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3	Yin-Yang control of energy balance by lipids in the hypothalamus:
4	the endocannabinoids vs bile acids case
5	Thomas H. Lee <sup>1</sup> , Daniela Cota <sup>1</sup> , Carmelo Quarta <sup>1#</sup>
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7	<sup>1</sup> University of Bordeaux, INSERM, Neurocentre Magendie, U1215, F-3300 Bordeaux, France.
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10	#Corresponding author: <u>carmelo.quarta@inserm.fr</u>
11	Inserm U1215, Neurocentre Magendie 146 rue Léo Saignat 33077 Bordeaux cedex - FRANCE
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13	Highlights
14	- Endocannabinoids and bile acids act as Yin-Yang regulators of the body energy needs
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16	- Master lipid sensors in the brain could integrate the actions of multiple lipid signals
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18	- Obesity might originate from dysregulation of master lipid sensors in the brain
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### 30 Abstract

Obesity is a chronic and debilitating disorder that originates from alterations in energysensing brain circuits controlling body weight gain and food intake. The dysregulated syntheses and actions of lipid mediators in the hypothalamus induce weight gain and overfeeding, but the molecular and cellular underpinnings of these alterations remain elusive.

In response to changes in the nutritional status, different lipid sensing pathways in the 35 36 hypothalamus direct body energy needs in a Yin-Yang model. Endocannabinoids orchestrate the crosstalk between hypothalamic circuits and the sympathetic nervous system to promote food 37 38 intake and energy accumulation during fasting, whereas bile acids act on the same top-down axis to reduce energy intake and possibly storage after the meal. In obesity, the bioavailability and 39 40 downstream cellular actions of endocannabinoids and bile acids are altered in hypothalamic neurons involved in body weight and metabolic control. Thus, the onset and progression of this 41 disease might result from an imbalance in hypothalamic sensing of multiple lipid signals, which 42 are possibly integrated by common molecular nodes. 43

In this viewpoint, we discuss a possible model that explains how bile acids and endocannabinoids may exert their effects on energy balance regulation via interconnected mechanisms at the level of the hypothalamic neuronal circuits. Therefore, we propose a new conceptual framework for understanding and treating central mechanisms of maladaptive lipid action in obesity.

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#### 50 Keywords

51 Obesity, endocannabinoids, bile acids, hypothalamus, lipid metabolism

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# 62 Abbreviations

Central nervous system, CNS; arcuate nucleus, ARC; neuropeptide Y, NPY; agouti-related 63 peptide, AgRP; amphetamine-regulated transcript, CART; proopiomelanocortin, POMC, blood-64 brain barrier, BBB; lipoprotein lipase, LPL; hormone-sensitive lipase, HSL; high-fat diet, HFD; 65 mediobasal hypothalamus, MBH; sympathetic nervous system, SNS;  $\Delta^9$ -tetrahydrocannabinol, 66 THC; cannabinoid type 1 receptors, CB1; brown adipose tissue, BAT; 2-arachidonoylglycerol, 2-67 68 AG; single-minded 1, SIM1; steroidogenic factor 1, SF1;  $2\alpha/\beta$ -hydrolase domain containing 6, ABHD6; ventromedial hypothalamic nucleus, VMH; 2-arachidonoylglycerol, 2-AG; nucleus 69 tractus solitarii, NTS, arachidonoylethanolamine, AEA; body mass index, BMI; farnesoid X 70 71 receptor, FXR; G-protein coupled receptor G-protein bile acid-activated receptor, GPBAR-72 1/TGR5; glucagon-like peptide-1, GLP-1; cholic acid, CA; chenodeoxycholic acid, CDCA; diettyrosine hydroxylase, 73 induced obese, DIO: TH: oleoylethanolamide, OEA: N-74 acylphosphatidylethanolamine-selective phospholipase D, NAPE-PLD; N-acylethanolamines, NAEs; small extracellular vesicles, sEV. 75

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### 77 1) Introduction

78 Over million years of evolution in an environment characterised by cataclysms and often scarce energy sources, the mammalian brain has developed adaptive mechanisms that promote 79 survival by manoeuvring the body energy state. Multiple studies have shed light on the 80 81 neurobiology of these mechanisms, and this advancement provides a more precise framework for 82 understanding and treating pathological conditions linked to altered energy balance regulation, such as obesity. Accordingly, we now know that obesity is primarily a brain disease since most of 83 84 the genetic mutations underlying disease progression map in the central nervous system (CNS). These mutations affect molecular factors responsible for synaptic transmission and neuronal 85 86 responses to hormones and energy substrates (1).

The outstanding ability of CNS neurons to monitor and maintain energy balance relies on its dynamic crosstalk with the peripheral organs. In the hypothalamic arcuate nucleus (ARC), the heterogeneous populations of neurons relay the peripheral signals by producing neurotransmitters

and neuropeptides that regulate feeding and metabolism (2-4). When energy reserves decline 90 91 during fasting, neuropeptide Y (NPY) and agouti-related peptide (AgRP) co-expressing neurons 92 are activated to promote food consumption and energy accumulation in peripheral organs (5). Conversely, when energy becomes available after dietary intake, neurons that co-express 93 amphetamine-regulated transcript (CART) and proopiomelanocortin (POMC) are recruited to 94 antagonise the actions of NPY/AgRP neurons to reduce food intake and stimulate whole-body 95 energy expenditure (6). AgRP and POMC neurons compose a unique neural circuit known as the 96 melanocortin system (6), which is modulated by the neighbouring non-neuronal cells, including 97 98 microglia, astrocytes, tanycytes, and endothelial cells (7). Non-neuronal cells regulate the melanocortin system by two mechanisms, the secretion of signalling molecules that influence 99 100 synaptic plasticity directly and the modulation of blood-brain barrier (BBB) permeability (7), 101 which subsequently impacts the transport of metabolic messengers from the bloodstream to the 102 brain. As thoroughly reviewed, hypothalamic neuronal circuits integrate these peripheral metabolic signals into output autonomic responses that influence feeding behaviour and 103 104 maintain the metabolic processes necessary for survival (8, 9).

