlorine pollution by chlordecone, an insecticide spread in the past in banana plantations, is now recognized as a major ecological, economic and social crisis in Guadeloupe and Martinique Islands. Due to its physical and chemical properties, this molecule is particularly persistent in the natural environment. Volcanic soil of Guadeloupe and Martinique contain allophanes (amorphous clays), which favor chlordecone trapping due to their structure and physical properties. Thus, with this trapping ability, allophanes could constitute a vector allowing chlordecone to contaminate runoff waters, and finally the sea. In the present publication, several studies recently conducted in the Lesser Antilles have been compiled in order to evaluate the desorption of chlordecone from allophanes when arriving in the estuarine environment and to determine the transfer of chlordecone along marine trophic food webs. The experiments showed that 20% of the initial quantity of chlordecone was released from allophones in estuarine conditions and 10% in marine environment. These results could explain the high level of contamination found in the suspended organic matter and zooplankton in the coastal areas located downstream the contaminated watersheds. The contamination of the marine food webs of mangroves seagrass beds and coral reefs, is dominated by a contamination 'by bath' in littoral waters containing chlordecone and by bioamplification seawards.
- 34 **Keywords**: organochlorine pollution, desorption, allophane, zooplankton, trophic food webs, bioamplification

Introduction

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37 The organochlorine pollution by chlordecone, an insecticide spread in the past in banana plantations, is now 38 recognized as a major ecological, economic and social crisis in the French West Indies. Used between 1972 and 39 1993 in fields of Guadeloupe and Martinique, chlordecone (commercialized as Kepone® then Curlone®) has 40 been globally banned by the Stockholm Convention on Persistent Pollutant since 2009. This molecule is 41 responsible for toxicological consequences such as an increased risk of prostate cancer, motor skills delays, an 42 increased risk of preterm birth, endocrine disruptions and reproduction impairments (Epstein 1978; Multigner et 43 al. 2010; Boucher et al. 2013; Kadhel et al. 2014; Multigner et al. 2016). 44 Due to its physical and chemical properties, this hydrophobic molecule has an affinity with soil organic matter 45 (partition coefficient *Koc* = 2500 l.kg⁻¹, Howard *et al.* 1981). Chlordecone has a low volatility (vapour pressure = 2.5 x 10⁻⁵ mm Hg), presents high thermodynamic stability and resistance to chemical or biological degradation, 46 47 involving a high persistence in the environment (Cabidoche et al. 2009). Consequently, it remains in the soil 48 before degradation into at least 5b-hydrochlordecone (Devault et al. 2016). The time to reach the depollution level (at the threshold of quantification of 10 µg.kg⁻¹) depends on the nature of the soils: a few decades for 49 50 nitisol, two to three centuries for ferralsol, five to six for andosol (Cabidoche et al. 2009). 51 Andosols contain amorphous clays, called allophanes, issued from the transformation of volcanic materials with 52 very specific properties. Allophanes present drastically different structures and physical properties compared to 53 usual clays, that is a very high poral volume and an important pores surface. Hygroscopic water contributed to 54 allophanes spherical form, leading them to stack in a three-dimensional fractal labyrinth in which chlordecone 55 could be trapped (Woignier et al. 2007). Due to its trapping ability, allophanes allow chlordecone to reaches 56 runoff and ground waters, and finally end up in the sea. Nitisols are tropical and subtropical deep (iron oxides 57 dominate), red, well-drained soils with clay content of more than 30%. Clays are more aged than andosols, i.e. 58 water content is less important and clays present flatten structure compared to andosols. Nitisols are well-drained 59 type of ferralsol, which are commonly low-draining and muddy due to the high clay content, and more aged than 60 andosols. Andosols and nitisols are particularly abundant in the south of Basse Terre (Guadeloupe) and in the 61 north of Martinique, in the areas where banana plantations occur (Sierra and Desfontaines 2018). 62 Indeed, in Guadeloupe, approximately 11,400 ha present a high risk of contamination that is 90% of probability 63 to find chlordecone in these areas. This area represents 25% of the land surface used for agriculture. In 64 Martinique, this value reaches 40% of agricultural lands (Direction de l'Alimentation, de l'Agriculture et de la 65 Forêt, data 2018). 66 Since 2003, several studies conducted in the Lesser Antilles have highlighted the presence of a contamination by 67 chlordecone in soil (Cabidoche et al. 2009), vegetables (Clostre et al. 2015), aquatic and marine organisms (Coat 68 et al. 2006; Bertrand et al. 2013; Coat et al. 2011; Bouchon et al. 2016; Dromard et al. 2016; Monti et al. 2016, 69 Méndez-Fernandez et al. 2018). However, few studies have been conducted on the fate of chlordecone between 70 agricultural fields and marine ecosystems, especially when it arrives in estuarine environments. The studies on 71 Kepone cycling in aquatic environments has been principally conducted in Virginia, because the original 72 production of Kepone began in Hopewell. In 1975, it was discovered that the Kepone factory had not only 73 exposed workers, but also severely contaminated the James River estuary (Nichols 1990; Luellen et al. 2006).

