1

Metabolic Messengers: Endocannabinoids

- 2 Arnau Busquets-Garcia^{1,+}, Juan P. Bolaños^{2,3,4,+}, Giovanni Marsicano^{5,6,+}
- 3 ¹"Cell-type mechanisms in normal and pathological behavior" Research Group. IMIM-
- 4 Hospital del Mar Medical Research Institute, PRBB, Barcelona, Spain.

⁵ ²Institute of Functional Biology and Genomics, University of Salamanca, CSIC, 37007

- 6 Salamanca, Spain
- 7 ³Centro de Investigación Biomédica en Red sobre Fragilidad y Envejecimiento Saludable
- 8 (CIBERFES), Instituto de Salud Carlos III, Madrid, Spain
- 9 ⁴Institute of Biomedical Research of Salamanca, University Hospital of Salamanca, University
- 10 of Salamanca, CSIC, 37007 Salamanca, Spain
- ⁵INSERM, U1215 NeuroCentre Magendie, Bordeaux, France
- ⁶University of Bordeaux, Bordeaux, France
- 13 ⁺ All authors shared first authorship and are corresponding authors (abusquets@imim.es;
- 14 jbolanos@usal.es; giovanni.marsicano@inserm.fr)
- 15

16 Abstract

The endocannabinoid system is considered one of the most important modulatory networks in the whole organism. Research has yielded great insight on the mechanisms that link endocannabinoids and several peripheral and brain metabolic functions. Here, we provide a brief overview of the metabolic roles of endocannabinoids in tissue-, cellular- and subcellular-dependent mechanisms. We highlight past discoveries and future open questions with a special focus on CB1 receptor-dependent astroglial and mitochondrial metabolic processes.

25

1. Introduction

With its almost 5000-year documented use, cannabis sativa is one of the oldest 26 cultivated plants, possibly the first bred for reasons independent from direct food 27 production^{1,2}. We can, however, presume that the first users soon realized that its 28 consumption induces, amongst a plethora of other effects, strong hunger and desire to 29 eat appetitive foods³. After the introduction of the plant into the Western 30 31 pharmacopeia in the 1800s for the treatment of various conditions, modern science, 32 starting from the middle of last century, provided key discoveries in the field (Figure 1). 33 The description of the chemical structure of the active components of the plant (cannabinoids, like the psychoactive Δ^9 -tetrahydrocannabinol, THC, and others)^{4,5}, of 34 the cellular type-1 and type-2 cannabinoid receptors (CB1 and CB2)^{6,7}, and of the 35 endogenous lipid ligands (endocannabinoids)⁸⁻¹⁰ with the machinery for their synthesis 36 and degradation¹¹ (Figure 1) are the historical milestone discoveries leading to the 37 definition of the so-called endocannabinoid system (ECS)¹², which is a fundamental 38 modulatory system in the whole organism. With just a little exaggeration, one could 39 affirm that there is almost nothing that happens in the body, from skin physiology, to 40 41 immune responses, to metabolic and mental processes, that would not involve the 42 activation of endocannabinoid signaling.

As we will see more in detail below, the majority of these diverse functions underlie the important general role of the ECS in modulating body energy metabolism. This short review will address these aspects, particularly focusing on how the modulation of "cellular" metabolism represents a novel way through which endocannabinoids control brain and body functions, and their interactions. This is linked to the discovery of novel cellular and subcellular localizations of CB1 receptors, which can deeply impact cellular metabolic processes with functional implications at the organismal level.

50 The definition of the ECS is continuously changing, with the addition of novel players 51 directly or indirectly involved in its functions. Due to space limitations, the list of 52 cannabinoid receptors and the endogenous molecules modulating their activity 53 provided in this article is necessarily limited and incomplete (See Refs. 2,13-15 for more 54 exhaustive descriptions). CB1 and CB2 are seven-transmembrane G Protein-Coupled 55 Receptors (GPCRs) and they were first discovered as the cellular targets of exogenous plant-derived (THC) or synthetic cannabinoids^{6,7}. This discovery implied the existence of 56 57 endogenous ligands, which were soon identified as lipid derivatives of arachidonic acid and called endo-cannabinoids^{8,9}. The most studied endocannabinoids are 2-58 arachidonoyl-glycerol (2AG) and anandamide (AEA), but new concepts have arisen that 59 should be taken into consideration when dealing with endocannabinoid signaling: (i) 60 2AG and AEA could bind to other targets than cannabinoid receptors¹⁴, implying that 61 their tissue level changes do not always reflect an activation of the ECS in strict sense; 62 (ii) peptide endogenous ligands (PepCans) can bind and modulate the activity of 63 cannabinoid receptors and were proposed to belong to the endocannabinoid family^{16,17}; 64 (iii) the synthetic and catabolic pathways of lipid endocannabinoids are redundant and 65 involve also other lipids, such as prostaglandins and others¹⁸; (iv) non-arachidonic acid-66 derived lipids were identified as endogenous allosteric regulators of cannabinoid 67 68 receptor activity, such as the anti-inflammatory Lipoxin A4 (Ref. 19) or the steroid precursor Pregnenolone²⁰, making these molecules members *de facto* of the ECS. 69

70 71

2. Endocannabinoid signaling, a cornerstone for whole-body energy metabolism

72 Few attempts have been made to obtain a unified vision of the large spectrum of 73 functions controlled by endocannabinoid signaling in animal physiology. One of the 74 most convincing ideas in our opinion is that endogenous CB1 receptor signaling might represent a powerful system to adjust body metabolism towards the accumulation of 75 energy reserves. As such, endocannabinoid signaling would exert a prototypical 76 77 exostatic function (accumulation of reserves for future potential needs), as opposed to endostatic ones, through which the body responds to present needs (see Ref. 21 for an 78 79 exhaustive discussion). Thus, amongst other functions, endocannabinoids can rewire behavior towards more food intake (particularly palatable and energy-rich)²², favor the 80 accumulation of fat in the adipose system²³, increase nutrients absorption in the 81 intestinal tract²⁴, decrease glucose use in the muscles²⁵, increase lipogenesis in the 82 liver²⁶ and increase insulin secretion in the pancreas²⁷ (Figure 2). In line with this, other 83 roles of endocannabinoid signaling such as in the reduction of passive fear responses²⁸, 84 of pain²⁹, of anxiety³⁰, or the gating of motivation for physical exercise³¹⁻³³ would help 85 individuals to be resilient against adversities and to physically engage in the search and 86

consumption of food²¹. Similarly, endocannabinoids seem to enhance sensory
 perception, like vision, olfaction or taste^{34,35}, all conditions that would favor the
 detection of nutrients in the environment²¹. Thus, endocannabinoids play a key role in
 the control of body energy metabolism.