The intercellular signalling between hypothalamic cells is tightly controlled by lipid-105 106 derived messengers, either produced from cellular precursors in the brain or transported from the bloodstream. Free fatty acids, for instance, can cross the BBB (10-13). These lipids can also 107 108 reach the hypothalamus as triglyceride-rich lipoproteins, which are then hydrolysed by enzymes, 109 such as the lipoprotein lipase (LPL) in neurons and glia (14, 15). Genetic ablation of LPL in glial 110 cells induces exaggerated body weight gain and glucose intolerance in mice exposed to a high-fat 111 diet (HFD), possibly impacting the function of hypothalamic neuronal circuits (15, 16). Likewise, non-selective deletion of LPL in the mediobasal hypothalamus (MBH) by viral transfection 112 113 induces weight gain and glucose intolerance in mice fed with a chow diet (17). Additionally, hormone-sensitive lipase (HSL), which regulates intracellular lipolysis, is expressed in 114 appetite-regulating hypothalamic neurons, and its activity in the MBH suppresses stress-115 induced food intake and HFD-induced obesity (18). 116

Hence, the action of lipids in the hypothalamus is critical for body weight control and the maintenance of energy homeostasis, although the resulting neurophysiological adaptations can be either beneficial or detrimental to metabolic health. The chemical configuration of the lipid species, for instance, the double bonds in the fatty acids, can affect the metabolic outcome. Accordingly, intracerebroventricular administration of the saturated palmitic acid enhances hepatic gluconeogenesis in mice, possibly due to central leptin resistance and inflammatory responses in the hypothalamus (19), whereas central administration of unsaturated oleic acid modulates the excitability of hypothalamic POMC neurons (20) and elicits beneficial metabolic responses, including inhibition of hepatic glucose production and food intake (21, 22).

Of note, certain lipid species, such as endocannabinoids (ECs), can act as retrograde 126 127 neuromodulators regulating neural plasticity in the CNS, including the hypothalamus. ECs are endogenous ligands of cannabinoid receptors that favour food intake and energy accumulation 128 129 when energy is scarce, for instance, during fasting (23). These lipid mediators inform the body about low energy availability by acting on the hypothalamus-sympathetic nervous system 130 131 (SNS) axis (23). In contrast, other periphery-derived lipids, such as bile acids (BAs), can convey information about energy accumulation in the body through the same top-down axis 132 133 (24, 25). After being produced by the liver and released in the gastrointestinal tract following dietary intake, BAs are reabsorbed into the circulation and act on the 134 135 hypothalamus to enhance SNS activity, promote energy dissipation, and inhibit appetite, 136 thus eliciting opposite physiological effects to ECs (24, 25). Intriguingly, the bioavailability and downstream signalling cascades of both ECs and BAs are altered in the hypothalamus 137 in obesity. These multiple lipid-mediated cascades might therefore converge to common 138 139 molecular nodes that act as 'super lipid sensors' in regulating metabolism, while the onset 140 and progression of metabolic diseases might result from the unbalanced regulations by these cascades. 141

Here we will discuss the recent evidence on the roles of ECs and BAs in fine-tuning the hypothalamic circuits regulating energy balance. Moreover, we will interrogate the potential interconnection of BAs- and ECs-mediated signalling pathways along the hypothalamusperiphery axes implicated in body weight control. Finally, we will highlight how these investigations have led us to propose a novel framework for understanding and possibly treating maladaptive changes in hypothalamic lipid action in obesity.

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149 2) Hypothalamic sensing of endocannabinoids: from physiology to obesity development

For centuries, marijuana (*Cannabis sativa*) has been known to stimulate food intake,
particularly for sweet and palatable food. However, the discovery of the biological mechanisms

underlying 'the munchies' started only in the 60s with the identification of  $\Delta^9$ -152 153 tetrahydrocannabinol (THC), the main psychoactive component of Cannabis sativa (26). Almost 30 years later, specific cannabinoid receptors were identified as the downstream targets of THC 154 (27, 28), which was followed by the characterisation of their endogenous lipid-derived ligands, 155 ECs (29, 30) and the enzymatic machinery necessary for ECs syntheses and degradations (31). 156 We now know that these molecular components form the endocannabinoid system, which mainly 157 158 operates to maximise the introduction, accumulation, and storage of energy substrates in the body (32). These effects are achieved by tissue-specific changes in ECs syntheses and the subsequent 159 160 activation of cannabinoid type 1 receptors (CB1) expressed in the brain and peripheral organs (23, 32, 33). From a biochemical standpoint, certain ECs, such as anandamide (AEA) and 2-161 162 arachidonoylglycerol (2-AG) are derived from the precursor arachidonic acid (AA) (33). Others, such as N-eicosapentaenoylethanolamide and N-docosahexaenoyl-ethanolamide, are 163 164 synthetized from the n-3 polyunsaturated fatty acids (PUFAs), eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), respectively (33). PUFAs, including linoleic acid and 165 166 linolenic acid, are essential fatty acids that must be obtained from the diet, and the amount and the types of PUFA from dietary intake can influence ECs biosyntheses. Hence, amongst 167 168 the several factors influencing ECs production, the availability of lipid precursors directly obtained through the diet plays a crucial role. 169

170 ECs are produced from several sources in the body, for example, the brain and peripheral tissues; their production is dependent on the global energy state of the organism. 171 172 When body energy level drops during fasting, brain ECs levels are increased to achieve energy 173 homeostasis by restoring the internal energy loss (23). ECs-mediated control of neuronal 174 functions contributes to this adaptive response. In 2010, our group characterised a transgenic mouse model bearing selective CB1 deletion in the principal forebrain (including the 175 176 hypothalamus) and sympathetic neurons (CB1-KO mice) and observed that these mutants are resistant to obesity with higher energy expenditure and enhanced brown adipose tissue (BAT) 177 thermogenesis (35). Since the SNS controls adaptive thermogenesis and BAT function (36), we 178 179 asked whether neuronal ECs actions modulate these peripheral metabolic outputs. Using chemical and surgical SNS denervation procedures, we have observed that the increased 180 181 functional activity in the BAT of CB1-KO mice results from an upregulated SNS tone (35). Likewise, Piomelli and his team have studied the phenotype of a transgenic murine model 182

183 overexpressing the presynaptic hydrolase monoacylglycerol lipase (MGL), an enzyme that 184 degrades 2-AG, in forebrain neurons. In this model, reduced 2-AG levels in the forebrain results in phenotypic changes that resemble those observed in the CB1-KO mice, including leanness, 185 elevated energy cost of activity, resistance to diet-induced obesity, and increased expression of 186 the thermogenic protein uncoupling protein 1 in the BAT (37). These two studies have set up a 187 well-accepted model whereby ECs operate in a CB1-dependent manner in the brain to modulate 188 189 whole-body energy homeostasis and thermogenesis via peripheral sympathetic 190 neurotransmission.

191 Subsequent studies have then uncovered the role of the hypothalamic circuits in mediating this top-down axis. Mice presenting specific genetic deletion of CB1 in different populations of 192 193 hypothalamic neurons, including single-minded 1 (SIM1)- or steroidogenic factor 1 (SF1)-194 expressing neurons, display increased energy expenditure and a modified sensitivity of the 195 peripheral SNS to circulating hormones, such as leptin and dietary cues (38, 39). Besides, genetic deletion of the ECs-degrading enzyme  $2\alpha/\beta$ -hydrolase domain containing 6 (ABHD6) in the 196 197 ventromedial hypothalamic nucleus (VMH), which promotes hypothalamic production of 2-AG, has been shown to provoke opposite metabolic effects compared with the prior CB1-KO model, 198 199 including impairments in adaptive thermogenesis in response to cold exposure or high-fat feeding (40). 200

201 Peripheral organs, such as the adipose tissue, also express enzymes and receptors 202 necessary for ECs syntheses and actions (34). To explore whether CB1 influences behavioural and metabolic responses in the adipose tissue, we have characterised a transgenic mouse model 203 204 with adipose-tissue-specific CB1 deletion. Notably, we observed adaptive changes in peripheral 205 SNS neurotransmission and protection from diet-induced obesity (41). Moreover, we have 206 documented that the hypophagic action of the CB1 antagonist rimonabant inhibits feeding by 207 activating visceral afferents and glutamatergic transmission in the brain stem nucleus tractus 208 solitarii (NTS) in a periphery-to-brain manner (42).

