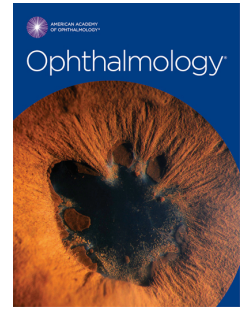


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Neuroretinal and RPE changes and susceptibility to Age-Related Macular Degeneration: insights from the longitudinal Alienor Study

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This article contains additional online-only material. The following should appear online-only: Figures S2, S4; Tables S2, S4 – S8.

Abbreviations: AMD: age-related macular degeneration, BLamDs: basal laminar deposits, BM: Bruch's membrane, CFH: complement factor H, CNV: choroidal neovascularization, ETDRS: Early Treatment Diabetes Retinopathy Study, GA: geographic atrophy, GWAS: genome-wide association study, HR: hazard ratio, GCL: ganglion cell layer, INL: inner nuclear layer, IPL: inner plexiform layer, IRL: inner retinal layers, ONL: outer nuclear layer, OPL: outer plexiform layer, ORL: retina retinal layers, PRS: polygenic risk scores, PSL: photoreceptor segment layer, RPE: retinal pigment epithelium, RNFL: retinal nerve fiber layer, SD-OCT: spectral-domain optical coherence tomography, SNP: single nucleotide polymorphism, TRL: total retinal layers.

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1 **Abstract**

2 **Purpose:** We assessed the associations of macular layer thicknesses, measured using
3 spectral-domain OCT (SD-OCT), with incident age-related macular degeneration (AMD)
4 and AMD polygenic risk scores (PRS).

5 **Design:** Population-based cohort study

6 **Participants:** 653 participants of the Alienor study, with biennial eye imaging from 2009
7 to 2024.

8 **Methods:** Macular layer thicknesses of eight distinct layers and three compound layers
9 were automatically segmented based on SD-OCT imaging of the macula. Total and
10 pathway specific PRS were calculated from previous AMD genome-wide association
11 studies summary statistics. Associations of macular layer thicknesses with incident
12 intermediate and advanced AMD were analyzed using time-dependent Cox proportional
13 hazards models. Associations of macular layer thicknesses with PRS were assessed
14 using linear mixed models.

15 **Main outcome measures:** Incident intermediate and advanced AMD based on fundus
16 colour photographs and SD-OCT.

17 **Results:** Mean age at first OCT examination of the 653 participants was 82.2 ± 4.2 years
18 and 61.3 % were women. In multivariate adjusted models, incident intermediate AMD was
19 associated with thicker retinal pigment epithelium (RPE) - Bruch's Membrane (BM)
20 complex in the 1 mm central circle (Hazard ratio (HR)= 1.13 for 1 μm increase; $P_{\text{FDR}} = 8.08$
21 $\times 10^{-4}$). Incident advanced AMD was associated with thicker RPE-BM complex in both the

22 central circle (HR= 1.09; P_{FDR} = 0.005) and the inner circle (1 mm – 3 mm) (HR= 1.28;
23 P_{FDR} = 1.61×10^{-5}).

24 Over the study period, RPE-BM complex thickening in the inner circle was more
25 pronounced in individuals with high total PRS (β = 0.06 $\mu\text{m}/\text{year}$ for 1 standard deviation
26 increase, P_{FDR} = 1.61×10^{-10}), high complement pathway PRS (β = 0.04 $\mu\text{m}/\text{year}$,
27 P_{FDR} = 3.23×10^{-5}), high lipid pathway PRS (β = 0.03 $\mu\text{m}/\text{year}$, P_{FDR} = 3.74×10^{-4}) and
28 ARMS2 (β = 0.03 $\mu\text{m}/\text{year}$, P_{FDR} = 0.002). Further, high total PRS and high complement-
29 specific PRS were associated with thinner photoreceptor segment layer (PSL) at baseline
30 and with thinning of the outer nuclear layer over the study period.

31 **Conclusion:** These results highlight the importance of RPE-BM complex thickening in
32 the pathophysiological sequence of AMD. Further longitudinal studies are needed, in
33 particular to determine the value of RPE-BM thickening and PSL thinning measured
34 using SD-OCT for the clinical follow-up of AMD patients.

35 **Introduction**

36 Age-related macular degeneration (AMD) is a complex multi-factorial disease and the
37 leading cause of vision loss among older adults in high income countries.^{1,2} Global pooled
38 data of people between 45 and 85 years estimate the worldwide prevalence of AMD at
39 8.7%.³

40 AMD is classified into early, intermediate and advanced stages based on the severity of
41 fundus lesions.⁴ Early AMD may be asymptomatic and is characterized by medium-sized
42 drusen (63-125 μm), which are clinically visible deposits of abnormal, incompletely
43 degraded material beneath the retinal pigment epithelium (RPE). Further progression to
44 intermediate AMD is marked by larger drusen ($>125 \mu\text{m}$) and pigmentary abnormalities of
45 the macula. Advanced AMD includes atrophic AMD (geographic atrophy, GA) and
46 neovascular AMD (choroidal neovascularization, CNV), both resulting in substantial visual
47 decline in most patients.