91 The link between endocannabinoid signaling and energy metabolism is based on an overwhelming amount of experimental and clinical data. In the second half of the years 92 2000, the CB1 receptor blocker rimonabant was commercialized as a therapeutic aid 93 against obesity and associated metabolic disorders³⁶. Before being withdrawn in 2008 94 because of undesired side effects linked to anxiety and depression³⁷, rimonabant 95 96 showed its efficacy in reducing not only body weight, but also many other obesityassociated metabolic dysfunctions^{36,38,39}. These aspects have been listed and discussed 97 by several reviews over the years^{3,40,41}. In this section, we will just shortly highlight how 98 endocannabinoid signaling through CB1 receptors can impact central and peripheral 99 100 mechanisms involved in energy metabolism. For space reasons, we will focus on CB1 101 receptor signaling, but it should be considered that the other main cannabinoid receptors (*i.e.*, CB2 receptors) have also been involved in whole-body metabolism^{42,43}. 102

103

104 Neuronal endocannabinoid signaling. CB1 receptors in the brain, especially those 105 located in the hypothalamic areas, play a key role in controlling appetite and modulating certain neuropeptides to favor energy accumulation and storage^{3,44}. In 106 107 other brain regions involved in reward (i.e., ventral tegmental area or nucleus accumbens), the endocannabinoid signaling has also been linked to the control of 108 hedonic feeding behavior^{45,46}. As happen with other brain functions⁴⁷, depending on the 109 brain regions where CB1 receptors are located, the control over food intake or energy 110 balance can be very different. Indeed, this control also depends on whether CB1 111 receptors are located on specific cell types^{22,48}. Thus, specific pools of CB1 receptors 112 present in particular hypothalamic cells, such as the POMC neurons, or in the 113 sympathetic system could modulate food intake^{3,44,48}. Accordingly, different doses of 114 (endo)cannabinoids could preferentially act on glutamatergic or GABAergic CB1 115 receptors to respectively increase or decrease food intake in mice²². Similar biphasic 116 effects of cannabis on food consumption were also observed in humans⁴⁹. 117

118

119 Endocannabinoid signaling in adipose tissue. Adipocytes also express CB1 receptors, which are involved in energy metabolism^{23,41}. Pharmacological CB1 receptor blockade 120 increased adiponectin messenger RNA expression in adipose cells and tissue, 121 respectively^{23,50}. These results suggest that CB1 receptors on adipocytes normally 122 suppress adiponectin activity. In addition, CB1-dependent mechanisms increase 123 lipoprotein lipase activity enhancing the storage of more fat^{41,51,52}. These results, 124 together with the finding that other components of the ECS are found in fat tissue⁵³, 125 126 demonstrate that endocannabinoid signaling can be an important target to modulate fat metabolism. Indeed, analysis of human adipose tissue of obese patients revealed a 127 128 decrease in the activity of the main AEA-degrading enzyme fatty acid amide hydrolase, increased endocannabinoid levels and decreased expression of CB1 receptors⁵⁴. 129 Moreover, CB1 receptor activation results in adipogenesis and lipogenesis⁴¹, which 130 leads to an impaired mitochondrial function in diet-induced obesity⁵⁵. Overall, the 131 132 specific targeting of adipocyte CB1 receptors, which have been recently shown to play a 133 key role in the crosstalk across adipocytes, immune cells and the sympathetic nervous system²³, represents an interesting therapeutic approach in order to treat obesity and 134 135 metabolic syndrome.

136

137 Hepatic endocannabinoid signaling. The deletion of CB1 receptors specifically from 138 mouse hepatocytes has brought important findings supporting the role of endocannabinoid signaling in the liver⁵⁶. High-fat diet exposure of animals lacking 139 hepatic CB1 receptors does not cause dyslipidemia, insulin/leptin resistance, hepatic 140 141 steatosis, which are key metabolic consequences associated with the diet-induced obesity^{57,58} (but see Ref. 59). In line with the general increase of endocannabinoid 142 signaling under obesogenic conditions, high fat diet- or alcohol-induced hepatic 143 steatosis is linked to an upregulation of liver CB1 receptors and the modulation of other 144 components of the ECS⁵⁸. Interestingly, studies on human patients with hepatic 145 steatosis have also detected increases in endocannabinoid levels⁶⁰ and hepatic CB1R 146 expression⁶¹. Moreover, hepatic CB1 receptors contribute to control bile acid 147 metabolism whereas bile acids can control the levels of certain endocannabinoids⁶². 148 149 However, the link between endocannabinoid signaling and bile acid composition and its 150 impact on body weight control is still unknown.

Endocannabinoid signaling in other peripheral sites. All key players of the ECS are 152 present in other peripheral tissues such as the gastrointestinal (GI) tract²⁴, where they 153 regulate motility, acid and fluid release, inflammation, vasodilation, but also food 154 intake⁶³. In addition, endocannabinoids in the GI tract affect the secretion of hormones 155 critically involved in whole body energy metabolism, such as ghrelin, CCK and GLP-1, 156 thereby exerting a crucial role in inter-organ endocrine communication processes⁶³⁻⁶⁵. 157 Neuronal bidirectional communication between the periphery and the brain is also 158 under the control of endocannabinoid signaling, as strong evidence indicates that both 159 160 the sympathetic and parasympathetic systems participate in its impact on metabolism and behavior^{63,66-68}. The ECS is also known for its contribution to blood glucose control, 161 mostly by the direct modulation of endocrine cell types of the pancreatic Langerhans 162 islets²⁷, although the exact mechanisms are not known yet. Finally, peripheral 163 164 endocannabinoid signaling is also present in skeletal muscle, where its activation decreases glucose uptake and oxidative metabolism²⁵. 165

166

151

167

7 **3.** Endocannabinoids and mitochondria.

168 History is an important factor to approach the study of the ECS. The search for the biological mechanisms underlying the effects of the *Cannabis sativa* plant became very 169 strong in the late 1960s and in the 1970s, as depicted by the relatively rapid raise of 170 171 publications during this period (Figure 1). However, this wave of interest was slowly 172 decreasing during the 1980s, likely because the lack of a precise mechanism of action of 173 cannabinoids made the research less attractive. By early 1990s, however, the discovery 174 of cannabinoid receptors and endocannabinoids fueled again a rapidly growing and still ongoing raise of enthusiasm (Figure 1). Interestingly, a series of "pre-receptor" papers 175 during the 1970s showed that plant-derived cannabinoids can alter mitochondrial 176 activity by affecting, for instance, NADH oxidase activity⁶⁹, ATP synthesis⁷⁰ and oxygen 177 consumption⁷¹. With the later discovery of cannabinoid receptors and their description 178 179 as GPCRs, however, these intracellular effects were revised and finally considered as artifacts or unspecific drug effects. Indeed, lipid cannabinoids were shown to alter lipid 180 membrane composition (including mitochondrial ones)⁷², thereby possibly altering the 181

182 activity of organelles in an unspecific manner. As GPCRs have been always known to be "plasma membrane" proteins, the possibility that some of the cannabinoid effects on 183 184 mitochondrial activity might be specifically and directly receptor-mediated was not 185 taken into consideration. To the best of our knowledge, the first functional and anatomical evidence of a receptor-mediated regulation of mitochondrial activity by 186 cannabinoids came in the years 2005-2010, when a series of studies showed the impact 187 of cannabinoids on mitochondrial activity in human sperms⁷³⁻⁷⁵, where the presence of 188 CB1 receptors on mitochondrial membranes was first observed. The specificity of these 189 extremely interesting human data could obviously not be confirmed by the use of 190 191 samples genetically lacking the CB1 receptor. This was done in 2012, when the 192 comparison and rigorous quantification of immunogold electron microscopy staining in 193 the brain of wild-type and CB1-KO mice allowed establishing the specific presence of mitochondria-associated CB1 receptors (mtCB1)⁷⁶. Likewise, the effects of cannabinoids 194 195 on oxygen consumption in brain-isolated mitochondria was shown to be present in wild-type, but not in CB1-KO mice⁷⁶. These data did not exclude that, depending on 196 197 doses and conditions, cannabinoids might also unspecifically alter mitochondrial 198 functions by modifying membrane lipid composition, but they strongly promoted the 199 idea that GPCRs can be active intracellularly and regulate the functions of organelles in a specific way⁷⁷⁻⁸⁰. Importantly, Benard et al also showed that endocannabinoids are 200 present in brain mitochondria and their levels are strikingly inversely proportional to 201 the respiration activity of the organelles⁷⁶, indicating that the CB1-dependent 202 modification of mitochondrial functions is not only a pharmacological effect of 203 204 exogenous cannabinoids, but it represents a physiological feature of the ECS. The functional presence of mtCB1 has been so far shown in different tissues and cell 205 types^{25,73,76,81-84}. For instance, mtCB1 has been associated to the regulation of motility 206 and other functions in sperms^{73-75,85}, muscle mtCB1 can regulate oxygen consumption²⁵, 207 and the presence of mtCB1 in progesterone-producing ovarian cells, recently suggested 208 its implication in peripheral hormonal regulation⁸². However, the brain is so far the 209 organ where most details have been obtained concerning the functions and the 210 mechanisms of action of mtCB1 receptors. Interestingly in the context of the present 211 212 article, the discovery of mtCB1 added a new element to the chain linking 213 (endo)cannabinoid signaling to metabolic processes of the cells. Thus, not only endocannabinoids can regulate body energy metabolism by typical GPCR-like signaling
at cellular plasma membranes, but they can use the modulation of cellular energy
transformation as a signal pathway to control organ functions and, in the brain, to
dictate behavioral choices. So far, amongst other behavioral processes, mtCB1 activity
has been involved in the regulation of food intake^{81,86}, hippocampal memory⁸⁴, social
interactions⁸³, nociception and motor control⁸⁶.