Altogether, these studies emphasise the existence of bidirectional brain-periphery signalling mechanisms, whereby ECs can act centrally and peripherally to modulate food intake and energy use through the SNS. An additional peripheral route through the vagus nerve is possibly involved in mediating brain-periphery crosstalk (43, 44). Hence, how can we

translate these empirical findings to delineate the neural basis of the onset and progression 213 214 of obesity? One should mention that brain ECs levels, particularly in the hypothalamus, vary in response to diet-induced metabolic stress. ECs act as a signal that alerts the brain 215 216 when energy reserves are low and need to be restored under physiological conditions. Paradoxically, their levels in the brain and peripheral organs remain elevated in obesity 217 (23, 45), a condition of excess energy. Several studies have reported that plasma ECs are 218 219 found at supraphysiological levels in obese individuals and positively correlate with body mass index (BMI) and several biomarkers of disease severity (23, 45). Dysregulated ECs 220 221 syntheses and degradation by the enzymatic machinery may be responsible for impaired energy balance regulation and disease development. Accordingly, ECs syntheses are 222 223 increased in association with reduced degradation in a tissue-dependent manner in obesity (46, 47), particularly in the adipose tissues (34). Besides, missense polymorphisms involving 224 225 the fatty acid amide hydrolase (FAAH), a key enzyme controlling AEA degradation, are associated with a high BMI in humans. Interestingly, the elevated circulating levels of the 226 227 ECs 2-AG and AEA in obese subjects after the exposure to rewarding food (23, 45) may augment their motivation to ingest the palatable food, which further exacerbates obesity. 228

229 Another example of how ECs are involved in the bidirectional brain-periphery crosstalk comes from a study investigating ECs levels during short-term exposure to HFD. Based on this 230 231 study, the hypothalamic 2-AG and AEA levels are transiently elevated 7 days after HFD feeding in mice, followed by a subsequent decline, with levels lower than those in chow-fed mice (48). 232 233 The initial transient elevation of the hypothalamic ECs levels is concomitant with the activation 234 peak of BAT thermogenesis due to the caloric overload (48). To reconcile the initial ECs surge in the hypothalamus, the authors acutely administered CL316,243, a  $\beta$ 3-adrenoceptor agonist that 235 stimulates BAT thermogenesis mainly via its peripheral action (49). This treatment increased 236 237 hypothalamic ECs levels substantially (48-50). These data not only reveal the existence of a 238 feedback loop linking peripheral changes in BAT function to central hypothalamic ECs levels, but also suggest that the elevated hypothalamic ECs levels in obesity may represent a 239 240 maladaptive counterregulatory mechanism that prevents excessive energy loss from the body in response to increased BAT thermogenesis. 241

Given that ECs signalling in the hypothalamus results in overfeeding and impaired SNSmediated energy dissipation, this maladaptive adipose tissue-ECs crosstalk accelerates weight gain after prolonged exposure to an energy-rich diet. However, why are the levels of hypothalamic ECs reduced after several weeks of HFD feeding when diet-induced obesity is established (48)? This could be explained by the existence of protective mechanisms that aim at hindering the upregulated peripheral EC tone (50) and, therefore, disease progression, albeit without success, since the animal continues to gain weight.

In conclusion, the dysregulated hypothalamic ECs action may cause, or be caused by, the maladaptive brain-periphery crosstalk mediated by the SNS and likely the vagus nerve in obesity. These signalling pathways dynamically contribute to the establishment and the progression of weight gain and its associated metabolic perturbations.

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#### 254 3) Bile acids: A novel hypothalamic brake on energy excess

BAs are liver-derived products of cholesterol metabolism that exert a series of metabolic functions via mainly (but not only) their signalling through the nuclear transcription factor farnesoid X receptor (FXR) and the seven-transmembrane G-protein coupled receptor G-protein bile acid-activated receptor (GPBAR)-1, also known as TGR5 (51).

A rapid elevation of hepatic BAs syntheses is observed during the transition from fasting to the fed state, which aids the absorption of ingested lipids in the gastrointestinal system (for a specific review on this subject, see (52)). Besides, BAs act as endocrine signals in the liver, the intestine, and the pancreas to modulate multiple metabolic outputs, including lipid and glucose metabolism (53), hepatic gluconeogenesis (53, 54), insulin and glucagon-like peptide-1 (GLP-1) release (53, 55), as well as mitochondrial respiration and thermogenesis in white and brown adipocytes (56-58).

Dietary supplementation of specific BAs, such as cholic acid (CA) and chenodeoxycholic acid (CDCA), prevents body weight gain and promotes weight loss significantly in diet-induced obese (DIO) mice through TGR5-mediated signalling (56, 59). These metabolic benefits are likely the result of increased thermogenesis and energy expenditure (56, 59). Indeed, circulating BAs levels correlate with energy expenditure in healthy human subjects (60) and with changes in energy substrate metabolism in obese subjects subjected to Roux-en-Y gastric bypass surgery (61). Hence, beyond their actions on nutrient absorption and glucose homeostasis, the observed
systemic elevation of BAs after dietary intake may signify positive energy balance and prompt
adaptive responses towards energy dissipation to restore energy homeostasis.

But can the brain be a possible target of BAs action? BAs are detected in several brain 275 areas, including the hypothalamus (25, 62). Under cholestasis, when the bile constituents are 276 277 accumulated in the blood due to the obstruction of bile ducts or excretory failure of hepatocytes, 278 the hypothalamic BAs level is augmented, promoting the synthesis of the hypothalamic hormone vasopressin to protect the liver from BAs-induced hepatotoxicity (63). Indirectly, BAs can 279 280 stimulate FGF15 release from the intestine, which enters the brain and activates FGF receptors in the hypothalamic AGRP/NPY neurons (64). The BAs-mediated gut-brain axis leads to 281 282 improvements in glucose tolerance that are likely mediated by changes in the peripheral 283 autonomic nervous system (64).