48 Over the last decades, the understanding of AMD has greatly increased. Large
49 epidemiological studies have identified risk factors such as cigarette smoking, nutritional
50 factors, cardiovascular diseases, and genetic markers.^{1,5} In 2016, a large genome-wide
51 association study (GWAS) identified 52 genetic variants at 34 loci that are associated
52 significantly with AMD. These 52 variants were used to compute a weighted polygenic risk
53 score (PRS), which was estimated to account for more than half of the genomic heritability
54 in the European-ancestry subjects.⁶ The genes at these 34 loci are primarily involved in
55 three biological pathways: the complement system, lipid metabolism, and extracellular
56 matrix (ECM) remodeling.

57 Furthermore, multimodal imaging modalities such as optical coherence tomography
58 (OCT) and fundus autofluorescence have made it possible to characterize in great detail
59 the morphological changes in the course of AMD.⁷ OCT changes in overall macular layer
60 thickness and reflectivity are thus widely used for diagnosis and monitoring of AMD and
61 other retinal and neurodegenerative diseases.⁸⁻¹⁰ However, only in recent years
62 automated OCT layer segmentation has made it possible to study in depth the changes
63 in distinct macular layers in retinal disease.¹¹⁻¹³

64 In normal aging, an overall thinning of the retina is observed, in particular due to loss of
65 neurons.¹⁴ Further, the aging RPE undergoes structural changes, including loss of
66 melanin granules, increase in the number of residual bodies and accumulation of the age
67 pigment lipofuscin.¹⁵ It is thought that these changes lead to reduced antioxidant capacity
68 of the RPE and progressive accumulation of basal laminar deposits (BLamDs) and
69 formation of drusen.^{16,17} BLamDs are histologically found between the RPE plasma
70 membrane and its basal lamina and would thus be measured as part of the RPE layer in
71 automated OCT layer segmentation.¹⁸ Even though continuous enlarging BLamDs may
72 indicate a strong risk for AMD progression and are accompanied by poor visual acuity and
73 RPE dysmorphia, eyes with early BLamDs may show normal fundus appearance and
74 visual acuity.¹⁶

75 Combined histopathological and functional studies have also evidenced that
76 photoreceptor degeneration and loss, with a preferential vulnerability of rods, occur before
77 disease progression to advanced AMD.¹⁹⁻²¹

78 Recent research showed that morphologic changes within the photoreceptors and RPE-
79 Bruch's Membrane (BM) complex may be linked to AMD and its progression. There is also

80 some evidence of thinner photoreceptor segment layer (PSL) and thicker RPE-BM in
81 individuals with high genetical susceptibility to AMD²²⁻²⁵. However, to date, there has been
82 no longitudinal analysis of the different macular layer thicknesses in a population-based
83 study in relation to AMD.

84 Thus, we analyzed the longitudinal associations of macular layer thicknesses, measured
85 using spectral-domain OCT (SD-OCT) with incident intermediate and advanced AMD and
86 AMD PRS in a population-based cohort of older adults.

87 **Methods**

88 Study cohort

89 The Alienor study is a prospective population-based cohort study. A total of 963
90 participants were recruited from 2006 through 2008 from the Bordeaux cohort of the Three
91 City (3C) study. The 3C study was initiated in 1999 and recruited individuals aged 65 years
92 or older from electoral registers. Both the Alienor study cohort and the 3C study cohort
93 have been described in detail elsewhere.^{5,26}

94 The Alienor study consists of biennial eye examinations at the Department of
95 Ophthalmology of the University Hospital of Bordeaux between 2006 and 2024
96 (<http://www.alienor-study.com/langue-english-1.html>, accessed on 19 September 2024).

97 The Alienor study followed the tenets of the Declaration of Helsinki and was approved
98 prospectively by the Ethical Committee of Bordeaux (Comité de Protection des Personnes
99 Sud-Ouest et Outre-Mer III, code 2006/10) in May 2006. Informed consent of all
100 participants of the Alienor study was obtained after explanation of the nature and possible
101 consequences of the study.

102 Ophthalmic examination, retinal imaging and macular layer segmentation

103 The initial Alienor study protocol includes visual acuity, refraction, intraocular pressure
104 and 45° high-resolution color fundus photographs (TRC NW6S; Topcon, Japan). Starting
105 from the first follow-up visit (2009-2011), SD-OCT examinations of the macula and the
106 optic nerve were included to the Alienor study protocol (Spectralis; Heidelberg
107 Engineering, Germany). The same OCT device was used at all follow-up visits.