Chlordecone desorption in the James River estuary has been studied by Nichols (1990) who showed that chlordecone stayed sorbed to organic particles when arriving in estuarine environment in the range of pH (7-8) and salinity (0.006 to 19.5). To our knowledge, chlordecone desorption has never been studied in the Lesser Antilles that present a particular context due to their tropical pedoclimatic conditions. Our first hypothesis is that chlordecone could be trapped by allophanes, due to their physical structure, and stayed sorbed during its transfer to the marine environment.

Along the James River and at the mouth of this river, contamination of aquatic fauna by chlordecone has been highlighted, starting with plankton, the first link in the trophic food web. Zooplankton is broadly highly impacted by this organochlorine pollution (Bahner *et al.* 1977; Luellen *et al.* 2006). Our second hypothesis is that zooplankton could represent a major way of transfer between chlordecone from the water column (adsorbed on terrestrial particles or dissolved) and others organisms from the trophic food-web.

Finally, some studies demonstrated that chlordecone passes through the different levels of the food chain and shows accumulation phenomena, like many other organochlorine pollutants (Bahner *et al.* 1977). In Guadeloupe and Martinique, the degree of contamination of marine organisms depends mostly on their location around the two islands (Dromard *et al.* 2016). Fish and crustacean are generally more contaminated when they are located downstream the contaminated watersheds and when they are close to the coast (Dromard *et al.* 2017). At the same time, detrivorous and carnivorous organisms generally display higher levels of chlordecone contamination than other trophic groups (Luellen *et al.* 2006; Dromard *et al.* 2017). Our third hypothesis is that chlordecone concentrates in marine organisms, depending on their location (close or far from the source of pollution) and on their feeding ecology.

The principal objective of the present study is to compile the results of recent studies on the environmental fate of chlordecone from estuaries to the marine ecosystems in the Lesser Antilles, with two main objectives: 1) to study chlordecone desorption as its arrival in the marine environment, 2) to evaluate the transfer of chlordecone in marine trophic food web, from plants to top predators. To do so, three studies recently conducted in Guadeloupe and Martinique are presented in the present work.

Material and Methods

Experimental protocol to measure chlordecone desorption on allophanes

A selected allophanic soil (6.1% w.w.) historically contaminated was sampled in Martinique and sieved on 2 mm-mesh stainless steel sieve. Samples of 1.5 g of soil has been incubated at dark and moderately stirred in glass vial during 24h in artificial sea salted water at 0, 10, 20 and 35 psu. Artificial seawater results from a mixture of Vittel® mineral water and coral reef "Instant ocean" sea salt from Aquarium Systems®. Each experiment was performed in triplicates.

In order to estimate chlordecone adsorption on the glass of the vials, three positive controls were implemented with Vittel water mixed with chlordecone up to a concentration of 0.1 µg.l⁻¹.

After incubation, suspended soils were filtered on Solid Phase Micro-Extraction (SPME) glass fiber filter (PDMS 100 μm Merlin): soil was lyophilized, then extracted in a microwave extractor (4 ml of dichloromethane, 30 watts, 10 minutes) and filtered again on a glass fiber filter in order to collect the organic extract.

The latter was partially evaporated under 70 % agitation and 51 % vacuum (900 mb), using a Rapidvap provided by Labocongo®, then reconditioned and fully evaporated under nitrogen stream. Extracts were reconstituted in 1 ml of acetonitrile and kept frozen until analysis.

Internal ¹³C chlordecone was added to filtered water (about 150 ml) then liquid-liquid extracted with 3 x 10 ml of dichloromethane. Extracts, dried on Na₂SO₄, were partially evaporated under 70 % agitation and 51 % vacuum (900 mb) then reconditioned and fully evaporated under nitrogen stream. Extracts were reconstituted in

1 ml of acetonitrile and kept frozen until analysis. Extraction recovery was respectively of 106 ± 6 % and 111 %.

For this experiment, solvents (dichloromethane, acetonitrile) were at least of analytical grade and provided by ICS (Belin-Beliet®, France). Certified solid standards for chlordecone were obtained from Cluzeau Info Labo.

Concentrations of chlordecone in water and soil were measured by liquid chromatography (Agilent series 1200)

using in tandem a mass spectrometer detector (Agilent® 6410a) that was provided by Agilent (Santa Clara,

California, USA).

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Samplings protocol to evaluate zooplankton contamination

Samples of zooplankton and seawater were collected at the mouth of two rivers in Guadeloupe (Rivière Grande Anse and Rivière du Grand Carbet) and two rivers in Martinique (Rivière Monsieur and Rivière Rouge) in December 2010. A second sampling campaign was done in Guadeloupe in May 2011. These four rivers have been chosen due to their high level of contamination by chlordecone. Samplings were conducted along three transects oriented from the coast to the open sea, using a 500 µm-meshed net for the plankton and glass bottles for seawater. On each site, nine stations were chosen to sample zooplankton and seawater (n = 3 replicates at each station for each compartment). Samples were freeze-dried before analysis. Quantification of chlordecone in zooplankton and seawater was conducted by the Center for Analytical Research and Technology at Liege University (CART, Belgium). Lipids and chlordecone were extracted with an accelerated solvent extractor using n-hexane-dichloromethane (90:10; v:v). The extracts were then dried under nitrogen flow until a constant weight is obtained. 3 ml n-hexane and a surrogate marker (PCB congener 112), at a concentration of 50 pg.µl⁻¹, were added. Then, the extracts underwent an acid clean-up carried out with 2 ml of sulphuric acid (98-100 %) in order to eliminate organic compounds (lipids, lipoproteins,...). Finally, the eluates were evaporated under an almost dry nitrogen stream. 100 ul n-hexane and 100 ul PCB 209 congener, used as an internal standard at a concentration of 50 pg.µl⁻¹, were added to the samples before injection. The analyses were performed by GCMS-MS (ThermoQuest ITQ 1100 ion trap). With this method, the lower limit of quantification (LOQ) was 0.010 µg.kg⁻¹ w.w. for zooplankton and 0.010 µg.l⁻¹ for seawater (Monti et al. 2017).