284 Our group has also observed that the hypothalamic BAs levels in mice change according to the nutritional state based on a fasting-refeeding paradigm experiment (25). We have inquired 285 286 whether activating TGR5 in the brain affects food intake and body weight in C57BL6 mice fed with a regular chow diet. After acute infusion of a BAs mix into the brain, we have observed a 287 288 significant reduction in food intake and the syntheses of the orexigenic peptides AgRP and NPY from the hypothalamus (25). To investigate whether hypothalamic TGR5 signalling mediates 289 290 long-term effects on body weight control, we have chronically infused a TGR5-specific, semi-291 synthetic BAs analogue into the cerebral ventricles. Coherently, food intake is transiently 292 reduced, but the prolonged activation of central BAs signalling does not lead to changes in food intake or body weight (25). Hence, BA-TGR5 signalling in the hypothalamus coordinates satiety 293 294 during the fasting-refeeding transition but is not involved in long-term body weight maintenance under physiological conditions. 295

As previously discussed, lipid-mediated homeostatic responses in the hypothalamus can be compromised in obesity. In an additional study, we have explored whether the brain BA-TGR5 axis might have a more dominant role in regulating energy balance in obesity. Using pharmacological and genetic approaches, we have observed that activating brain TGR5 signalling counteracts diet-induced obesity (DIO) progression in mice by reducing food intake and increasing energy expenditure through increased SNS activity. On the other hand, genetic downregulation of hypothalamic TGR5 actions accelerates obesity development and progression (24). Thus, the role of the hypothalamic BA-TGR5 pathway in the top-down control of body weight seems more prominent in obesity. This also implies that hypothalamic BAs sensing may be impaired in obesity, which is supported by the observation that levels of BAs species acting as TGR5 agonists in the hypothalamus are reduced in DIO mice (24).

Due to their heterogeneous structure and target specificity, BAs may confer different 307 metabolic effects through different downstream receptors. Brain infusion of the FXR agonist 308 309 GW4064 reduces the expression of tyrosine hydroxylase (TH), the rate-limiting enzyme in catecholamine synthesis, and subsequently, the sympathetic tone (65). The reduction in the 310 311 sympathetic tone might involve changes in hypothalamic neuronal activity, given that hypothalamic TH expression is reduced following brain GW4064 infusion (65). Conversely, in 312 313 agreement with our observations (24), brain infusion of tauro-lithocholic acid, a TGR5 agonist, promotes lipid metabolism and enhances the SNS tone (66). Hence, the membrane receptor 314 315 TGR5 and the nuclear receptor FXR might have opposing functions on energy balance regulation 316 in the hypothalamus in response to changes in BAs species.

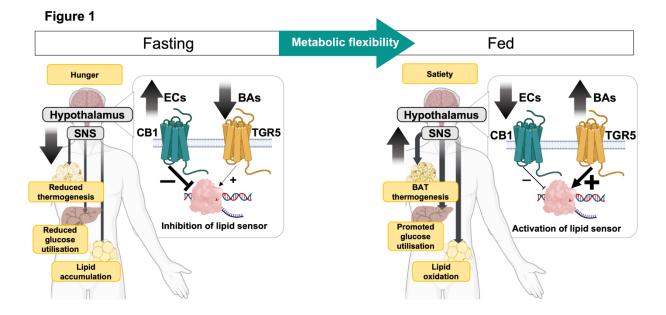
In summary, the hypothalamic BA-TGR5 pathway affects a top-down neurophysiological mechanism that leads to satiety and enhanced energy dissipation by changing peripheral SNS activity. This pathway may operate via the same hypothalamic circuits sensitive to ECs action, but with an opposite physiological goal: to counteract pathological conditions of energy excess, such as obesity.

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#### 323 4) Concluding remarks and perspectives

324 Under physiological conditions, the levels of BAs and ECs are regulated in an 325 opposite manner in the hypothalamus in response to the nutritional state. The Yin-Yang 326 regulation allows plastic changes in peripheral SNS neurotransmission and may affect metabolic flexibility between the fast-and-fed transition (Figure 1). ECs and BAs-mediated 327 328 actions in the hypothalamus may act on common intracellular lipid sensors that integrate 329 neuronal CB1 and TGR5 signalling to regulate food intake and the autonomic nervous 330 system (Figure 1). The identities of these molecular underpinnings and the neuronal circuits involved are far from being elucidated. However, addressing this knowledge gap may 331 332 provide progress towards understanding the aetiology of metabolic disorders, such as obesity. 333







336 Figure 1. Graphical representation of a possible yin-yang mechanism of energy balance 337 regulation by CB1 and TGR5 in the hypothalamus under physiological conditions. Endocannabinoids (ECs) and bile acids (BAs) might regulate whole-body metabolic flexibility 338 in the hypothalamus through CB1 and TGR5 receptors, respectively. During fasting (left), the 339 hypothalamic levels of ECs are increased, whereas BAs signalling is suppressed. This 340 341 imbalance results in hunger and possibly in modification of the sympathetic nervous system 342 (SNS) tone to impede energy storage. An opposite situation is observed after a meal (right), as hypothalamic ECs levels are reduced, whereas central BAs signalling is stimulated. This leads 343 to satiety and dissipation of excess energy by thermogenesis via upregulated SNS activity. The 344 'super lipid sensor' might integrate the inhibitory CB1 signalling and the activatory TGR5 345 signalling in the hypothalamus to facilitate energy balance regulation and metabolic flexibility. 346

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Obesity is a disease characterized by the maladaptive upregulation of the ECs tone and the concomitant downregulation of central BAs actions. Of note, the hypothalamic expression of BAs transporters exhibits plastic changes during the transition from fasting to refeeding in control lean mice but not in DIO mice (24). Thus, dysregulated hypothalamic BAs and ECs availability and sensing in the brain might contribute to obesity development, possibly causing impaired hypothalamus-SNS communication and metabolic inflexibility (Figure 2).



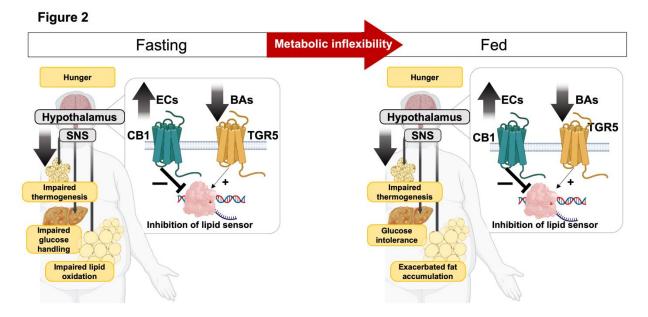




Figure 2. Graphical representation of a possible mechanism underlying maladaptive 356 357 hypothalamic CB1 and TGR5 signallings in obese pathophysiology. In obesity, ECs levels in 358 the hypothalamus are increased, while BAs levels are reduced in both the fasted and the fed state. As a result, the peripheral SNS does not cope with changes in energy availability during 359 the transition from fasting to the fed state. The transition might lead to a constant feeling of 360 361 hunger and a maladaptive drive towards excessive energy accumulation. An intracellular lipid 362 sensor might mediate alterations in this top-down axis in hypothalamic neurons. The 'super lipid sensor' might be controlled in an opposite manner by ECs and BAs-mediated signalling 363 via CB1 and TGR5, respectively. 364

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Based on these proposed models, we envision that ECs and BAs-mediated actions on 366 energy balance may be interconnected. Accordingly, peripheral administration of 2-AG 367 promotes hepatic BAs syntheses (67), suggesting that the increased levels of certain ECs 368 369 during a negative energy state during fasting may elicit BAs syntheses, perhaps to prepare 370 the body for BAs release during the upcoming meal. On the other hand, BAs availability may influence ECs production in response to changes in the body energy needs. This latter 371 hypothesis is supported by the observation that certain BAs, such as deoxycholic acid, can 372 target specific binding sites of N-acylphosphatidylethanolamine-selective phospholipase D 373

374 (NAPE-PLD) (68), the enzyme required for the syntheses of ECs-like lipid species known as
375 *N*-acylethanolamines (NAEs) (69).