The brain is the most complex organ of the body and, accordingly, its bioenergetic requirements are very high. Many brain cells contribute to bioenergetic roles, but glial cells, and in particular astrocytes, are major effectors of central metabolic functions in mammals. In the next section, we will see how astroglial mtCB1 receptors can regulate behavior by reprogramming brain metabolic activity.

225 **4. Endocannabinoids and astrocytes.**

226 Neurons and astrocytes are metabolically different. For their bioenergetic needs, 227 astrocytes mainly rely on glycolysis⁸⁷⁻⁹⁰, whereas neurons use principally the 228 mitochondrial oxidative phosphorylation (OXPHOS) pathway^{91,92}. Given their glycolytic 229 phenotype, astrocytes are net producers of lactate, a metabolite now considered an 230 energy substrate for neighboring neurons^{93,94}. This astrocyte-neuron intercellular 231 metabolic coupling through lactate actually represents a key element sustaining brain 232 functions⁹⁵ such as long-term memory formation⁹⁶ or depressive-like behaviors⁹⁷.

233 OXPHOS, besides being a major site for ATP generation, actively produces reactive oxygen species (ROS) with physiological signaling properties⁹⁸. Mitochondrial complex I 234 (CI) is an important site for ROS production under physiological conditions⁹⁹ that is 235 amenable to regulation by assembling with complex III (CIII)¹⁰⁰. In astrocytes, CI is 236 237 partially disassembled from CIII, determining low OXPHOS energy efficiency, but high ROS generation under physiological conditions¹⁰⁰. Indeed, downmodulating 238 239 mitochondrial ROS abundance in astrocytes by a genetic approach causes profound metabolic changes, negatively impacting on neighboring neuronal integrity and 240 behavior in mice¹⁰¹. 241

242 Interestingly, recent work proposed a possible connection between mtCB1-mediated inhibition of CI activity in the brain^{76,84} and CI production of ROS in astrocytes⁸³. For ROS 243 formation, CI requires an intact N-module, i.e., the functional domain of the complex 244 that is responsible for electron acceptance from NADH(H+)^{102,103}. Activation of mtCB1 in 245 astrocytes causes the loss of CI N-module stability, without affecting those of the Q- and 246 the P-modules⁸³. Earlier studies revealed that the PKA-dependent phosphorylation of 247 the CI N-module subunit NDUFS4 is required to sustain CI activity¹⁰⁴. Interestingly, 248 activation of mtCB1 causes NDUFS4 dephosphorylation that wholly accounts for the 249 loss of the CI N-module stability and the impairment of mitochondrial ROS formation in 250 astrocytes⁸³. Notably, mitochondrial ROS stabilizes hypoxia-inducible factor-1a 251 (HIF1a)¹⁰⁵, a transcription factor that sustains the expression of glycolytic genes¹⁰⁶. In 252 line with this, mtCB1-induced decrease of ROS causes HIF1a destabilization and 253 debilitates glycolysis in astrocytes, leading to a reduced release of astroglial lactate, 254 eventually causing neuronal bioenergetic dysfunction⁸³. Interestingly, this cascade 255 256 results in reduced mouse social interactions, which can be reverted by exogenous 257 lactate⁸³. Thus, the mtCB1-dependent control of cellular metabolic pathways is 258 translated into key behavioral functions.

259

260 **4. Conclusions and future perspectives**

Metabolism can be defined as "the chemical processes that occur within a living 261 262 organism in order to maintain life". Endocannabinoids are surely important metabolic molecules both at cellular and whole-body levels. As such, they control energy 263 metabolism, even beyond the immediate needs, to "be ready" for future scarcity of 264 energy sources and, thereby to maintain life through "exostatic" mechanisms. This is 265 266 obtained by a myriad of alternative, overlapping or distinct cellular mechanisms that rely on different signaling couplings and/or subcellular locations of the cannabinoid 267 receptors. The continuous identification of new mechanisms and/or new locations will 268 tell us how many cellular processes can be impacted by the endocannabinoid signaling. 269 270 The discovery that endocannabinoids can alter cellular mitochondrial functions represents a key step towards a full understanding of their metabolic impact. 271 272 Moreover, the cumulating evidence that other GPCRs are also functionally present at 273 mitochondrial membranes suggests that the control of bioenergetic processes can be a 274 generalized means of receptor-dependent regulation of cell metabolism and functions. 275 In this context, the fact that typical signaling receptors directly "use" metabolic 276 processes to signal, suggests that the classical separation between metabolism and signaling is possibly less clear cut than previously thought. So far, mtCB1 receptors have 277 been mainly studied as the pharmacological targets of exogenous cannabinoid drugs. 278 279 Interestingly, the data seem to indicate that whereas certain cannabinoid effects (e.g. analgesia) do not require mtCB1 receptors, undesired side-effects of these drugs such 280 281 as decreased social behavior, impairment of memory or alteration of motor responses 282 are mediated by direct mitochondrial alterations. All these observations are necessarily 283 partial, but they suggest that also different physiological functions of endocannabinoids 284 might require or not mitochondrial and metabolic signaling. In the brain, astrocytes are 285 key metabolic cells, whose activity is largely dedicated to the maintenance of energy 286 levels in the organ. The fact that astroglial metabolism is under endocannabinoid 287 control is a further indication of the key metabolic functions of these molecules. 288 However, astrocytes are not only metabolic cells and strongly participate in the 289 signaling necessary for synaptic activity. The recent results indicating that 290 endocannabinoids can control synaptic plasticity by regulating mitochondrial calcium signaling¹⁰⁷ is another clear indication that metabolic processes can be directly used as 291 292 signaling in the brain. In addition, the control of body metabolism is clearly the result of 293 a constant interaction between the periphery and the brain. Endocannabinoid signaling, 294 through its almost ubiquitous presence and its ability to modulate key metabolic, 295 hormonal and neuronal processes, is perfectly placed to control this bidirectional 296 interaction. Future studies will for sure reveal new aspects and implications of this pivot 297 role of endocannabinoids.

298

This short article tried to provide a rapid overview of some of the myriad pieces of evidence that support the key role of endocannabinoids in the control of body and cellular metabolism and of how this control translates into regulation of very different processes, from biochemistry to behavior. The exciting avenues opened by old and recent discoveries on endocannabinoid signaling will lead us to a better understanding of metabolic regulation and to the design of novel approaches for the treatment of 305 metabolic disorders. Moreover, accumulating evidence indicates that many diseases, 306 whose metabolic component has been so far overlooked, actually involve alterations of 307 cellular metabolism at different levels. Therefore, the raising role of endocannabinoids 308 in the control of this function will extend our understanding of the therapeutic 309 properties of cannabinoid-related drugs.