108 For the OCT acquisitions, a standardized imaging protocol was employed. All OCT
109 assessments were performed by the same experienced technician. The Spectralis® OCT
110 provides an automatic real-time function that adjusts for eye movement and increases
111 image quality. For the macular cube acquisition, the following conditions were used:
112 resolution mode, high speed; scan angle, 20°; number of B-scans, 19; pattern size, 20° ×
113 15°; centered on the fovea and distance between B-scans, 236 μm. Single horizontal and
114 vertical B-scan images (1536 A-scans) centered on the fovea were also performed.

115 The OCT-layer segmentation was performed using the inbuilt automated segmentation
116 algorithm of the Heidelberg Eye Explorer (HEYEX 2) on the macular volume scan. The
117 HEYEX segmentation algorithm delineates the following macular layers (**Figure 1**): retinal
118 nerve fiber layer (RNFL), ganglion cell layer (GCL), inner plexiform layer (IPL), inner
119 nuclear layer (INL), outer plexiform layer (OPL), outer nuclear layer (ONL), and retinal
120 pigment epithelium (RPE). With modern SD-OCT devices, the single band previously
121 attributed to the RPE can now occasionally be seen as 2 distinctive hyperreflective bands
122 separated by a hyporeflective zone, particularly when pathology is present. We thus used
123 the term RPE-BM complex instead of RPE, consistent with recent literature.²⁷
124 Furthermore, the layers from RNFL to ONL are combined to inner retinal layers (IRL), the
125 photoreceptors and the RPE-BM complex to outer retinal layers (ORL), and all layers to
126 total retinal layers (TRL). An additional layer distinct to the above listed can be calculated
127 by subtraction of the RPE-BM complex from the ORL. This layer represents the
128 photoreceptor inner and outer segments including the interdigitation zone and we will refer
129 to it as photoreceptor segment layer (PSL) in the following. For each layer, the device
130 reports the average thickness (in μm) in nine sectors (center, superior inner, temporal

131 inner, inferior inner, nasal inner, superior outer, temporal outer, inferior outer and nasal
132 outer) of the 6 mm diameter Early Treatment Diabetes Retinopathy Study (ETDRS) grid
133 centered on the fovea. As a recent cross-sectional study showed associations between
134 macular layer thicknesses and early AMD mainly in the central (1 mm in diameter) and
135 inner circle (consisting of the superior inner, temporal inner, inferior inner and nasal inner
136 sector, 1–3 mm) of the ETDRS grid,²⁴ we focused our analysis on those two circles. In
137 addition, our macular cube is too small to assess the outer ETDRS circle. The mean
138 thickness of the inner circle was determined by calculating the average of the four inner
139 sectors: superior inner, temporal inner, inferior inner, nasal inner.

140 The accuracy and reproducibility of the inbuilt automated segmentation algorithm of the
141 HEYEX has been previously reported.²⁸

142 All available OCT data of the Alienor cohort were exported between January and April
143 2024. OCT images of participants with missing OCT data, retinal diseases and identified
144 outliers as well as signal strength lower than 15 dB were ophthalmologically reviewed for
145 segmentation quality. Acquisitions with artefacts and segmentation errors were excluded
146 from the analysis.

147 Classification of AMD

148 For fundus photography, the AMD grading scheme followed the International
149 Classification and Grading System by Bird et al., modified for drusen size, location and
150 area according to the Multi-Ethnic Study of Atherosclerosis.^{29,30} Grading was performed
151 from color fundus photographs in duplicate by two trained graders and inconsistencies
152 between the two graders were adjudicated by a retina specialist.⁵ In addition, SD-OCT
153 macular scans were interpreted for signs of advanced atrophic AMD and neovascular

154 AMD. Finally, classification of advanced AMD was performed by retina specialists, using
155 all available information (ophthalmological history and treatments, color fundus
156 photographs, SD-OCT scans).

157 To increase the comparability of this study with clinical routine data, in the present study,
158 we used the Beckman AMD classification,⁴ with intermediate AMD defined as presence
159 of large drusen and/or pigmentary abnormalities on fundus photographs, in the absence
160 of advanced AMD (atrophic and or neovascular).

161 Incidence of intermediate AMD was defined as the eye progressing from no or early AMD
162 at baseline OCT examination (2009–2010) to intermediate AMD at any time-point during
163 the study period (2009–2020). Incidence of advanced AMD was defined as the eye
164 progressing from no, early AMD or intermediate at baseline OCT examination (2009–
165 2010) to advanced AMD at any time-point during the study period (2009–2020). The date
166 of occurrence of intermediate/advanced AMD was calculated as the midpoint of the
167 interval between the last visit without intermediate /advanced AMD and the first visit with
168 intermediate /advanced AMD.