376 If confirmed, these peripheral mechanisms of communication between BAs and ECs may impact the hypothalamic circuits via different routes. First, BAs (63) and ECs (70) can 377 cross the blood-brain barrier. Also, CB1- and TGR5-expressing hypothalamic neurons in 378 the ARC are ideally positioned in close contact with the fenestrated capillaries to sense 379 380 systemic changes in ECs and BAs levels. Thus, hypothalamic neurons may be capable of responding to changes in the peripheral bioavailability of these lipid mediators. 381 382 Additionally, although several peripheral organs are potential sources of ECs release in the bloodstream (71), the enzymatic machinery necessary for ECs syntheses and degradations 383 384 is also expressed in the brain (31, 72). Therefore, it is tempting to speculate that the ECs syntheses in the hypothalamic neurons may be adjusted in response to changes in BAs levels 385 386 and actions. Given that ECs are modulators of synaptic plasticity (31), modulating ECs productions may fine-tune synaptic functions to orchestrate whole body changes in energy 387 388 needs.

389 It is noteworthy that EC-like species can be highly heterogeneous in their molecular functions and physiological effects. Oleoylethanolamide (OEA), for instance, is a shorter 390 monounsaturated analogue of the endocannabinoid AEA. Unlike AEA, OEA acts 391 392 independently of the CB1 signalling pathway and can suppress appetite (33), possibly via 393 regulation of the nuclear peroxisome proliferator-activated receptor-alpha receptor (PPARa) and the G protein-coupled receptor GPR119 (33, 73). OEA can also modulate BAs 394 395 syntheses, conjugation, and transport via PPAR $\alpha$ -mediated activation (74). Moreover, the 396 OEA receptor GPR119 mediates some of the effects of BAs on gastric emptying and 397 satiation (74). Thus, the complexity may go beyond our proposed conceptual framework 398 (Figure 1) because certain canonical ECs-like molecules may operate via CB1-dependent 399 signalling in an opposite manner to BAs. In contrast, others, such as OEA, may resemble BAs-elicited physiological responses, such as satiety, through modulaiton of common 400 401 hypothalamic downstream receptors.

402 **Overall, this evidence suggests the existence of intracellular 'super lipid sensors' that** 403 **integrate the extracellular actions of multiple lipid species in the hypothalamus.** Uncovering 404 the identity of these master regulators of neuronal lipid action might have therapeutic implications, for instance, the development of novel anti-obesity pharmacological paradigms that
target specific molecular pathways in specific neuronal populations. Several advancements have
recently been made in this direction. For instance, small extracellular vesicles (sEVs) have been
used to shuttle a pharmacological inhibitor of the energy sensor AMPK in hypothalamic SF1expressing neurons in the obese murine models (75), and this approach lowers the body weight of
obese mice by sympathetic activation of BAT function (75).

411 In the past few years, we have contributed to the generation and functional validation of novel unimolecular conjugates that combine GLP-1 analogues with synthetic activators of 412 metabolic transcription factors, such as the glucocorticoid receptor or PPARs (76, 77). The GLP-413 1 moiety of these conjugates is designed to internalise the nuclear ligand in GLP-1R-expressing 414 415 cells to target organs such as the hypothalamus, where GLP-1R expression is abundant. However, organs with negligible or low GLP-1R expression are spared from these conjugates, which can 416 417 overcome the potentially toxic and off-target effects of glucocorticoids or PPARs (76, 77). Chronic treatments of obese mice with a conjugate that co-activates PPARa and PPARy in GLP-418 419 1R positive cells elicit clear-cut anti-obesity and anti-diabetic effects partly through their actions on the hypothalamus (77). Notably, this cell-specific targeting approach does not induce 420 421 cardiovascular and kidney dysfunctions associated with non-specific PPAR agonism (77). Thus, this chemical conjugation strategy combining GLP-1R agonists and nuclear-acting metabolic 422 423 hormones offers a novel therapeutic option for ameliorating obesity and its co-morbidities in a safe and cell-specific manner. 424

To tackle the growing obesity epidemic and its negative impact on health (78), there is an urgent need to decipher the neural mechanisms leading to the dysregulation of energy homeostasis. Unidentified molecular pathways might integrate the action of multiple lipidssensing mechanisms in the hypothalamus, thus contributing to disease development. Uncovering these pathways will expand our understanding of the aetiology and treatment of obesity. Towards this goal, exploring the functional interaction of ECs and BAs in the brain warrants further investigation.

432

#### 433 Author contributions

434 T.H.L. and C.Q. co-wrote the manuscript, D.C. contributed to writing and editing.

435

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# 447 **References**

Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, Powell C, Vedantam S, Buchkovich ML,
 Yang J, et al. Genetic studies of body mass index yield new insights for obesity biology. Nature.
 2015;518(7538):197-206.

451 2. Dietrich MO, Horvath TL. Hypothalamic control of energy balance: insights into the role of 452 synaptic plasticity. Trends Neurosci. 2013;36(2):65-73.

453 3. Schwartz MW, Woods SC, Porte D, Jr., Seeley RJ, Baskin DG. Central nervous system control of 454 food intake. Nature. 2000;404(6778):661-71.

Joly-Amado A, Cansell C, Denis RG, Delbes AS, Castel J, Martinez S, Luquet S. The hypothalamic
 arcuate nucleus and the control of peripheral substrates. Best Pract Res Clin Endocrinol Metab.
 2014;28(5):725-37.

458 5. Krashes MJ, Koda S, Ye C, Rogan SC, Adams AC, Cusher DS, Maratos-Flier E, Roth BL, Lowell BB.
459 Rapid, reversible activation of AgRP neurons drives feeding behavior in mice. J Clin Invest.
460 2011;121(4):1424-8.

461 6. Quarta C, Fioramonti X, Cota D. POMC Neurons Dysfunction in Diet-induced Metabolic Disease:
462 Hallmark or Mechanism of Disease? Neuroscience. 2020;447:3-14.