310 References

- 311
- 3121Pisanti, S. & Bifulco, M. Medical Cannabis: A plurimillennial history of an evergreen. J313Cell Physiol 234, 8342-8351, doi:10.1002/jcp.27725 (2019).
- 3142Crocq, M. A. History of cannabis and the endocannabinoid system. Dialogues Clin315Neurosci 22, 223-228, doi:10.31887/DCNS.2020.22.3/mcrocq (2020).
- Mazier, W., Saucisse, N., Gatta-Cherifi, B. & Cota, D. The Endocannabinoid System:
 Pivotal Orchestrator of Obesity and Metabolic Disease. *Trends Endocrinol Metab* 26,
 524-537, doi:10.1016/j.tem.2015.07.007 (2015).
- Adams, R., Aycock, B. F., Jr. & Loewe, S. Tetrahydrocannabinol homologs. *J Am Chem Soc* 70, 662-664, doi:10.1021/ja01182a067 (1948).
- Mechoulam, R. & Gaoni, Y. Hashish. IV. The isolation and structure of cannabinolic
 cannabidiolic and cannabigerolic acids. *Tetrahedron* 21, 1223-1229,
 doi:10.1016/0040-4020(65)80064-3 (1965).
- Matsuda, L. A., Lolait, S. J., Brownstein, M. J., Young, A. C. & Bonner, T. I. Structure of
 a cannabinoid receptor and functional expression of the cloned cDNA. *Nature* 346,
 561-564, doi:10.1038/346561a0 (1990).
- 3277Munro, S., Thomas, K. L. & Abu-Shaar, M. Molecular characterization of a peripheral328receptor for cannabinoids. *Nature* **365**, 61-65, doi:10.1038/365061a0 (1993).
- 3298Devane, W. A. *et al.* Isolation and structure of a brain constituent that binds to the330cannabinoid receptor. *Science* **258**, 1946-1949, doi:10.1126/science.1470919 (1992).
- Sugiura, T. *et al.* 2-Arachidonoylglycerol: a possible endogenous cannabinoid
 receptor ligand in brain. *Biochem Biophys Res Commun* 215, 89-97,
 doi:10.1006/bbrc.1995.2437 (1995).
- Mechoulam, R. *et al.* Identification of an endogenous 2-monoglyceride, present in
 canine gut, that binds to cannabinoid receptors. *Biochem Pharmacol* 50, 83-90,
 doi:10.1016/0006-2952(95)00109-d (1995).
- 33711Petrosino, S. & Di Marzo, V. FAAH and MAGL inhibitors: therapeutic opportunities338from regulating endocannabinoid levels. *Curr Opin Investig Drugs* **11**, 51-62 (2010).
- Kano, M., Ohno-Shosaku, T., Hashimotodani, Y., Uchigashima, M. & Watanabe, M.
 Endocannabinoid-mediated control of synaptic transmission. *Physiol Rev* 89, 309380, doi:10.1152/physrev.00019.2008 (2009).
- 342
 13
 Pertwee, R. G. Endocannabinoids and Their Pharmacological Actions. Handb Exp

 343
 Pharmacol 231, 1-37, doi:10.1007/978-3-319-20825-1_1 (2015).
- Ligresti, A., De Petrocellis, L. & Di Marzo, V. From Phytocannabinoids to Cannabinoid
 Receptors and Endocannabinoids: Pleiotropic Physiological and Pathological Roles
 Through Complex Pharmacology. *Physiol Rev* 96, 1593-1659,
 doi:10.1152/physrev.00002.2016 (2016).
- 34815Maccarrone, M. et al. Endocannabinoid signaling at the periphery: 50 years after349THC. Trends Pharmacol Sci **36**, 277-296, doi:10.1016/j.tips.2015.02.008 (2015).
- 35016Bauer, M. et al. Identification and quantification of a new family of peptide351endocannabinoids (Pepcans) showing negative allosteric modulation at CB1352receptors. J Biol Chem 287, 36944-36967, doi:10.1074/jbc.M112.382481 (2012).
- 35317Hofer, S. C. *et al.* Localization and production of peptide endocannabinoids in the354rodent CNS and adrenal medulla. Neuropharmacology98, 78-89,355doi:10.1016/j.neuropharm.2015.03.021 (2015).

- 356 Buisseret, B., Alhouayek, M., Guillemot-Legris, O. & Muccioli, G. G. Endocannabinoid 18 357 and Prostanoid Crosstalk in Pain. Trends Mol Med 25, 882-896, 358 doi:10.1016/j.molmed.2019.04.009 (2019).
- Pamplona, F. A. *et al.* Anti-inflammatory lipoxin A4 is an endogenous allosteric
 enhancer of CB1 cannabinoid receptor. *Proc Natl Acad Sci U S A* 109, 21134-21139,
 doi:10.1073/pnas.1202906109 (2012).
- Vallee, M. *et al.* Pregnenolone can protect the brain from cannabis intoxication.
 Science 343, 94-98, doi:10.1126/science.1243985 (2014).
- 36421Piazza, P. V., Cota, D. & Marsicano, G. The CB1 Receptor as the Cornerstone of365Exostasis. Neuron 93, 1252-1274, doi:10.1016/j.neuron.2017.02.002 (2017).
- 36622Bellocchio, L. *et al.* Bimodal control of stimulated food intake by the367endocannabinoid system. *Nat Neurosci* 13, 281-283, doi:10.1038/nn.2494 (2010).
- Ruiz de Azua, I. *et al.* Adipocyte cannabinoid receptor CB1 regulates energy
 homeostasis and alternatively activated macrophages. *J Clin Invest* 127, 4148-4162,
 doi:10.1172/JCI83626 (2017).
- Lee, Y., Jo, J., Chung, H. Y., Pothoulakis, C. & Im, E. Endocannabinoids in the
 gastrointestinal tract. *Am J Physiol Gastrointest Liver Physiol* **311**, G655-G666,
 doi:10.1152/ajpgi.00294.2015 (2016).
- Mendizabal-Zubiaga, J. *et al.* Cannabinoid CB1 Receptors Are Localized in Striated
 Muscle Mitochondria and Regulate Mitochondrial Respiration. *Front Physiol* 7, 476,
 doi:10.3389/fphys.2016.00476 (2016).
- Kunos, G. & Osei-Hyiaman, D. Endocannabinoids and liver disease. IV.
 Endocannabinoid involvement in obesity and hepatic steatosis. *Am J Physiol Gastrointest Liver Physiol* 294, G1101-1104, doi:10.1152/ajpgi.00057.2008 (2008).
- 38027Doyle, M. E. The role of the endocannabinoid system in islet biology. Curr Opin381Endocrinol Diabetes Obes 18, 153-158, doi:10.1097/MED.0b013e32834455a8 (2011).
- Lutz, B., Marsicano, G., Maldonado, R. & Hillard, C. J. The endocannabinoid system in
 guarding against fear, anxiety and stress. *Nat Rev Neurosci* 16, 705-718,
 doi:10.1038/nrn4036 (2015).
- Corcoran, L., Roche, M. & Finn, D. P. The Role of the Brain's Endocannabinoid System
 in Pain and Its Modulation by Stress. *Int Rev Neurobiol* 125, 203-255,
 doi:10.1016/bs.irn.2015.10.003 (2015).
- 388 30 Moreira, F. A. & Wotjak, C. T. Cannabinoids and anxiety. *Curr Top Behav Neurosci* 2, 429-450, doi:10.1007/7854_2009_16 (2010).
- 390 31 Dubreucq, S. *et al.* Ventral tegmental area cannabinoid type-1 receptors control
 391 voluntary exercise performance. *Biol Psychiatry* 73, 895-903,
 392 doi:10.1016/j.biopsych.2012.10.025 (2013).
- 39332Muguruza, C. *et al.* The motivation for exercise over palatable food is dictated by394cannabinoid type-1 receptors. *JCl Insight* **4**, doi:10.1172/jci.insight.126190 (2019).
- 33 Fuss, J. *et al.* A runner's high depends on cannabinoid receptors in mice. *Proc Natl* 396 *Acad Sci U S A* **112**, 13105-13108, doi:10.1073/pnas.1514996112 (2015).
- 39734Soria-Gomez, E. *et al.* The endocannabinoid system controls food intake via olfactory398processes. Nat Neurosci **17**, 407-415, doi:10.1038/nn.3647 (2014).
- 39935Yazulla, S. Endocannabinoids in the retina: from marijuana to neuroprotection. *Prog*400*Retin Eye Res* 27, 501-526, doi:10.1016/j.preteyeres.2008.07.002 (2008).