169 Covariates

170 Polygenic risk score (PRS): Genotyping was performed by the French Centre National de
171 Génotypage at 3C study baseline (1999–2001) using Illumina Human610-Quad
172 BeadChips.³¹ The present PRS is based on the weighted risk score published by Fritsche
173 and colleagues and was described in detailed elsewhere.^{6,32–34} The score corresponds to
174 the sum of the corresponding betas multiplied by the number of minor alleles for each
175 single nucleotide polymorphism (SNP). The betas used are calculated from the fully
176 conditioned odds ratios of Fritsche et al. Due to the high number of missing data for the

177 three following SNPs: TRPM3 rs71507014, CNN2 rs67538026, and MMP9 rs142450006,
178 these SNPs have been excluded from the risk score calculation. The present total PRS is
179 thus based on 49 SNPs and was calculated for all participants who had available data for
180 at least the five majors AMD-related genes (CFH rs10922109, CFH rs570618, C2
181 rs11603772, C3 rs2230199, and ARMS2 rs3750846).^{32,34} The pathway-specific PRS were
182 calculated for the three main biological pathways: C2, C3, C9, CFH, CFI and TMEM97 for
183 the complement pathway, ADAMTS9, COL4A3, COL8A1, SYN3 and VEGFA for the ECM
184 pathway and ABCA1, APOE, CETP, LIPC for the lipid pathway.³⁴ The PRS were used as
185 quantitative variables after Z score standardization.

186 Sociodemographic data, lifestyle characteristics and medical variables were collected at
187 3C baseline.²⁶

188 Medical variables including diabetes (fasting blood glucose ≥ 7 mmol/L and/or diabetes
189 medication), hypertension (blood pressure $>140/90$ mm Hg and/or antihypertensive
190 medication), and systemic lipid-lowering medications were also assessed at 3C baseline.
191 Fasting plasma lipids were measured at the Biochemistry Laboratory of the University
192 Hospital of Dijon, France using routine enzymatic techniques. Plasma esterified 3-hydroxy
193 fatty acids (3-OH FAs) were quantified as a proxy of total plasma LPS burden using liquid
194 chromatography tandem mass spectrometry (LC-tandem MS).³⁵ All plasma
195 measurements were assessed using blood samples collected during the 3C baseline visit
196 (1999–2001).

197 The Mediterranean diet score is based on MEDI-LITE score developed by Sofi et al.³⁶
198 This score is based on a 148-items validated food frequency questionnaire and reflects

199 the level of adherence to the Mediterranean diet, ranging from 0 (poor adherence) to 18
200 (highest adherence).

201 Statistical analyses

202 *Cox proportional hazards model:*

203 Associations of each macular layer thickness with incident intermediate / advanced AMD
204 were estimated using Cox proportional hazards models. The individual eye was used as
205 the unit of analysis. To take into account intra-individual correlation between eyes of one
206 participant, the “cluster” term was included in the using Cox proportional hazards models
207 in R (R Foundation for Statistical Computing, Vienna, Austria).

208 Time-dependent Cox proportional hazards model were used as an extension of the
209 standard Cox model, allowing to include the repeated OCT measurements of macular
210 layer thickness data.

211 Cox proportional hazards model was not performed for incident early AMD due to
212 insufficient power, as 23.2 % (n = 223) of the cohort already had early AMD at baseline of
213 this study.

214 All conditions for the application of the Cox model were verified. Linearity was checked
215 using penalized splines with four degrees of freedom (pspline function in the Coxph
216 function of R). The proportional hazard assumption was checked using Schoenfeld
217 residuals.

218 *Multivariate adjustments:*

219 For the Cox proportional hazards model, we used two different sets of adjustments: Model
220 1 was adjusted for age at baseline and sex. Model 2 was further adjusted for the most

221 relevant potential confounders identified from the literature and with a directed acyclic
222 graph (DAG). As some data on these variables were missing, we used multivariate
223 imputation by chained equations (MICE) assuming that missing data were Missing At
224 Random (MAR). Model 2 was thus adjusted after multivariate imputation.

225 *Linear mixed model:*

226 To study the evolution of each distinct macular layer thickness over the study period, we
227 used linear mixed models (LMM), with the eye as the unit of analysis, as described
228 previously.^{37,38} One model was fitted per layer and the outcome variable of each model
229 was the macular layer thickness by circle (central or inner). The longitudinal relationship
230 of each macular layer thickness with the fixed effects age at baseline, sex and follow-up
231 time was studied. Interactions of age at baseline with follow-up time and sex with follow-
232 up time were not used, as the Akaike Information Criterion (AIC) and Bayesian Information
233 Criterion (BIC) were favorable of the less complex models.

234 To study the longitudinal evolution of each macular layer according to the genetic risk of
235 AMD, the fixed effects age, follow-up time and sex, as well as PRS and the interactions
236 between PRS and follow-up time were included in LMMs.