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Samplings protocol to evaluate trophic-food web contamination

To study the contamination of marine food webs by chlordecone, samplings were conducted at Petit-Bourg in Guadeloupe, in three habitats: mangrove, seagrass beds (located at 500 m from the coast) and coral reefs (4 km from the coast), between January 2014 and February 2015. On each habitat, sediment, Superficial Sediment Layer (SSL), Suspended Organic Matter (SOM), vegetal matter (macroalgae and seagrass) and consumers from different trophic levels (primary consumers, secondary consumers, top predators) were collected. The classification of species into trophic group was done according to the literature (Randall 1967, Froese and Pauly 2017). The full list of collected species is given in S1 Table. Macroalgae, fishes and crustaceans were collected by hand, spearfishing or using nets in seagrass beds and mangroves. Whenever possible, three replicates were made for each species. Each individual was rinsed, measured (total length (TL) in centimeters) and weighed (w.w. in grams). For each organism, a piece of flesh was collected and frozen (-18°C) until analyses. To sample the suspended organic matter (SOM), seawater was collected in the three habitats in acid-cleaned plastic drums. Water was then filtered on Whatman® GF/F 47 mm filters. Sediments were recovered using a corer to a depth of about 20 cm. Superficial sediment layer (SSL) was sampled on the first 2 cm of sediment. Concentrations in chlordecone were quantified by LABOCEA (Plouzané, France) with liquid chromatography coupled to mass spectrometry in tandem (UPLC-MS/MS). The lower LOQ with this method was 1 µg.kg⁻¹ (w.w.) for organic material and 10 µg.kg⁻¹ (w.w.) for sediment.

162 Results

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Chlordecone desorption on allophanes

No adsorption on the glass of the vials has been observed on controls.

Initial concentration of chlordecone measured in soil was $957 \pm 13.2 \, \mu g.kg^{-1}$ that is about $1.145 \, \mu g$ of chordecone in each sample of $1.5 \, g$ of soil. After 24h of incubation, the maximal desorption of chlordecone was observed in the minimal salinity conditions (0 psu) while the minimal desorption was observed in the maximal salinity condition (35 psu) (Table 1).

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Table 1 Concentrations of chlordecone ([CDC]) measured in soil after 24h of incubation. Initial concentration of chlordecone in soil was $957 \pm 13.2 \ \mu g.kg^{-1}$. SD: Standard Deviation; RSD: Relative Standard Deviation.

		[CDC]	Mean [CDC] ± SD	RSD
Salinity (psu)	Replicates	μg.kg ⁻¹	μg.kg ⁻¹	%
	1	729		
0	2	983	788 ± 173.1	22.0
	3	653		
	1	905		
10	2	733	799 ± 92.6	11.6
	3	759		
	1	821		
20	2	902	837 ± 57.8	6.9
	3	790		
	1	674		
35	2	1 002	814 ± 169.2	20.8
	3	766		

After 24h of incubation, 78 % of the initial quantity of chlordecone was still adsorbed on soil in the freshwater condition (0 psu) and 90 % for the marine condition (35 psu) (Fig. 1). About 20 % of the initial quantity of chlordecone was desorbed from soil in estuarine conditions (10 and 20 psu).

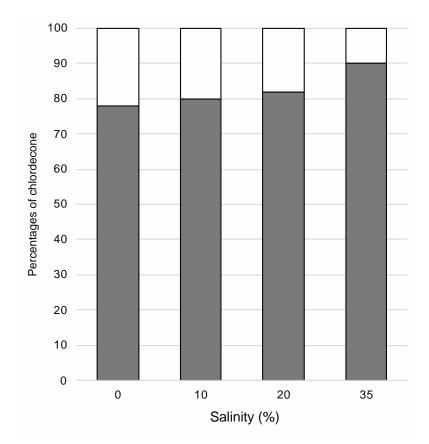


Fig. 1 Percentages of chlordecone in water (white) and in soil (grey) related to salinity after 24h of incubation.

Contamination of the planktonic compartment

In Martinique, the minimal concentration of chlordecone found in zooplankton was 22 μ g.kg⁻¹ (Rivière Rouge) while the maximal concentration was measured at the mouth of Rivière Monsieur (272 μ g.kg⁻¹). In Guadeloupe, concentrations in chlordecone varied between 40 and 306 μ g.kg⁻¹ (Fig. 2). Concentrations of chlordecone in seawater were under the LOQ in 53% of the studied stations. In the other stations, concentrations varied from 0.01 to 0.053 μ g.L⁻¹. No correlation was found between the concentration of chlordecone measured in seawater and zooplankton.