Garcia-Caceres C, Balland E, Prevot V, Luquet S, Woods SC, Koch M, Horvath TL, Yi CX, Chowen
JA, Verkhratsky A, et al. Role of astrocytes, microglia, and tanycytes in brain control of systemic
metabolism. Nat Neurosci. 2019;22(1):7-14.

466 8. Luquet S, Magnan C. The central nervous system at the core of the regulation of energy
467 homeostasis. Front Biosci (Schol Ed). 2009;1(2):448-65.

468 9. Mirzadeh Z, Faber CL, Schwartz MW. Central Nervous System Control of Glucose Homeostasis: A
469 Therapeutic Target for Type 2 Diabetes? Annu Rev Pharmacol Toxicol. 2022;62:55-84.

470 10. Rapoport SI, Chang MC, Spector AA. Delivery and turnover of plasma-derived essential PUFAs in
471 mammalian brain. J Lipid Res. 2001;42(5):678-85.

11. Smith QR, Nagura H. Fatty acid uptake and incorporation in brain: studies with the perfusion

473 model. J Mol Neurosci. 2001;16(2-3):167-72; discussion 215-21.

474 12. Miller JC, Gnaedinger JM, Rapoport SI. Utilization of plasma fatty acid in rat brain: distribution of
475 [14C]palmitate between oxidative and synthetic pathways. J Neurochem. 1987;49(5):1507-14.

476 13. Rapoport SI. In vivo labeling of brain phospholipids by long-chain fatty acids: relation to turnover
477 and function. Lipids. 1996;31 Suppl:S97-101.

478 14. Wang H, Eckel RH. Lipoprotein lipase in the brain and nervous system. Annu Rev Nutr.479 2012;32:147-60.

480 15. Gao Y, Vidal-Itriago A, Kalsbeek MJ, Layritz C, Garcia-Caceres C, Tom RZ, Eichmann TO, Vaz FM,

Houtkooper RH, van der Wel N, et al. Lipoprotein Lipase Maintains Microglial Innate Immunity in
Obesity. Cell Rep. 2017;20(13):3034-42.

483 16. Gao Y, Layritz C, Legutko B, Eichmann TO, Laperrousaz E, Moulle VS, Cruciani-Guglielmacci C,
484 Magnan C, Luquet S, Woods SC, et al. Disruption of Lipid Uptake in Astroglia Exacerbates Diet-Induced
485 Obesity. Diabetes. 2017;66(10):2555-63.

Laperrousaz E, Moulle VS, Denis RG, Kassis N, Berland C, Colsch B, Fioramonti X, Philippe E,
Lacombe A, Vanacker C, et al. Lipoprotein lipase in hypothalamus is a key regulator of body weight gain
and glucose homeostasis in mice. Diabetologia. 2017;60(7):1314-24.

489 18. Hundahl C, Kotzbeck P, Burm HB, Christiansen SH, Torz L, Helge AW, Madsen MP, Ratner C, Serup
490 AK, Thompson JJ, et al. Hypothalamic hormone-sensitive lipase regulates appetite and energy

491 homeostasis. Mol Metab. 2021;47:101174.

492 19. Cheng L, Yu Y, Szabo A, Wu Y, Wang H, Camer D, Huang XF. Palmitic acid induces central leptin

resistance and impairs hepatic glucose and lipid metabolism in male mice. J Nutr Biochem.

494 2015;26(5):541-8.

- 495 20. Jo YH, Su Y, Gutierrez-Juarez R, Chua S, Jr. Oleic acid directly regulates POMC neuron excitability
  496 in the hypothalamus. J Neurophysiol. 2009;101(5):2305-16.
- 497 21. Obici S, Feng Z, Arduini A, Conti R, Rossetti L. Inhibition of hypothalamic carnitine
- 498 palmitoyltransferase-1 decreases food intake and glucose production. Nat Med. 2003;9(6):756-61.
- Ross RA, Rossetti L, Lam TK, Schwartz GJ. Differential effects of hypothalamic long-chain fatty
   acid infusions on suppression of hepatic glucose production. Am J Physiol Endocrinol Metab.

501 2010;299(4):E633-9.

23. Quarta C, Mazza R, Obici S, Pasquali R, Pagotto U. Energy balance regulation by

endocannabinoids at central and peripheral levels. Trends Mol Med. 2011;17(9):518-26.

- Castellanos-Jankiewicz A, Guzman-Quevedo O, Fenelon VS, Zizzari P, Quarta C, Bellocchio L,
   Tailleux A, Charton J, Fernandois D, Henricsson M, et al. Hypothalamic bile acid-TGR5 signaling protects
   from obesity. Cell Metab. 2021;33(7):1483-92 e10.
- 507 25. Perino A, Velazquez-Villegas LA, Bresciani N, Sun Y, Huang Q, Fenelon VS, Castellanos-Jankiewicz
  508 A, Zizzari P, Bruschetta G, Jin S, et al. Central anorexigenic actions of bile acids are mediated by TGR5.
  509 Nat Metab. 2021;3(5):595-603.
- 510 26. Gaoni Y, Mechoulam R. Isolation, Structure, and Partial Synthesis of an Active Constituent of 511 Hashish. Journal of the American Chemical Society. 1964;86(8):1646-7.
- 512 27. Devane WA, Dysarz FA, 3rd, Johnson MR, Melvin LS, Howlett AC. Determination and
- characterization of a cannabinoid receptor in rat brain. Mol Pharmacol. 1988;34(5):605-13.

51428.Munro S, Thomas KL, Abu-Shaar M. Molecular characterization of a peripheral receptor for515cannabinoids. Nature. 1993;365(6441):61-5.

- 516 29. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, Gibson D, Mandelbaum A,
  517 Etinger A, Mechoulam R. Isolation and structure of a brain constituent that binds to the cannabinoid
  518 receptor. Science. 1992;258(5090):1946-9.
- 519 30. Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR, Gopher A, Almog S,
- Martin BR, Compton DR, et al. Identification of an endogenous 2-monoglyceride, present in canine gut,
   that binds to cannabinoid receptors. Biochem Pharmacol. 1995;50(1):83-90.
- 522 31. Piomelli D. The molecular logic of endocannabinoid signalling. Nat Rev Neurosci. 2003;4(11):873523 84.

524 32. Piazza PV, Cota D, Marsicano G. The CB1 Receptor as the Cornerstone of Exostasis. Neuron.
525 2017;93(6):1252-74.

S26 33. Rahman SMK, Uyama T, Hussain Z, Ueda N. Roles of Endocannabinoids and EndocannabinoidLike Molecules in Energy Homeostasis and Metabolic Regulation: A Nutritional Perspective. Annu Rev
Nutr. 2021;41:177-202.