237 Finally, three sensitivity analysis were performed using LMMs, the first also included AMD
238 stage at baseline (no/early/intermediate/advanced) as fixed effect, the second excluded
239 all participants with any AMD stage at baseline and the third excluded all participants with
240 intermediate or advanced AMD stage at baseline.

241 All conditions for the application of the linear mixed model were verified. The linearity of
242 quantitative variables was investigated using restricted cubic spline functions.

243 *Correction for multiple testing*

244 To address multiple testing, the Benjamini-Hochberg procedure was applied for false
245 discovery rate (FDR) correction.³⁹ The Benjamini-Hochberg procedure adjusts p-values
246 in a stepwise manner aiming to limit the proportion of false positives: After p-values are
247 assigned ranks according to their ascending value, adjusted p-values were calculated
248 using the following formula, where:

$$249 \quad p_{adjusted(i)} = \min \left(1, \min_{k \geq i} \left(\frac{m}{k} \cdot p_{(k)} \right) \right)$$

- 250 ▪ adjusted p(i) represents the adjusted p-value for the i-th ranked p-value,
- 251 ▪ p(k) represents the k-th ranked p-value and
- 252 ▪ m represents the total number of hypotheses being tested.

253 Statistical significance was determined using an FDR-corrected P value (P FDR) of less
254 than 0.05.

255 R software version 4.3.3 (R Foundation for Statistical Computing, Vienna, Austria) was
256 used for all statistical analyses.

257 **Results**

258 Characteristics of the studied sample

259 OCT examinations were included to the Alienor study protocol from 2009. Thus, the
260 studied sample comprises all participants of the Alienor study receiving at least one OCT
261 examinations between the first follow-up visit (2009-2010) and the last follow up visit in
262 February 2024. A total of 3532 OCT scans of 721 subjects were exported. Of these 344
263 OCT scans of 94 subjects were excluded due to low quality. Further 408 OCT scans of
264 174 subjects were excluded due to errors of the automatic segmentation (**Figure S2**).
265 Thus, in total 2780 OCT examinations of 653 subjects were analyzed.

266 Among the 653 included participants, mean age at first OCT examination was 82.2 years
267 (\pm 4.2) and 400 (61.3%) were female (**Table 1**). Approximately two-thirds had never
268 smoked and 15.8 % were obese (Body mass index, BMI > 30 kg/m²). Prevalent
269 hypertension was reported in 473 (72.4 %) of the participants and prevalent diabetes
270 mellitus in 46 (7.0 %). A total of 236 (36.1 %) participants received one OCT examination,
271 166 (25.4 %) received two, 109 (16.7 %) received three, 82 (12.6 %) received four and 60
272 (9.2 %) received at least five. For the 417 participants receiving more than one OCT
273 examination, the average follow-up time was 4.8 years (range 0.3 to 14.1).

274 Measurements of Macular Layer Thicknesses

275 The thickness of eight separate macular layers and three compound layer complexes was
276 measured in micrometers, including the RNFL, GCL, IPL, INL, OPL, ONL, PSL, RPE-BM
277 complex, IRL complex and ORL complex and TRL complex. The mean macular layer

278 thicknesses of the 1 mm central and the 1 mm - 3 mm inner ETDRS circle of these layers
279 and layer complexes at baseline OCT examination can be found in **Table S2**.

280 *Macular layer thicknesses and the risk of incident intermediate and advanced AMD*

281 We analyzed the associations of macular layer thicknesses with incident intermediate and
282 advanced AMD. To take into account the data of all available OCT follow-up examinations,
283 we used time-dependent proportional hazard Cox Models (**Table 3**).

284 Incident intermediate AMD: After adjustment for age and sex, we observed an association
285 of incident intermediate AMD with thicker RPE-BM complex in the 1 mm central circle
286 (Hazard ratio (HR) = 1.12 for 1 μm increase; 95% confidence interval (CI), [1.08; 1.18]; $P_{\text{FDR}} = 9.75 \times 10^{-4}$, **Table 3**). The association was similar after adjustment for AMD stage at
287 baseline, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL)
288 cholesterol, triglycerides (TG), diet quality, body mass index (BMI), lipid-lowering
289 medication, physical activity, 3-hydroxy fatty acids as a proxy of Lipopolysaccharide -type
290 endotoxins, smoking and PRS (**Table 3**). No significant association was observed of
291 incident intermediate AMD with any other macular layer, neither in the central circle, nor
292 in the inner circle.