Temporal variations in the level of contamination of zooplankton were tested in Guadeloupe. No statistical significant difference was found on the level of contamination in zooplankton between samples collected at the end of the dry season (May) and those collected at the end of the wet season (December), according to the two sites in Guadeloupe.

Spatial variations in the level of contamination of zooplankton were tested between the four sites (two in Martinique and two in Guadeloupe) sampled in December 2010. Mean concentrations of chlordecone in zooplankton show significant spatial differences (Kruskal-Wallis, $X^2 = 13.9$, p = 0.002). These spatial

differences were globally due to the lower concentrations of chlordecone measured in zooplankton at the mouth of Rivière Rouge compared to the other sites.

The bioconcentration factor (BCFw), calculated as the ratio between concentrations of chlordecone measured in zooplankton and those measured in seawater, varied from 440 and 27200 (Table 2).

Table 2 Mean concentrations of chlordecone (min - max) measured in zooplankton (in $\mu g.kg^{-1}$ w.w.), seawater (in $\mu g.l^{-1}$) and mean bioconcentration factors (BCFw) at the mouth of four rivers. WS: end of the wet season (December), DS: end of the dry season (may), n: number of samples, "-" indicates absence of measures.

Islands	Rivers	Seasons	n	Zooplankton	Seawater	BCFw
Martinique	Rivière Rouge	WS	9	62.3 (22 - 99)	0.028 (0.01 - 0.05)	4320 (440 - 9900)
Martinique	Rivière Monsieur	WS	9	164.3 (79 - 272)	0.01	27 200
Guadeloupe	Grande Anse	WS	9	104 (40 - 294)	0.023 (0.02 - 0.027)	5338 (2095 - 10889)
Guadeloupe	Grande Anse	DS	9	167.8 (79.5 - 271.8)	-	-
Guadeloupe	Grand Carbet	WS	9	133.0 (50.9 - 231.7)	=	-
Guadeloupe	Grand Carbet	DS	9	164.7 (100 - 306)	0.027 (0.017 - 0.053)	7 422 (2509 - 14647)

Mean BCFw was maximal at the mouth of Rivière Monsieur in Martinique. The three other sites reached similar order of magnitude: 4320 at Rivière Rouge, 7422 at Grand Carbet and 5338 at Grande Anse (Fig. 2).

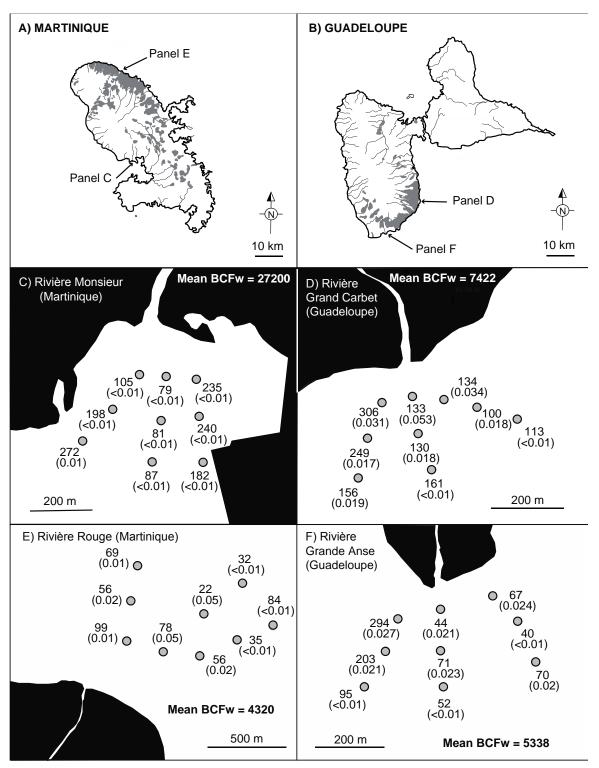


Fig. 2 Mean concentration of chlordecone measured in zooplankton (in μg.kg⁻¹ w.w.) and seawater (into brackets, in μg.l⁻¹ w.w.) in Martinique (A) and in Guadeloupe (B) at the mouth of four rivers (C to F). BCFw: Bioconcentration factor (data from Monti *et al.* 2012); grey surfaces in A and B panels indicate soils contaminated by chlordecone.

Contamination of the trophic food web

Concentrations in chlordecone were measured in different sources of matter and marine organisms in three marine habitats: mangrove, seagrass beds and coral reefs (Fig. 3).



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SOM

Vegetal

matter

Biofilm

feeders

Susp

НВ

feeders (Invert.)