529 34. Kuipers EN, Kantae V, Maarse BCE, van den Berg SM, van Eenige R, Nahon KJ, Reifel-Miller A,

530 Coskun T, de Winther MPJ, Lutgens E, et al. High Fat Diet Increases Circulating Endocannabinoids

531 Accompanied by Increased Synthesis Enzymes in Adipose Tissue. Front Physiol. 2018;9:1913.

53235.Quarta C, Bellocchio L, Mancini G, Mazza R, Cervino C, Braulke LJ, Fekete C, Latorre R, Nanni C,533Bucci M, et al. CB(1) signaling in forebrain and sympathetic neurons is a key determinant of

endocannabinoid actions on energy balance. Cell Metab. 2010;11(4):273-85.

535 36. Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. Physiol
 536 Rev. 2004;84(1):277-359.

Jung KM, Clapper JR, Fu J, D'Agostino G, Guijarro A, Thongkham D, Avanesian A, Astarita G,
DiPatrizio NV, Frontini A, et al. 2-arachidonoylglycerol signaling in forebrain regulates systemic energy
metabolism. Cell Metab. 2012;15(3):299-310.

540 38. Cardinal P, Bellocchio L, Guzman-Quevedo O, Andre C, Clark S, Elie M, Leste-Lasserre T, Gonzales

541 D, Cannich A, Marsicano G, et al. Cannabinoid type 1 (CB1) receptors on Sim1-expressing neurons 542 regulate energy expenditure in male mice. Endocrinology. 2015;156(2):411-8.

S43 39. Cardinal P, Bellocchio L, Clark S, Cannich A, Klugmann M, Lutz B, Marsicano G, Cota D.
S44 Hypothalamic CB1 cannabinoid receptors regulate energy balance in mice. Endocrinology.
S45 2012;153(9):4136-43.

Fisette A, Tobin S, Decarie-Spain L, Bouyakdan K, Peyot ML, Madiraju SRM, Prentki M, Fulton S,
Alquier T. alpha/beta-Hydrolase Domain 6 in the Ventromedial Hypothalamus Controls Energy
Metabolism Flexibility. Cell Rep. 2016;17(5):1217-26.

41. Ruiz de Azua I, Mancini G, Srivastava RK, Rey AA, Cardinal P, Tedesco L, Zingaretti CM, Sassmann
A, Quarta C, Schwitter C, et al. Adipocyte cannabinoid receptor CB1 regulates energy homeostasis and
alternatively activated macrophages. J Clin Invest. 2017;127(11):4148-62.

42. Bellocchio L, Soria-Gomez E, Quarta C, Metna-Laurent M, Cardinal P, Binder E, Cannich A,

Delamarre A, Haring M, Martin-Fontecha M, et al. Activation of the sympathetic nervous system
mediates hypophagic and anxiety-like effects of CB(1) receptor blockade. Proc Natl Acad Sci U S A.
2013;110(12):4786-91.

43. Gomez R, Navarro M, Ferrer B, Trigo JM, Bilbao A, Del Arco I, Cippitelli A, Nava F, Piomelli D,

Rodriguez de Fonseca F. A peripheral mechanism for CB1 cannabinoid receptor-dependent modulation
of feeding. J Neurosci. 2002;22(21):9612-7.

559 44. Berland C, Castel J, Terrasi R, Montalban E, Foppen E, Martin C, Muccioli GG, Luquet S,

560 Gangarossa G. Identification of an endocannabinoid gut-brain vagal mechanism controlling food reward 561 and energy homeostasis. Mol Psychiatry. 2022;27(4):2340-54.

562 45. Gatta-Cherifi B, Matias I, Vallee M, Tabarin A, Marsicano G, Piazza PV, Cota D. Simultaneous
563 postprandial deregulation of the orexigenic endocannabinoid anandamide and the anorexigenic peptide
564 YY in obesity. Int J Obes (Lond). 2012;36(6):880-5.

565 46. Starowicz KM, Cristino L, Matias I, Capasso R, Racioppi A, Izzo AA, Di Marzo V. Endocannabinoid
566 dysregulation in the pancreas and adipose tissue of mice fed with a high-fat diet. Obesity (Silver Spring).
567 2008;16(3):553-65.

568 47. Engeli S. Dysregulation of the endocannabinoid system in obesity. J Neuroendocrinol. 2008;20569 Suppl 1:110-5.

48. Miralpeix C, Fosch A, Casas J, Baena M, Herrero L, Serra D, Rodriguez-Rodriguez R, Casals N.
Hypothalamic endocannabinoids inversely correlate with the development of diet-induced obesity in

572 male and female mice. J Lipid Res. 2019;60(7):1260-9.

49. Richard JE, Lopez-Ferreras L, Chanclon B, Eerola K, Micallef P, Skibicka KP, Wernstedt Asterholm
I. CNS beta3-adrenergic receptor activation regulates feeding behavior, white fat browning, and body
weight. Am J Physiol Endocrinol Metab. 2017;313(3):E344-E58.

576 50. Rodriguez-Rodriguez R, Miralpeix C, Fosch A, Pozo M, Calderon-Dominguez M, Perpinya X,

577 Vellvehi M, Lopez M, Herrero L, Serra D, et al. CPT1C in the ventromedial nucleus of the hypothalamus is 578 necessary for brown fat thermogenesis activation in obesity. Mol Metab. 2019;19:75-85.

579 51. Chiang JY. Bile acids: regulation of synthesis. J Lipid Res. 2009;50(10):1955-66.