294 Incident advanced AMD: After adjustment for age and sex, the time-dependent Cox Model
295 showed that thicker RPE-BM complex in the 1 mm central circle was associated with
296 higher risk of advanced AMD (HR = 1.16 for 1 μm increase; 95% CI, [1.10; 1.22]; $P_{\text{FDR}} =$
297 1.01×10^{-7}) (**Table 3**). The association was similar in the fully-adjusted model (**Table 3**).
298 Also, in the 1 – 3 mm inner circle, thicker RPE-BM complex was strongly associated with
299 higher risk of advanced AMD in the age and sex adjusted model (HR = 1.42 for 1 μm

300 increase; 95% CI, [1.30; 1.56]; $P_{FDR} = 7.49 \times 10^{-13}$), as well as in the fully-adjusted model
301 (HR = 1.28 for 1 μm increase; 95% CI, [1.16; 1.42]; $P_{FDR} = 1.61 \times 10^{-5}$, **Table 3, Figure**
302 **3**). This corresponds to a multiplication of the risk of advanced AMD by 3.43 for an RPE-
303 BM complex of 15 μm and by 11.8 for an RPE-BM complex of 20 μm , in comparison with
304 an RPE-BM complex of 10 μm , in the fully-adjusted model. (**Figure 3**). We observed no
305 statistically significant associations with other retinal layers.

306 Evolution of macular layers with age, sex and follow-up time

307 Using linear mixed model analysis, we analyzed the evolution of each macular layer in
308 depth, first looking into the associations with age, sex and time (**Table S4, Figure S4**).

309 In the 1 mm central circle, the four innermost macular layers differed significantly at
310 baseline according to sex, with female sex being significantly associated with thinner
311 RNFL (Coefficient Beta (β) = -1.13 μm for female sex; 95% CI, [-1.53; -0.73]; $P_{FDR} = 1.79$
312 $\times 10^{-7}$), GCL ($\beta = -1.62 \mu\text{m}$ for female sex; 95% CI, [-2.34; -0.90]; $P_{FDR} = 3.33 \times 10^{-5}$),
313 IPL ($\beta = -1.75 \mu\text{m}$ for female sex; 95% CI, [-2.33; -1.17]; $P_{FDR} = 3.12 \times 10^{-8}$) and INL (β
314 = -3.90 μm for female sex; 95% CI, [-4.83; -2.97]; $P_{FDR} = 8.88 \times 10^{-15}$). Older age at
315 baseline was significantly associated with thinner PSL at baseline ($\beta = -0.15 \mu\text{m}$ for 1 year
316 of age increase; 95% CI, [-0.20; -0.09]; $P_{FDR} = 1.32 \times 10^{-6}$).

317 In the 1 – 3 mm inner circle, older age at baseline was associated with thinner GCL ($\beta =$
318 -0.31 μm for 1 year of age increase; 95% CI, [-0.43; -0.19]; $P_{FDR} = 1.47 \times 10^{-6}$) and
319 thinner IPL ($\beta = -0.20 \mu\text{m}$ for 1 year of age increase; 95% CI, [-0.27; -0.13]; $P_{FDR} = 1.79$
320 $\times 10^{-7}$) at baseline. Again, the female sex was significantly associated with thinner INL (β
321 = -1.20 μm for female sex; 95% CI, -1.73; -0.68]; $P_{FDR} = 2.67 \times 10^{-5}$) and also with

322 thinner ONL ($\beta = -3.37 \mu\text{m}$ for female sex; 95% CI, $-4.70; -2.03$]; $P_{\text{FDR}} = 3.23 \times 10^{-6}$) at
323 baseline.

324 Macular layer thicknesses in the 1 – 3 mm inner circle all decreased significantly with
325 follow-up time, except for RNFL which increased. In the 1 mm central circle, INL and RPE-
326 BM complex increased with follow-up time, while ONL and PSL decreased.

327 AMD polygenic risk score (PRS) and macular layers

328 Using linear mixed model analysis, we then analyzed the association of each layer
329 according to their genetic risk using the PRS for AMD, adjusting for age at baseline and
330 sex (**Table S5**).

331 The AMD PRS was significantly associated with thinner PSL at baseline in the 1 mm
332 central circle ($\beta = -0.63 \mu\text{m}$ for 1 standard deviation (SD)-increase of the PRS, 95% CI, [$-$
333 $0.86; -0.40$], $P_{\text{FDR}} = 2.42 \times 10^{-6}$, **Table S5, Figure 5**) and in the 1 – 3 mm inner circle ($\beta =$
334 -0.39 , 95% CI [$-0.52; -0.26$], $P_{\text{FDR}} = 2.46 \times 10^{-7}$, **Table S5, Figure 5**), but was not
335 significantly associated with the evolution of PSL with follow-up time.