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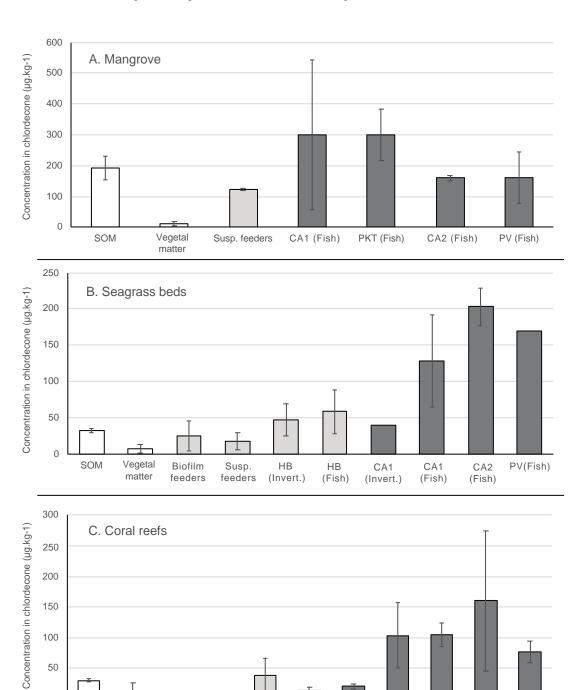


Fig. 3 Mean concentration of chlordecone \pm SD (in $\mu g.kg^{-1}$) measured in sources of carbon (in white, SOM: Suspended Organic Matter; Vegetal matter: algal turf, macroalgae, seagrass), primary consumers (in light grey, Susp. Feeders: suspension feeders, HB: herbivores) and secondary consumers (in dark grey, CA1: invertebrate feeders, PKT: planktivores, CA2: fish and invertebrate feeders, PV: piscivores), in mangrove (A), seagrass beds (B) and coral reefs (C).

ΗВ

(Fish)

(Invert.)

CA1

(Fish)

PKT

(Fish)

CA2

PV(Fish)

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Mean concentrations per trophic category were calculated in pooling all individuals from similar trophic group. Concentrations in chlordecone in sediment and superficial sediment layer (SL) were found under the limit of quantification (10 µg.kg⁻¹). However, chlordecone was found in SOM and vegetal matter in the different habitat. SOM exhibited the highest values among the different sources of carbon. Primary consumers, including biofilm feeders, suspension feeders and herbivorous organisms, presented intermediate mean concentrations of chlordecone from $122.3 \pm 3.8 \,\mu g. \text{Kg}^{-1}$ in mangrove to $17.0 \pm 22.0 \,\mu g. \text{Kg}^{-1}$ in coral reefs. Secondary consumers, that are carnivorous organisms (invertebrates feeders, invertebrates and fish feeders and piscivorous), showed the highest mean concentrations of chlordecone among the studied trophic categories from 232.6 \pm 172.3 $\mu g.Kg^{-1}$ in mangrove to $91.9 \pm 57.6 \,\mu g. \text{Kg}^{-1}$ in coral reefs. In each habitat, significant differences in the mean concentrations of chlordecone were found between the food sources (SOM, vegetal matter) and the trophic categories (primary consumers and secondary consumers) (Table 3). In mangrove, multiple post-hoc comparisons tests indicated that differences of chlordecone concentrations between trophic categories are due to the low value in vegetal matter. In this habitat, SOM, primary and secondary consumers showed similar levels of contamination but were statistically different from the level in vegetal matter. In seagrass beds and coral reefs, the patterns of contamination were similar: SOM, vegetal matter and primary consumers were not statistically different according to their level of contamination. However, these three trophic categories significantly displayed lower concentrations of chlordecone than secondary consumers (Table 3).

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Table 3 Mean concentrations of chlordecone (\pm SD) in μ g.kg⁻¹ measured in food sources (SOM and vegetal matter), primary consumers and secondary consumers. Differences in chlordecone concentrations between the trophic categories were tested with a Kruskal-Wallis test. Letters indicate the results of multiple comparisons conducted with post-hoc tests.

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Trophic categories	Mangrove	Seagrass beds	Coral reefs
SOM	191.3 ± 38.5 ^b	31.7 ± 2.9^{a}	30.3 ± 2.1 ^a
Vegetal matter	12.3 ± 6.4 ^a	6.8 ± 5.6^{a}	12.3 ± 13.5 ^a
Primary consumers	$122.3 \pm 3.79^{\ b}$	34.6 ± 25.9 ^a	17.0 ± 22.0^{a}
Secondary consumers	$232.6 \pm 172.3^{\ b}$	$147.5 \pm 66.0^{\ b}$	$91.9 \pm 57.6^{\ b}$
\mathbf{X}^2	15.1	44.1	35.6
p values	0.002	< 0.0001	< 0.0001

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Discussion

In the Lesser Antilles, all the environmental compartments that have been in contact with chlordecone present a signal of contamination: agricultural soil, rivers, seawater, zooplankton and fauna from the marine food webs. In the present work, the experiments conducted to study the behavior of chlordecone during its arrival in estuarine and marine environments show that molecules stay strongly linked to the particles of terrestrial organic matter, whatever the salinity (0 to 35 psu). About 20 % of the initial quantity of chlordecone was desorbed from soil in estuarine conditions (10 and 20 psu) and 10 % was released in marine conditions (35 psu). Thus, even if furthers