- Scheen, A. J. *et al.* Efficacy and tolerability of rimonabant in overweight or obese
 patients with type 2 diabetes: a randomised controlled study. *Lancet* 368, 16601672, doi:10.1016/S0140-6736(06)69571-8 (2006).
- 404 37 Mitchell, P. B. & Morris, M. J. Depression and anxiety with rimonabant. *Lancet* 370, 1671-1672, doi:10.1016/S0140-6736(07)61705-X (2007).
- 406 38 Despres, J. P., Golay, A., Sjostrom, L. & Rimonabant in Obesity-Lipids Study, G. Effects
 407 of rimonabant on metabolic risk factors in overweight patients with dyslipidemia. *N*408 *Engl J Med* **353**, 2121-2134, doi:10.1056/NEJMoa044537 (2005).
- Pi-Sunyer, F. X. et al. Effect of rimonabant, a cannabinoid-1 receptor blocker, on 409 39 410 weight and cardiometabolic risk factors in overweight or obese patients: RIO-North 411 randomized controlled trial. JAMA America: а 295, 761-775, 412 doi:10.1001/jama.295.7.761 (2006).
- 40 Gatta-Cherifi, B. & Cota, D. New insights on the role of the endocannabinoid system 414 in the regulation of energy balance. *Int J Obes (Lond)* **40**, 210-219, 415 doi:10.1038/ijo.2015.179 (2016).
- 416 41 Cota, D. *et al.* The endogenous cannabinoid system affects energy balance via central
 417 orexigenic drive and peripheral lipogenesis. *J Clin Invest* **112**, 423-431,
 418 doi:10.1172/JCI17725 (2003).
- 419 42 Deveaux, V. *et al.* Cannabinoid CB2 receptor potentiates obesity-associated
 420 inflammation, insulin resistance and hepatic steatosis. *PLoS One* 4, e5844,
 421 doi:10.1371/journal.pone.0005844 (2009).
- 422 43 Agudo, J. *et al.* Deficiency of CB2 cannabinoid receptor in mice improves insulin 423 sensitivity but increases food intake and obesity with age. *Diabetologia* **53**, 2629-424 2640, doi:10.1007/s00125-010-1894-6 (2010).
- 425 44 Simon, V. & Cota, D. MECHANISMS IN ENDOCRINOLOGY: Endocannabinoids and 426 metabolism: past, present and future. *Eur J Endocrinol* **176**, R309-R324, 427 doi:10.1530/EJE-16-1044 (2017).
- 428 45 Melis, T. et al. The cannabinoid antagonist SR 141716A (Rimonabant) reduces the 429 increase of extra-cellular dopamine release in the rat nucleus accumbens induced by 430 palatable novel high food. Neurosci Lett 419, 231-235, а 431 doi:10.1016/j.neulet.2007.04.012 (2007).
- 432 46 Zangen, A., Solinas, M., Ikemoto, S., Goldberg, S. R. & Wise, R. A. Two brain sites for 433 cannabinoid reward. *J Neurosci* **26**, 4901-4907, doi:10.1523/JNEUROSCI.3554-434 05.2006 (2006).
- 43547Busquets-Garcia, A. et al. Dissecting the cannabinergic control of behavior: The436where matters. Bioessays 37, 1215-1225, doi:10.1002/bies.201500046 (2015).
- 43748Mazier, W. et al. mTORC1 and CB1 receptor signaling regulate excitatory438glutamatergic inputs onto the hypothalamic paraventricular nucleus in response to439energy availability. Mol Metab 28, 151-159, doi:10.1016/j.molmet.2019.08.005440(2019).
- 49 Jarbe, T. U. & DiPatrizio, N. V. Delta9-THC induced hyperphagia and tolerance
 442 assessment: interactions between the CB1 receptor agonist delta9-THC and the CB1
 443 receptor antagonist SR-141716 (rimonabant) in rats. *Behav Pharmacol* 16, 373-380,
 444 doi:10.1097/00008877-200509000-00009 (2005).
- Tam, J. *et al.* Role of adiponectin in the metabolic effects of cannabinoid type 1
 receptor blockade in mice with diet-induced obesity. *Am J Physiol Endocrinol Metab* **306**, E457-468, doi:10.1152/ajpendo.00489.2013 (2014).

- Vettor, R. & Pagano, C. The role of the endocannabinoid system in lipogenesis and
 fatty acid metabolism. *Best Pract Res Clin Endocrinol Metab* 23, 51-63,
 doi:10.1016/j.beem.2008.10.002 (2009).
- 451 52 Ruby, M. A. *et al.* Overactive endocannabinoid signaling impairs apolipoprotein E-452 mediated clearance of triglyceride-rich lipoproteins. *Proc Natl Acad Sci U S A* **105**, 453 14561-14566, doi:10.1073/pnas.0807232105 (2008).
- You, T., Disanzo, B. L., Wang, X., Yang, R. & Gong, D. Adipose tissue endocannabinoid
 system gene expression: depot differences and effects of diet and exercise. *Lipids Health Dis* 10, 194, doi:10.1186/1476-511X-10-194 (2011).
- 457 54 Engeli, S. *et al.* Activation of the peripheral endocannabinoid system in human 458 obesity. *Diabetes* **54**, 2838-2843, doi:10.2337/diabetes.54.10.2838 (2005).
- Tedesco, L. *et al.* Cannabinoid type 1 receptor blockade promotes mitochondrial
 biogenesis through endothelial nitric oxide synthase expression in white adipocytes. *Diabetes* 57, 2028-2036, doi:10.2337/db07-1623 (2008).
- Kim, Y. *et al.* Hepatocyte cannabinoid 1 receptor nullification alleviates toxin-induced
 liver damage via NF-kappaB signaling. *Cell Death Dis* **11**, 1044, doi:10.1038/s41419020-03261-8 (2020).
- 465 57 Irungbam, K. *et al.* Cannabinoid receptor 1 knockout alleviates hepatic steatosis by
 466 downregulating perilipin 2. *Lab Invest* 100, 454-465, doi:10.1038/s41374-019-0327-5
 467 (2020).
- 468 58 Osei-Hyiaman, D. *et al.* Hepatic CB1 receptor is required for development of diet469 induced steatosis, dyslipidemia, and insulin and leptin resistance in mice. *J Clin Invest*470 **118**, 3160-3169, doi:10.1172/JCI34827 (2008).
- Wang, S. *et al.* Cannabinoid receptor 1 signaling in hepatocytes and stellate cells
 does not contribute to NAFLD. *J Clin Invest* **131**, doi:10.1172/JCI152242 (2021).
- 47360Kimberly, W. T. *et al.* Metabolite profiling identifies anandamide as a biomarker of474nonalcoholic steatohepatitis. *JCl Insight* **2**, doi:10.1172/jci.insight.92989 (2017).
- Liu, J. *et al.* Hepatic cannabinoid receptor-1 mediates diet-induced insulin resistance
 via inhibition of insulin signaling and clearance in mice. *Gastroenterology* 142, 12181228 e1211, doi:10.1053/j.gastro.2012.01.032 (2012).
- 47862Margheritis, E. *et al.* Bile Acid Recognition by NAPE-PLD. ACS Chem Biol **11**, 2908-4792914, doi:10.1021/acschembio.6b00624 (2016).
- 480 63 Berland, C. *et al.* Identification of an endocannabinoid gut-brain vagal mechanism
 481 controlling food reward and energy homeostasis. *Mol Psychiatry*,
 482 doi:10.1038/s41380-021-01428-z (2022).
- 48364Tellez, L. A. et al. A gut lipid messenger links excess dietary fat to dopamine484deficiency. Science 341, 800-802, doi:10.1126/science.1239275 (2013).
- 48565DiPatrizio, N. V. Endocannabinoids and the Gut-Brain Control of Food Intake and486Obesity. Nutrients 13, doi:10.3390/nu13041214 (2021).
- Burdyga, G., Varro, A., Dimaline, R., Thompson, D. G. & Dockray, G. J. Expression of
 cannabinoid CB1 receptors by vagal afferent neurons: kinetics and role in influencing
 neurochemical phenotype. *Am J Physiol Gastrointest Liver Physiol* 299, G63-69,
 doi:10.1152/ajpgi.00059.2010 (2010).
- 491 67 Quarta, C. *et al.* CB(1) signaling in forebrain and sympathetic neurons is a key
 492 determinant of endocannabinoid actions on energy balance. *Cell Metab* 11, 273-285,
 493 doi:10.1016/j.cmet.2010.02.015 (2010).