336 The AMD PRS was associated with a small but significant ONL thinning ($\beta = -0.11 \mu\text{m}/$
337 year for 1 SD-increase of the PRS, 95% CI, [$-0.16; -0.06$], $P_{\text{FDR}} = 3.08 \times 10^{-4}$, **Table S5**)
338 and RPE-BM complex thickening ($\beta = 0.06 \mu\text{m}/\text{year}$ for 1 SD-increase of the PRS, 95%
339 CI, [$0.04; 0.07$], $P_{\text{FDR}} = 1.61 \times 10^{-10}$, **Table S5**) in the 1 – 3 mm inner circle. As shown in
340 **Figure 6**, ONL was not different at baseline in participants with high (standardized PRS =
341 $+1$) or low (standardized PRS = -1) genetic susceptibility, but decreased more rapidly in
342 those with high genetic susceptibility. Regarding the RPE-BM complex, again there was
343 no significant difference at baseline between participants with high and low PRS, but RPE-

344 BM complex increased with time in those with high PRS, while it decreased in those with
345 low PRS.

346 Of note, other layers were not associated with AMD PRS, either at baseline or during
347 follow-up.

348 Three sensitivity analyses which showed similar results i) after inclusion of AMD stage
349 (no/early/intermediate/advanced) at baseline as fixed effect, ii) after exclusion of all
350 participants with any AMD at baseline and iii) after exclusion of all participants with
351 intermediate and advanced AMD at baseline (**Table S6, S7 and S8**).

352 Pathway-specific polygenic risk scores (PRS) and macular layers

353 Using linear mixed model analysis, we analyzed the genetic risk profile in more detail by
354 looking into the association of the pathway-specific PRS with the macular layers showing
355 significant associations with the AMD PRS (**Table 9**).

356 We observed a significant association of the complement pathway with ONL thinning ($\beta =$
357 $-0.16 \mu\text{m}/\text{year}$, 95% CI $[-0.21; -0.11]$, $P_{\text{FDR}} = 2.46 \times 10^{-7}$) in the 1 -3 mm inner circle, while
358 ARMS2 was associated with ONL thickening in both the 1 mm central ($\beta = 0.12 \mu\text{m}/\text{year}$,
359 95% CI $[0.03; 0.21]$, $P_{\text{FDR}} = 0.04$) and the 1 -3 mm inner circle ($\beta = 0.09 \mu\text{m}/\text{year}$, 95% CI
360 $[0.03; 0.15]$, $P_{\text{FDR}} = 0.02$, **Table 9**).

361 We also observed a significant association of the complement pathway with thinner
362 baseline PSL in the 1 mm central circle ($\beta = -0.65 \mu\text{m}$, 95 % CI $[-0.93; -0.38]$, $P_{\text{FDR}} = 4.72$
363 $\times 10^{-5}$) and in the 1 – 3 mm inner circle ($\beta = -0.40 \mu\text{m}$, 95 % CI $[-0.56; -0.25]$, $P_{\text{FDR}} = 6.66$
364 $\times 10^{-6}$). However, PSL decreased less rapidly with time in those with high complement

365 pathway PRS, both in the central ($\beta = 0.07 \mu\text{m} / \text{year}$, 95 % CI [0.03; 0.11], $P_{\text{FDR}} = 3.31 \times$
366 10^{-3}) and the inner circle ($\beta = 0.03 \mu\text{m} / \text{year}$, 95 % CI [0.01; 0.05], $P_{\text{FDR}} = 0.01$, **Table 9**).
367 We did not observe significant associations of PSL with the other pathway-specific PRS.

368 Finally, we observed a significant association of the complement pathway ($\beta = 0.04 \mu\text{m} /$
369 year , 95 % CI [0.02; 0.06], $P_{\text{FDR}} = 3.23 \times 10^{-5}$), the lipid pathway ($\beta = 0.03 \mu\text{m} / \text{year}$, 95
370 % CI [0.02; 0.05], $P_{\text{FDR}} = 3.74 \times 10^{-4}$) and ARMS2 ($\beta = 0.03 \mu\text{m} / \text{year}$, 95 % CI [0.02;
371 0.05], $P_{\text{FDR}} = 0.002$) with RPE-BM complex thickening.

372 Discussion

373 This study evidenced a strong association between thicker foveal RPE-BM complex and
374 both incident intermediate and advanced AMD. Longitudinal RPE-BM complex thickening
375 was found to be strongly associated with total AMD PRS, and specifically with the
376 complement, lipid metabolism and ARMS2 pathways PRS. Further, both the total AMD
377 PRS and the complement-specific PRS were associated with thinner PSL at baseline and
378 with longitudinal ONL thinning. Overall, our data contribute to and highlight the following
379 results in the light of known pathophysiological mechanisms and recent literature.