investigations should be conducted to verify this affirmation, marine conditions seem to be the least conducive to desorption. These results are in accordance with those of Nichols (1990), who demonstrated that once chlordecone is sorbed it stays adsorbed in the range of pH (7-8) and salinity (0.006 to 19.5). Bakir et al. (2014) demonstrated that salinity generally does not affect desorption rates of persistent organic pollutants (POP), however their study was conducted on sorption of POP on microplastics. The capacity of POP sorption and desorption on allophanes could be different. In the present study, salinity appears to limit chlordecone desorption because of a process which needs furthers investigations. The present experiments could explain the high concentrations of chlordecone measured in the suspended organic matter (SOM), collected after filtration of seawater in the coastal areas (from 28 to 233 µg.kg⁻¹). Chlordecone was not encountered in superficial sediment layers and marine sediments cores (all samples presented concentrations inferior to the limit of quantification: 10 μg.kg⁻¹). These results could be due to the fact that some samples of sediments collected in the present study were poor in organic matter (for example, less of 2 % of OM in sediments collected in coral reefs). Indeed, the level of contamination of sediments is generally associated with the nature of sediment: organic-rich sediment has much greater capacity to sorb chlordecone than organic-poor sand or kaolinite (O'Connor and Connolly 1980). Bodiguel et al. (2011) and Robert (2012) also found a very low level of contamination for these two compartments in a bay in Martinique (22 samples, all were < 0.5 µg.kg⁻¹). In 2005, Bocquené and Franco found a disparity between the concentrations of chlordecone measured in sediment (<10 µg.kg⁻¹) and SOM (52 and 22 μg.kg⁻¹). Chlordecone can be transferred to estuaries and the marine environment in particulate form (Crabit et al. 2016) via SOM to which it is firmly attached. This suspended matter does not seem to settle on the bottom directly when it arrives at sea, since the sediment compartment is very little affected by the contamination. Several hypothesis could explain these observations: 1) contaminated SOM is constituted by very fine and low density particles which may settle far from estuarine and coastal areas (but this explanation cannot be plausible in semi-enclosed bays), 2) contaminated SOM may be directly consumed in the water column by planktonic compartment upon arrival in the marine environment, 3) contaminated SOM may settle in estuaries and coastal areas but may be rapidly degraded by microorganisms in the sedimentary compartment, 4) samples were collected during the wet season, which can induce intense rainfall and disturbances of estuarine and coastal waters, preventing sedimentation of contaminated SOM (Eggleton and Thomas 2004; Noegrohati et al. 2008). Levels of contamination of zooplankton compartment were investigated to verify the second hypothesis on chlordecone pathway (see above). Zooplankton is closely linked and in direct contact to SOM in estuarine and coastal waters. Indeed, previous studies have highlighted the sensibility to this compartment towards organochlorine pollutants, probably due to its lipid content (Jordan 1979; Day 1990; Nichols 1990). Nichols (1990) measured a mean concentration equal to 4 800 µg.kg⁻¹ in zooplankton from the James River estuary. Coat et al. (2011) reported a mean concentration equal to 5 100 μg.kg⁻¹ in the river mouth and 3 500 μg.kg⁻¹ in the coastal waters of Guadeloupe. In the present study, concentrations of chlordecone measured in zooplankton were considerably lower and varied between 22 and 306 µg.kg⁻¹. Concentrations of chlordecone in zooplankton also spatially varied. The lowest concentrations were observed in front of Rivière Rouge, probably due to the exposure of the site to swell on this Atlantic coast. No temporal variations of chlordecone concentrations were observed between zooplankton collected at the end of the wet season and those collected at the end of the dry season. However, the number of samples was relatively low and samples were taken in the same day during each campaign. Because the planktonic compartment displays very short turn-over, further sampling effort should

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highlight temporal variations in the level of chlordecone contamination of zooplankton. Bioconcentration factors (BCFw) were calculated as the ratio between concentrations of chlordecone in zooplankton and those in seawater. The highest BCFw was found at the mouth of Rivière Monsieur, probably due to its high degree of confinement (semi-enclosed bay). Zooplankton is a keystone component of both marine and rivers systems. It represents a link between invertebrates and fish by providing food and by recycling essential nutrients through feeding on living and detrital material. The accumulation of organochlorine pollutants in zooplankton is a severe threat to ecosystems and a potential way of transfer of the molecule along marine and freshwater food webs. Bioaccumulation phenomena of chlordecone have been demonstrated experimentally (Bahner 1977) and in the field (Dromard et al. 2018). Two processes of bioaccumulation were demonstrated in the present study. In mangrove, the ambient level of contamination is high (SOM: 191.3 µg.kg⁻¹) and this value from the basis of the trophic food web is not statistically different from the other trophic categories (excepted with vegetal matter). Indeed, in mangrove, the entire trophic food web exhibits high concentrations of chlordecone, reflecting the contamination of the baseline. This first way of contamination indicates a prevalence of a contamination "by bath", i.e. by contact between organisms (via teguments and gills) and contaminated surrounding waters. In seagrass beds and coral reefs, food sources and primary consumers shared similar level of contamination that were significantly different from those of the secondary consumers. In these two habitats, a phenomenon of bioamplification, i.e. a contamination by trophic way, is highlighted. The introduction of carnivory in fish diet is linked to a significant increase in the chlordecone concentrations of marine organisms, probably due to the higher lipid content in the preys. In the Lesser Antilles, the degree of contamination of marine systems by chlordecone was drastically lower than that measured in the James River estuary, due to the difference of pollution discharge between the two sites. However, the majority of the studied species were impacted by chlordecone and showed concentrations above the maximal residue limit (LMR) authorized by the French food and safety authorities for the consumption and the commercialization of seafood products (LMR = $20 \mu g.kg^{-1}$). The variations among species from similar trophic category were sometimes very high, suggesting that other processes could influence the level of contamination of marine fishes, such as their physiology, their size or their movement among the different habitats. Indeed, Luellen et al. (2006) indicated that fish ecology could considerably influence the level of contamination of marine fishes, especially for migratory species that spend a period of the year in an estuarine environment. To conclude, molecules of chlordecone can reach marine ecosystems with two principal processes. Firstly, dissolved molecules of chlordecone can infiltrate ground waters during percolation process and join marine environment with resurgences inside rivers or in the sea (Crabit et al. 2016). In the present study, this pollution was highlighted by the concentrations of chlordecone measured in seawater samples. Secondly, due to its strong affinity with soil particle (allophane or others terrestrial particles), molecules of chlordecone can reach runoff waters, rivers and estuaries by leaching of the contaminated soil throughout the erosion process of the catchment (Crabit et al. 2016). When chlordecone arrive in marine environment, it seems to stay sorbed on allophanes and this fact could be similar with other organic compounds, as salinity generally does not affect desorption rates (Bakir et al. 2014). Chlordecone, adsorbed or dissolved, is then integrated in SOM and zooplankton. These two compartments are located at the base of trophic food-web and contribute to the contamination of the entire food chain, via contact or trophic way.