- 494 68 Bellocchio, L. *et al.* Activation of the sympathetic nervous system mediates
 495 hypophagic and anxiety-like effects of CB(1) receptor blockade. *Proc Natl Acad Sci U*496 S A 110, 4786-4791, doi:10.1073/pnas.1218573110 (2013).
- 497 69 Bartova, A. & Birmingham, M. K. Effect of delta9-tetrahydrocannabinol on 498 mitochondrial NADH-oxidase activity. *J Biol Chem* **251**, 5002-5006 (1976).
- 49970Chari-Bitron, A. & Bino, T. Effect of 1-tetrahydrocannabinol on ATPase activity of rat500liver mitochondria. Biochem Pharmacol **20**, 473-475, doi:10.1016/0006-5012952(71)90084-0 (1971).
- 502 71 Chiu, P., Karler, R., Craven, C., Olsen, D. M. & Turkanis, S. A. The influence of delta9503 tetrahydrocannabinol, cannabinol and cannabidiol on tissue oxygen consumption.
 504 *Res Commun Chem Pathol Pharmacol* 12, 267-286 (1975).
- Sarkar, C. & Ghosh, J. J. Effect of delta-9-tetrahydrocannabinol administration on the
 lipid constituents of rat brain subcellular fractions. *J Neurochem* 24, 381-385,
 doi:10.1111/j.1471-4159.1975.tb11891.x (1975).
- Rossato, M., Ion Popa, F., Ferigo, M., Clari, G. & Foresta, C. Human sperm express
 cannabinoid receptor Cb1, the activation of which inhibits motility, acrosome
 reaction, and mitochondrial function. *J Clin Endocrinol Metab* **90**, 984-991,
 doi:10.1210/jc.2004-1287 (2005).
- 51274Aquila, S. et al. Human sperm anatomy: ultrastructural localization of the513cannabinoid1 receptor and a potential role of anandamide in sperm survival and514acrosome reaction. Anat Rec (Hoboken) 293, 298-309, doi:10.1002/ar.21042 (2010).
- 515 75 Badawy, Z. S. *et al.* Cannabinoids inhibit the respiration of human sperm. *Fertil Steril*516 **91**, 2471-2476, doi:10.1016/j.fertnstert.2008.03.075 (2009).
- 517 76 Benard, G. *et al.* Mitochondrial CB(1) receptors regulate neuronal energy 518 metabolism. *Nat Neurosci* **15**, 558-564, doi:10.1038/nn.3053 (2012).
- 51977Jong, Y. I., Harmon, S. K. & O'Malley, K. L. Intracellular GPCRs Play Key Roles in520SynapticPlasticity.ACSChemNeurosci9,2162-2172,521doi:10.1021/acschemneuro.7b00516 (2018).
- 522 78 Wang, Q. *et al.* 5-HTR3 and 5-HTR4 located on the mitochondrial membrane and 523 functionally regulated mitochondrial functions. *Sci Rep* **6**, 37336, 524 doi:10.1038/srep37336 (2016).
- 525 79 Lahuna, O. & Jockers, R. [Mitochondrial signaling of G protein-coupled receptors].
 526 *Biol Aujourdhui* 212, 21-26, doi:10.1051/jbio/2018024 (2018).
- 527 80 Belous, A. *et al.* Mitochondrial P2Y-Like receptors link cytosolic adenosine 528 nucleotides to mitochondrial calcium uptake. *J Cell Biochem* **92**, 1062-1073, 529 doi:10.1002/jcb.20144 (2004).
- 53081Koch, M. *et al.* Hypothalamic POMC neurons promote cannabinoid-induced feeding.531Nature **519**, 45-50, doi:10.1038/nature14260 (2015).
- 53282Kamnate, A. et al. Mitochondrial Localization of CB1 in Progesterone-producing Cells533of Ovarian Interstitial Glands of Adult Mice. J Histochem Cytochem,534221554211063516, doi:10.1369/00221554211063516 (2021).
- 535 83 Jimenez-Blasco, D. *et al.* Glucose metabolism links astroglial mitochondria to 536 cannabinoid effects. *Nature* **583**, 603-608, doi:10.1038/s41586-020-2470-y (2020).
- 53784Hebert-Chatelain, E. *et al.* A cannabinoid link between mitochondria and memory.538Nature 539, 555-559, doi:10.1038/nature20127 (2016).