- 580 52. Staels B, Fonseca VA. Bile acids and metabolic regulation: mechanisms and clinical responses to 581 bile acid sequestration. Diabetes Care. 2009;32 Suppl 2:S237-45.
- 582 53. Kuipers F, Bloks VW, Groen AK. Beyond intestinal soap--bile acids in metabolic control. Nat Rev 583 Endocrinol. 2014;10(8):488-98.
- 584 54. Perino A, Demagny H, Velazquez-Villegas L, Schoonjans K. Molecular Physiology of Bile Acid 585 Signaling in Health, Disease, and Aging. Physiol Rev. 2021;101(2):683-731.
- 586 55. Kumar DP, Rajagopal S, Mahavadi S, Mirshahi F, Grider JR, Murthy KS, Sanyal AJ. Activation of 587 transmembrane bile acid receptor TGR5 stimulates insulin secretion in pancreatic beta cells. Biochem 588 Biophys Res Commun. 2012;427(3):600-5.
- 589 56. Watanabe M, Houten SM, Mataki C, Christoffolete MA, Kim BW, Sato H, Messaddeq N, Harney 590 JW, Ezaki O, Kodama T, et al. Bile acids induce energy expenditure by promoting intracellular thyroid 591 hormone activation. Nature. 2006;439(7075):484-9.
- 57. Broeders EP, Nascimento EB, Havekes B, Brans B, Roumans KH, Tailleux A, Schaart G, Kouach M,
  593 Charton J, Deprez B, et al. The Bile Acid Chenodeoxycholic Acid Increases Human Brown Adipose Tissue
  594 Activity. Cell Metab. 2015;22(3):418-26.
- 58. Velazquez-Villegas LA, Perino A, Lemos V, Zietak M, Nomura M, Pols TWH, Schoonjans K. TGR5
  signalling promotes mitochondrial fission and beige remodelling of white adipose tissue. Nat Commun.
  2018;9(1):245.
- 598 59. Teodoro JS, Zouhar P, Flachs P, Bardova K, Janovska P, Gomes AP, Duarte FV, Varela AT, Rolo AP,
  599 Palmeira CM, et al. Enhancement of brown fat thermogenesis using chenodeoxycholic acid in mice. Int J
  600 Obes (Lond). 2014;38(8):1027-34.
- 60. Ockenga J, Valentini L, Schuetz T, Wohlgemuth F, Glaeser S, Omar A, Kasim E, duPlessis D,
  602 Featherstone K, Davis JR, et al. Plasma bile acids are associated with energy expenditure and thyroid
  603 function in humans. J Clin Endocrinol Metab. 2012;97(2):535-42.
- 604 61. Simonen M, Dali-Youcef N, Kaminska D, Venesmaa S, Kakela P, Paakkonen M, Hallikainen M, 605 Kolehmainen M, Uusitupa M, Moilanen L, et al. Conjugated bile acids associate with altered rates of 606 glucose and lipid oxidation after Roux-en-Y gastric bypass. Obes Surg. 2012;22(9):1473-80.
- 607 62. Higashi T, Watanabe S, Tomaru K, Yamazaki W, Yoshizawa K, Ogawa S, Nagao H, Minato K, 608 Maekawa M, Mano N. Unconjugated bile acids in rat brain: Analytical method based on LC/ESI-MS/MS
- with chemical derivatization and estimation of their origin by comparison to serum levels. Steroids.
   2017;125:107-13.
- 611 63. Doignon I, Julien B, Serriere-Lanneau V, Garcin I, Alonso G, Nicou A, Monnet F, Gigou M,
- Humbert L, Rainteau D, et al. Immediate neuroendocrine signaling after partial hepatectomy through
  acute portal hyperpressure and cholestasis. J Hepatol. 2011;54(3):481-8.
- 614 64. Liu S, Marcelin G, Blouet C, Jeong JH, Jo YH, Schwartz GJ, Chua S, Jr. A gut-brain axis regulating
- 615 glucose metabolism mediated by bile acids and competitive fibroblast growth factor actions at the
- 616 hypothalamus. Mol Metab. 2018;8:37-50.

617 65. Deckmyn B, Domenger D, Blondel C, Ducastel S, Nicolas E, Dorchies E, Caron E, Charton J, Vallez 618 E, Deprez B, et al. Farnesoid X Receptor Activation in Brain Alters Brown Adipose Tissue Function via the

619 Sympathetic System. Front Mol Neurosci. 2021;14:808603.

66. Eggink HM, Tambyrajah LL, van den Berg R, Mol IM, van den Heuvel JK, Koehorst M, Groen AK,
Boelen A, Kalsbeek A, Romijn JA, et al. Chronic infusion of taurolithocholate into the brain increases fat
oxidation in mice. J Endocrinol. 2018;236(2):85-97.

67. Chanda D, Kim YH, Li T, Misra J, Kim DK, Kim JR, Kwon J, Jeong WI, Ahn SH, Park TS, et al. Hepatic
cannabinoid receptor type 1 mediates alcohol-induced regulation of bile acid enzyme genes expression
via CREBH. PLoS One. 2013;8(7):e68845.

626 68. Margheritis E, Castellani B, Magotti P, Peruzzi S, Romeo E, Natali F, Mostarda S, Gioiello A, 627 Piomelli D, Garau G. Bile Acid Recognition by NAPE-PLD. ACS Chem Biol. 2016;11(10):2908-14.

628 69. Egertova M, Simon GM, Cravatt BF, Elphick MR. Localization of N-acyl phosphatidylethanolamine
 629 phospholipase D (NAPE-PLD) expression in mouse brain: A new perspective on N-acylethanolamines as
 630 neural signaling molecules. J Comp Neurol. 2008;506(4):604-15.

631 70. Maccarrone M, Fiori A, Bari M, Granata F, Gasperi V, De Stefano ME, Finazzi-Agro A, Strom R.
632 Regulation by cannabinoid receptors of anandamide transport across the blood-brain barrier and
633 through other endothelial cells. Thromb Haemost. 2006;95(1):117-27.

Mazier W, Saucisse N, Gatta-Cherifi B, Cota D. The Endocannabinoid System: Pivotal Orchestrator
 of Obesity and Metabolic Disease. Trends Endocrinol Metab. 2015;26(10):524-37.

Nomura DK, Morrison BE, Blankman JL, Long JZ, Kinsey SG, Marcondes MC, Ward AM, Hahn YK,
Lichtman AH, Conti B, et al. Endocannabinoid hydrolysis generates brain prostaglandins that promote
neuroinflammation. Science. 2011;334(6057):809-13.

639 73. Overton HA, Babbs AJ, Doel SM, Fyfe MC, Gardner LS, Griffin G, Jackson HC, Procter MJ,

Rasamison CM, Tang-Christensen M, et al. Deorphanization of a G protein-coupled receptor for
 oleoylethanolamide and its use in the discovery of small-molecule hypophagic agents. Cell Metab.

642 2006;3(3):167-75.

643 74. Li T, Chiang JY. Regulation of bile acid and cholesterol metabolism by PPARs. PPAR Res.
644 2009;2009:501739.

645 75. Milbank E, Dragano NRV, Gonzalez-Garcia I, Garcia MR, Rivas-Limeres V, Perdomo L, Hilairet G,

Ruiz-Pino F, Mallegol P, Morgan DA, et al. Small extracellular vesicle-mediated targeting of hypothalamic
AMPKalpha1 corrects obesity through BAT activation. Nat Metab. 2021;3(10):1415-31.

648 76. Quarta C, Clemmensen C, Zhu Z, Yang B, Joseph SS, Lutter D, Yi CX, Graf E, Garcia-Caceres C,
649 Legutko B, et al. Molecular Integration of Incretin and Glucocorticoid Action Reverses Immunometabolic
650 Definition and Objective Collective Collective 2017 26(4) 620 22 cf.

by 50 Dysfunction and Obesity. Cell Metab. 2017;26(4):620-32 e6.

77. Quarta C, Stemmer K, Novikoff A, Yang B, Klingelhuber F, Harger A, Bakhti M, Bastidas-Ponce A,
Bauge E, Clemmensen C, et al. GLP-1-mediated delivery of the PPARalpha/gamma dual-agonist

653 Tesaglitazar improves obesity and glucose metabolism in mice. Research Square; 2021. DOI:

654 10.21203/rs.3.rs-1081959/v1

655 78. Collaborators GBDO, Afshin A, Forouzanfar MH, Reitsma MB, Sur P, Estep K, Lee A, Marczak L,
656 Mokdad AH, Moradi-Lakeh M, et al. Health Effects of Overweight and Obesity in 195 Countries over 25
657 Years. N Engl J Med. 2017;377(1):13-27.

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