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344	In perspectives, several aspects of chlordecone transfer between terrestrial, rivers and marine ecosystems should
345	be developed in future studies. The low proportion of chlordecone desorbed from allophanes requires further
346	studies on the physical and chemical behavior of allophanes when arriving in marine environments. The
347	potential ways of chlordecone degradation in marine sediment could also be investigated in order to understand
348	the low concentrations of chlordecone found in marine sediment. Measurements of chlordecone metabolites in
349	the sediment could help with the understanding of a potential bacterial degradation in this compartment. Finally,
350	studies on the kinetic of contamination versus decontamination in marine fauna could be useful to explain the
351	high variations in the concentrations of chlordecone between species and individuals.
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358	References
359 360	Bahner LH, Wilson AJ, Sheppard JM, Patrick JM, Goodman LR, Walsh GE (1977) Kepone® Bioconcentration, accumulation, loss and transfer through estuarine food chains. Chesap Sci 18: 299-308
361 362	Bakir A, Rowland SJ, Thompson RC (2014) Transport of persistent organic pollutants by microplastics in estuarine conditions. Estuar Coast Mar Sci 140: 14-21
363 364 365	Bertrand JA, Guyader O, Reynal L (2013) Characterization of the contamination of the halieutic fauna by chlordecone around Guadeloupe (surveys 2008 to 2011). Ifremer, 39 pp (in French) http://archimer.ifremer.fr/doc/00136/24762
366 367 368	Bodiguel X, Bertrand JA, Frémery J (2011) Transfert of the chlordecone in the trophic foodwebs of commercial marine species in the Lesser Antilles (Chloretro). Ifremer, 46 pp (in French) http://archimer.ifremer.fr/doc/00036/14684/
369 370	Bocquené G, Franco A (2005) Pesticide contamination of the coastline of Martinique. Marine Pollut Bull 21: 9511-9521
371 372 373	Boucher O, Simard MN, Muckle G, Rouget F, Kadhel P, Bataille H, Chajès V, Dallaire R, Monfort C, thomé JP, Multigner L, Cordier S (2013) Exposure to an organochlorine pesticide (chlordecone) and development of 18-month-old infants. NeuroToxicology 35: 162-168
374 375 376	Bouchon C, Lemoine S, Dromard CR, Bouchon-Navaro Y (2016) Level of contamination by metallic trace elements and organic molecules in the seagrass beds of Guadeloupe Island. Environ Sci Pollut Res 23: 61-72
377 378 379	Cabidoche YM, Achard R, Cattan P, Clermont-Dauphin C, Massat F, Sansoulet J (2009) Long-term pollution by chlordecone of tropical volcanic soils in the French West Indies: A simple leaching model accounts for current residue. Environ Pollut 157: 1697-1705
380 381	Clostre F, Letourmy P, Lesueur-Jannoyer M (2015) Organochlorine (chlordecone) uptake by root vegetables. Chemosphere 118: 96-102
382 383	Coat S, Bocquené G, Godard E (2006) Contamination of some aquatic species with the organochlorine pesticide chlordecone in Martinique. Aquat Living Resour 19: 181-187