- Amoako, A. A. *et al.* Anandamide modulates human sperm motility: implications for
 men with asthenozoospermia and oligoasthenoteratozoospermia. *Hum Reprod* 28,
 2058-2066, doi:10.1093/humrep/det232 (2013).
- 54286Soria-Gomez, E. *et al.* Subcellular specificity of cannabinoid effects in striatonigral543circuits. *Neuron* **109**, 1513-1526 e1511, doi:10.1016/j.neuron.2021.03.007 (2021).
- 54487Almeida, A., Moncada, S. & Bolanos, J. P. Nitric oxide switches on glycolysis through545the AMP protein kinase and 6-phosphofructo-2-kinase pathway. Nat Cell Biol 6, 45-54651, doi:10.1038/ncb1080 (2004).
- 547 88 Bolanos, J. P., Peuchen, S., Heales, S. J., Land, J. M. & Clark, J. B. Nitric oxide-548 mediated inhibition of the mitochondrial respiratory chain in cultured astrocytes. *J* 549 *Neurochem* **63**, 910-916, doi:10.1046/j.1471-4159.1994.63030910.x (1994).
- 55089Supplie, L. M. *et al.* Respiration-Deficient Astrocytes Survive As Glycolytic Cells In551Vivo. J Neurosci **37**, 4231-4242, doi:10.1523/JNEUROSCI.0756-16.2017 (2017).
- 552 90 Fiebig, C. *et al.* Mitochondrial Dysfunction in Astrocytes Impairs the Generation of 553 Reactive Astrocytes and Enhances Neuronal Cell Death in the Cortex Upon 554 Photothrombotic Lesion. *Front Mol Neurosci* **12**, 40, doi:10.3389/fnmol.2019.00040 555 (2019).
- 55691Bolanos, J. P., Almeida, A. & Moncada, S. Glycolysis: a bioenergetic or a survival557pathway? *Trends Biochem Sci* **35**, 145-149, doi:10.1016/j.tibs.2009.10.006 (2010).
- Herrero-Mendez, A. *et al.* The bioenergetic and antioxidant status of neurons is
 controlled by continuous degradation of a key glycolytic enzyme by APC/C-Cdh1. *Nat Cell Biol* 11, 747-752, doi:10.1038/ncb1881 (2009).
- Machler, P. *et al.* In Vivo Evidence for a Lactate Gradient from Astrocytes to Neurons. *Cell Metab* 23, 94-102, doi:10.1016/j.cmet.2015.10.010 (2016).
- Allaman, I., Belanger, M. & Magistretti, P. J. Astrocyte-neuron metabolic
 relationships: for better and for worse. *Trends Neurosci* 34, 76-87,
 doi:10.1016/j.tins.2010.12.001 (2011).
- 56695Bonvento, G. & Bolanos, J. P. Astrocyte-neuron metabolic cooperation shapes brain567activity. *Cell Metab* **33**, 1546-1564, doi:10.1016/j.cmet.2021.07.006 (2021).
- 56896Suzuki, A. *et al.* Astrocyte-neuron lactate transport is required for long-term memory569formation. *Cell* **144**, 810-823, doi:10.1016/j.cell.2011.02.018 (2011).
- 570 97 Carrard, A. *et al.* Peripheral administration of lactate produces antidepressant-like 571 effects. *Mol Psychiatry*, doi:10.1038/mp.2016.237 (2016).
- 572 98 Sena, L. A. & Chandel, N. S. Physiological roles of mitochondrial reactive oxygen 573 species. *Mol Cell* **48**, 158-167, doi:10.1016/j.molcel.2012.09.025 (2012).
- 574 99 Murphy, M. P. How mitochondria produce reactive oxygen species. *Biochem J* **417**, 1-575 13, doi:10.1042/BJ20081386 (2009).
- Lopez-Fabuel, I. *et al.* Complex I assembly into supercomplexes determines
 differential mitochondrial ROS production in neurons and astrocytes. *Proc Natl Acad Sci U S A* 113, 13063-13068, doi:10.1073/pnas.1613701113 (2016).
- Vicente-Gutierrez, C. *et al.* Astrocytic mitochondrial ROS modulate brain metabolism
 and mouse behaviour. *Nat Metab* 1, 201-211, doi:10.1038/s42255-018-0031-6
 (2019).
- 582 102 Mimaki, M., Wang, X., McKenzie, M., Thorburn, D. R. & Ryan, M. T. Understanding
 583 mitochondrial complex I assembly in health and disease. *Biochim Biophys Acta* 1817,
 584 851-862, doi:10.1016/j.bbabio.2011.08.010 (2012).

- 585 103 Guaras, A. M. & Enriquez, J. A. Building a Beautiful Beast: Mammalian Respiratory 586 Complex I. *Cell Metab* **25**, 4-5, doi:10.1016/j.cmet.2016.12.019 (2017).
- 587 104 De Rasmo, D. *et al.* Activation of the cAMP cascade in human fibroblast cultures 588 rescues the activity of oxidatively damaged complex I. *Free Radic Biol Med* **52**, 757-589 764, doi:10.1016/j.freeradbiomed.2011.11.030 (2012).
- Patten, D. A. *et al.* Hypoxia-inducible factor-1 activation in nonhypoxic conditions:
 the essential role of mitochondrial-derived reactive oxygen species. *Mol Biol Cell* 21,
 3247-3257, doi:10.1091/mbc.E10-01-0025 (2010).
- 593 106 Semenza, G. L., Roth, P. H., Fang, H. M. & Wang, G. L. Transcriptional regulation of 594 genes encoding glycolytic enzymes by hypoxia-inducible factor 1. *J Biol Chem* **269**, 595 23757-23763 (1994).
- 596107Serrat, R. et al.AstroglialER-mitochondriacalciumtransfermediates597endocannabinoid-dependentsynapticintegration.CellRep37,110133,598doi:10.1016/j.celrep.2021.110133 (2021).
- 599 600

601 Acknowledgements

602 This study is dedicated to the memory of our friend and colleague Federico Massa. We 603 thank Daniela Cota and Francis Chaouloff for critically reading the manuscript. We also 604 thank all the members of Busquets-Garcia's, Bolaños's and Marsicano's labs for useful 605 discussions and for their invaluable support. The work of A.Busquets-Garcia is supported by: 606 Agencia Estatal de Investigación-FEDER (RTI2018-093667-A-100); the IBRO Return Home 607 Fellowships 2019 and the Ramon v Cajal programme (RYC-2017-21776) funded by MCIN/ 608 AEI/10.13039/501100011033 and FSE). The work of J.P. Bolanos is funded by the Agencia 609 Estatal de Investigación (PID2019-105699RB-I00; PDC2021-121013-I00; RED2018-102576-T; 610 MCIN/AEI/10.13039/501100011033 & European Union NextGenerationEU/PRTR), Plan Nacional de Drogas (Ministerio de Sanidad; 2020I028), Junta de Castilla y León (CS/151P20 611 and Escalera de Excelencia CLU-2017-03), and Fundación Ramón Areces. The work of G. 612 Marsicano is funded by: INSERM, European Research Council (Endofood, ERC-2010-StG-613 ERC-2014-PoC-640923, MiCaBra, 614 260515 and CannaPreg, ERC-2017-AdG-786467), Fondation pour la Recherche Medicale (FRM, DRM20101220445), the Human Frontiers 615 616 Science Program, Region Nouvelle Aquitaine, Agence Nationale de la Recherche (ANR, NeuroNutriSens ANR-13-BSV4-0006 and ORUPS ANR-16-CE37-0010-01 and BRAIN ANR-10-617 618 LABX-0043).

620 Figure Legends

621

622Figure 1. Timeline of (endo)cannabinoid research.On the left, Timeline of recorded623publications (green bars) using the keywords "Cannabis OR Cannabinoid OR624Endocannabinoid" (source PubMed,

625 <u>https://pubmed.ncbi.nlm.nih.gov/?term=cannabinoid+or+endocannabinoid+or+cannabis</u>).

626 Time is selected after 1940 to improve clarity (No more than 1-2 publications per year from 627 1840s until 1940s). First publications on cannabis appear in the middle of 1800, when the 628 plant is imported from East. The chemical composition of the plant is characterized between 629 1940 and 1964. Notice the peak of publications on the subject in the following period. The number of publications then decreases, likely reflecting the difficulty in finding a clear 630 mechanism of action of cannabinoids. Publication rate starts again to grow exponentially 631 after the characterization of receptors and endocannabinoids around early 1990s. The 632 633 number of publications per year did not stop growing since. In 2021, 5,302 papers appeared on the subject. Adding the keyword "metabolism" reduces the absolute numbers (2,923 634 635 papers in 2021) but maintains the same trend overtime (not shown). On the right, schematic 636 and simplified representation of the main processes involved in the synthesis and 637 degradation of the two main endocannabinoids, 2-AG and AEA.

638

Figure 2. Target locations and metabolic activities of endocannabinoids. Endocannabinoids 639 640 and their receptors (here CB1 receptors) are highly conserved between mice and humans. 641 Most observations have been made in rodents but are also supported by human data. 642 Amongst other functions, endocannabinoids control brain functions to increase food intake 643 (particularly palatable and energy-rich) and regulate bioenergetic functions such as 644 mitochondria respiration and astroglial glucose metabolism. In addition, endocannabinoids 645 increase lipogenesis in the liver, increase insulin secretion in the pancreas, increase 646 nutrients absorption in the intestinal tract, decrease glucose use in the muscle and favor the 647 accumulation of fat in the adipose system.

648

Figure 3. Diverse CB1R signalling cascades and their functions. A representative and
 simplified description of the CB1 receptor signalling depending on their location is shown: 1)
 CB1 receptors are (mostly) integrated in the membranes of presynaptic neurons where are

652 coupled to Gi/o proteins that control several signalling cascades including the PI3K-AKT, the 653 MAPK and the PKA cascades. This neuronal CB1 receptors are involved in the control of 654 neurotransmission (NT) and cell survival. 2) Astroglial CB1 receptors are mainly coupled to 655 Gq protein. Their activacion increases calcium influx and regulates gliotransmission, among 656 other processes. As represented in the Figure, other G proteins have been linked with CB1 657 receptors being involved in CB1 signalling. 3) CB1 receptors associated to mitochondrial membranes are involved in the regulation of mitochondrial respiration through a specific 658 659 signalling cascade, which impact neuronal functions and astroglial glycolisis and lactate 660 release. In adddition, astroglial mtCB1 receptors are involved in the calcium transfer 661 between ER and mitochondrial, which impacts lateral synaptic potentiation.

1840-60: Introduction of Cannabis to West

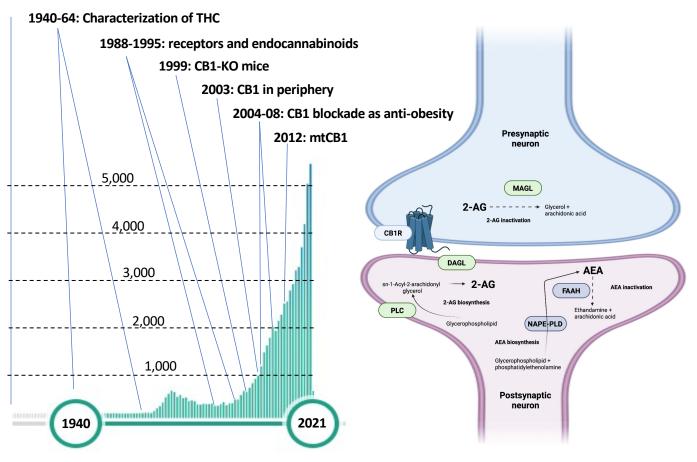


Figure 1. Timeline of (endo)cannabinoid research. On the left, Timeline of recorded publications (green bars) using the keywords "Cannabis OR Cannabinoid OR Endocannabinoid" (source PubMed, https://pubmed.ncbi.nlm.nih.gov/?term=cannabinoid+or+endocannabinoid+or+cannabis). Time is selected after 1940 to improve clarity (No more than 1-2 publications per year from 1840s until 1940s). First publications on cannabis appear in the middle of 1800, when the plant is imported from East. The chemical composition of the plant is characterized between 1940 and 1964. Notice the peak of publications on the subject in the following period. The number of publications then decreases, likely reflecting the difficulty in finding a clear mechanism of action of cannabinoids. Publication rate starts again to grow exponentially after the characterization of receptors and endocannabinoids around early 1990s. The number of publications per year did not stop growing since. In 2021, 5,302 papers appeared on the subject. Adding the keyword "metabolism" reduces the absolute numbers (2,923 papers in 2021), but maintains the same trend overtime (not shown). On the right, schematic and simplified representation of the main processes involved in the synthesis and degradation of the two main endocannabinoids, 2-AG and AEA.

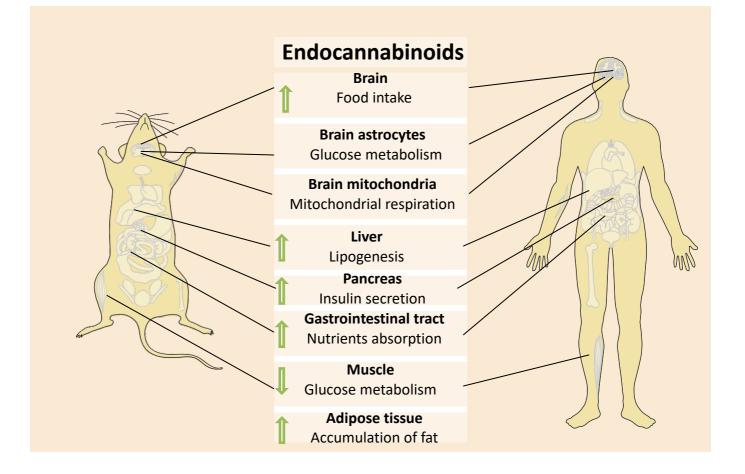
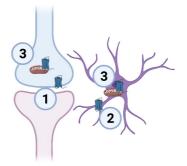


Figure 2. Target locations and metabolic activities of endocannabinoids. Endocannabinoids and their receptors (here CB1 receptors) are highly conserved between mice and humans. Most observations have been made in rodents but are also supported by human data. Amongst other functions, endocannabinoids control brain functions in order to increase food intake (particularly palatable and energy-rich) and regulate bioenergetic functions such as mitochondria respiration and astroglial glucose metabolism. In addition, endocannabinoids increase lipogenesis in the liver, increase insulin secretion in the pancreas, increase nutrients absorption in the intestinal tract, decrease glucose use in the muscle and favor the accumulation of fat in the adipose system.



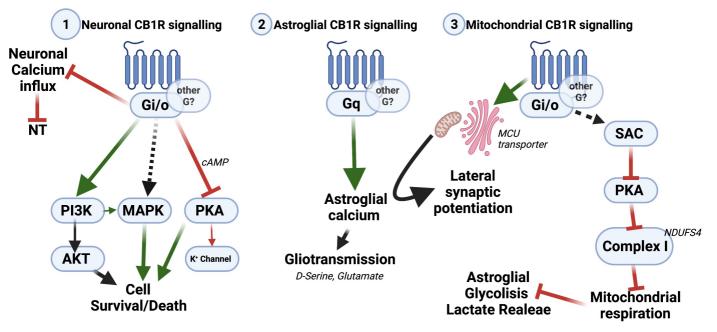


Figure 3. Diverse CB1R signalling cascades and their functions. A representative and simplified description of the CB1 receptor signalling depending on their location is shown: 1) CB1 receptors are (mostly) integrated in the membranes of presynaptic neurons where are coupled to Gi/o proteins that control several signalling cascades including the PI3K-AKT, the MAPK and the PKA cascades. This neuronal CB1 receptors are involved in the control of neurotransmission (NT) and cell survival. 2) Astroglial CB1 receptors are mainly coupled to Gq protein. Their activacion increases calcium influx and regulates gliotransmission, among other processes. As represented in the Figure, other G proteins have been linked with CB1 receptors being involved in CB1 signalling. 3) CB1 receptors associated to mitochondrial membranes are involved in the regulation of mitochondrial respiration through a specific signalling cascade, which impact neuronal functions and astroglial glycolisis and lactate release. In adddition, astroglial mtCB1 receptors are involved in the calcium transfer between ER and mitochondrial, which impacts lateral synaptic potentiation.