384 385 386	Coat S, Monti D, Legendre P, Bouchon C, Massat F, Lepoint G (2011) Organochlorine pollution in tropical rivers (Guadeloupe): role of ecological factors in food web bioaccumulation. Environ Pollut 159: 1692-1701
387 388	Crabit A, Cattan P, Colin F, Voltz (2016) Soil and river contamination patterns of chlordecone in a tropical volcanic catchment in the French West Indies (Guadeloupe). Environ Pollut 212: 615-626
389	Day KE (1990) Pesticide residues in freshwater and marine zooplankton: a review. Environ Pollut 67: 205-222
390 391 392	Devault DA, Laplanche C, Pascaline H, Bristeau S, Mouvet C, Macarie H (2016) Natural transformation of chlordecone into 5b-hydrochlordecone in French West Indies soils: statistical evidence for investigating long-term persistence of organic pollutants. Environ Sci Pollut Res 23: 81-97
393 394 395	Dromard CR, Bodiguel X, Lemoine S, Bouchon-Navaro Y, Reynal L, Thouard E, Bouchon C (2016) Assessment of the contamination of marine fauna by chlordecone in Guadeloupe and Martinique (Lesser Antilles). Environ Sci Pollut Res 23: 73-80
396 397 398	Dromard CR, Guéné M, Bouchon-Navaro Y, Lemoine S, Cordonnier S, Bouchon C (2017) Contamination of marine fauna by chlordecone in Guadeloupe: evidence of a seaward decreasing gradient. Environ Sci Pollut Res 25: 14294-14301
399 400 401	Dromard CR, Bouchon-Navaro Y, Cordonnier S, Guéné M, Harmelin-Vivien M, Bouchon C (2018) Different transfer pathways of an organochlorine pesticide across marine tropical food webs assessed with stable isotope analysis. PLoS ONE 13(2): e0191335
402 403	Eggleton J, Thomas KV (2004) A review of factors affecting the release and bioavailability of contaminants during sediment disturbance events. Environ Int 30: 973-980
404	Epstein SS (1978) Kepone – Hazard evaluation. Sci Total Environ 9: 1-62
405	Froese R, Pauly D (2017) FishBase: World Wide Web electronic publication. Available: www.fishbase.org.
406 407	Jordan RA, Goodwin PA, Sutton CE, Lascara VJ, VanVeld PA, Carpenter RK (1979) An evaluation of the Kepone contamination of the plankton of the James River. Virginia Institute of Marine Science, 51 pp
408 409	Kadhel P, Monfort C, Costet N, Rouget F, Thomé JP, Multigner L, Cordier S (2014) Chlordecone exposure, length of gestation, and risk of preterm birth. Am J Epidemiol 179(5): 536-544
410 411	Luellen DR, Vadas GG, Unger MA (2006) Kepone in James River fish: 1976-2002. Sci Total Envion 358: 286-297
412 413 414	Méndez-Fernandez P, Kiska JJ, Heilthaus MR, Beal A, Vandersarren G, Caurant F, Spitz J, Taniguchi S, Montone RC (2018) From banana fields to the deep blue: assessment of chlordecone contamination of oceanic cetaceans on the eastern Caribbean. Mar Poll Bull 137: 56-60
415 416 417 418	Monti D, Forget-Leray J, Lagadic L, Thomé JP, Boulangé-Lecomte C, Zimmerman-Chancerel G, Géraudie P, Louvet M, Vassaux D, Boucher P, Herman F, Renia L, Yang-Ting L, Letellier J, Dromard C, Urvoix L (2012) Vulnerability of Guadeloupe and Martinique freshwater nurseries to chlordecone contamination in water. Report UAG/ONEMA/ODE/OEG/DEAL Martinique/DEAL Guadeloupe, 43 pp (in French)
419 420 421	Monti D, Rey P, Thomé JP (2016) Contamination of freshwater fauna. In: Lesueur Jannoyer M, Cattan P, Woignier T (ed), Clostre Florence (ed.) Crisis management of chronic pollution: contaminated soil and human health. CRC Press, Boca Raton, pp 91-103
422 423	Monti D, Rey P, Thomé JP (2017) Contamination of freshwater fauna. In: Jannoyer M, Cattan P, Woignier T, Clostre F (eds) Crisis Management of Chronic Pollution, 1st edn. CRC Press, Boca Raton, pp 91-103

424 425	Multigner L, Ndong JR, Giusti A, Romana M, Delacroix-Maillard H, Cordier S, Jégou B, Thome JP, Blanchet P (2010) Chlordecone exposure and risk of prostate cancer. Am J Clin Oncol 28: 3457-3462
426 427	Multigner L, Kadhel P, Rouget F, Blanchet P, Cordier S (2016) Chlordecone exposure and adverse effects in French West Indies populations. Environ Sci Pollut Res 23: 3-8
428 429	Nichols MM (1990) Sedimentologic fate and cycling of Kepone in an estuarine system: example from the James River estuary. Sci Total Environ 97/98: 407-440
430 431	Noegrohati S, Narsito, Hadi S, Sanjayadi (2008) Fate and behavior of organochlorine pesticides in the Indonesian tropical climate: a study in the Segara Anakan estuarine ecosystem. Clean 36: 767-774
432 433	O'Connor DJ, Connolly PJ (1980) The effect of concentration of adsorbing solids on the partition coefficient. Water Res 14: 1517-1523
434	Randall JE (1967) Food habits of reef fishes of the West Indies. Stud Trop Oceanogr 5: 665-847
435 436	Robert S (2012) History of chlordecone contamination of coastal sediments in the French West Indies (ChloSed). Ifremer, 93 pp (in French)
437 438	Sierra J, Desfontaines L (2018) Les sols de la Guadeloupe-Génèse, distribution et propriétés. Rapport INRA Guadeloupe, 23 pp (in French)
439 440	Woignier T, Pochet G, Doumenc H, Dieudonné P, Duffours L (2007) Allophane: A natural gel in volcanic soils with interesting environmental properties. J Sol-Gel Sci Technol 41(1): 25-30
441	
442	
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