380 First, RPE-BM complex thickening is a hallmark of AMD progression. In aging RPE cells,
381 cumulative oxidative damage contributes to anatomical and physiological changes,
382 including the accumulation of byproducts of the visual cycle leading to the accumulation
383 of drusen (between the RPE basal lamina and Bruch's membrane).⁴⁰ Distinct of drusen,
384 BLamDs, found between the RPE plasma membrane and its basal lamina, also represent
385 early pathogenic changes in AMD.¹⁶ Thus thickening of the RPE-BM complex measured
386 using OCT possibly include drusen, BLamDs, accumulation of lipofuscin, increase in the
387 density of residual bodies and diffuse thickening of Bruch's membrane.¹⁴

388 In 2019, a cross-sectional analysis of the German population-based AugUR Study also
389 observed RPE-BM complex thickening and PSL thinning in early AMD.²⁴ Similarly, the
390 cross-sectional Portuguese Coimbra study showed that RPE-BM complex thickening and
391 PSL thinning is found in more severe early AMD.²⁵ Here, we show that RPE-BM complex
392 is associated with future development of intermediate or advanced AMD, independently
393 of potential confounders, including AMD stage at baseline.

394 Schmidt-Erfurth et al. used a machine learning–based predictive model assessing the risk
395 of conversion to advanced AMD and described the reduction of neurosensory structures,
396 including the PSL, as an important determinant in the progression to GA.⁴¹ Further, they
397 reported RPE-drusen complex thickening as a prominent predictor for the development of
398 CNV. In the present study, however, we could not distinguish atrophic and neovascular
399 AMD, due to the low number of incident advanced AMD cases.

400 A recent study by Zekavat et al., underlined the significance of early changes in the PSL,
401 using the data of the UK Biobank with a broad AMD classification and including patients
402 of an age span from 40 to 70 years.²² They reported that PSL thinning and RPE-BM
403 complex thickening were both significantly associated with increased AMD prevalence
404 and incidence. Interestingly, they observed accelerated PSL thinning starting at around
405 45 years of age, 12 years before RPE-BM complex thickening, which highlights the
406 potential pathophysiological sequence in morphological changes in the development of
407 AMD.²² This potential chronological sequence is in line with our finding of a highly
408 significant association of RPE-BM complex thickening with incident intermediate and
409 advanced AMD and could also explain the lack of finding PSL thinning, as this thinning
410 might have occurred before the baseline OCT examination of our study cohort of
411 participants aged 75 years or above at baseline.

412 Second, using repeated OCT measurements of the same individuals, we are the first to
413 report the longitudinal evolution of each distinct macular layer according to age and
414 gender in a cohort of people above the age of 75 at baseline. Using this data, we could
415 show distinct sex-related differences, e.g. thinner inner macular layers in women which is
416 in agreement with previous cross-sectional data.^{28,42}

417 Third, our results showed that the total PRS based on 49 of the 52 genetic variants
418 associated with AMD at a genome-wide significance level for AMD⁶ was associated with
419 thinner PSL at baseline. The total PRS was not associated with thicker RPE-BM complex
420 at baseline, but with longitudinal RPE-BM complex thickening, which is again in line with
421 the hypothesized pathophysiological chronological sequence of OCT-related changes by
422 Zekavat et al.²² Further, we also found an association between the total AMD PRS and
423 longitudinal ONL thinning. This seems consistent with the cross-sectional data of Brandl
424 et al. and Farinha et al. showing that ONL is decreased in participants with severe early
425 AMD.^{24,25}

426 Fourth, our results pinpoint specific mechanisms in PRS associated macular layer
427 changes. The above used total PRS was grouped into four main biological pathways by
428 an international consortium.³⁴ the complement system, lipid metabolism, ECM remodeling
429 and ARMS2. Looking into those pathway specific PRS, the complement pathway was
430 strongly associated with thinner PLS at baseline as well as longitudinal ONL thinning and
431 RPE-BM complex thickening. Even though the genetic risk of AMD is considered to be
432 linked to multiple genetic loci of small to modest effect, one of two major SNPs linked to
433 advanced AMD is a variant in the complement factor H gene (CFH).⁴³⁻⁴⁶ This variant
434 affects the protein's ability to regulate inflammation, leading to increased susceptibility to
435 chronic inflammation and damage in the retina. Associations with factor B and
436 complement component C2 genes further emphasized the critical role of the complement
437 system in the development of AMD.⁴⁷ Furthermore, systemic measurements of
438 complement activation products like C3a, C3d, and C5a have consistently shown elevated
439 levels in AMD.⁴⁸ However, using total macular thickness OCT data, a recent GWAS by

440 Gao et al. did not find an association between variants in the CFH gene within the
441 outermost circle of the ETDRS grid.⁴⁹ By contrast, another study by Zouache et al. found
442 that variants in the CFH gene were associated with perifoveal changes in macular
443 thickness on OCT.⁵⁰ Finally, the study by Zekavat et al., reported the variants
444 CFH:rs570618-T, CHF:10922109-C, CFH:rs187328863-T and ARMS2:rs3750846-C to
445 be significantly associated with PSL thinning.²²

446 Looking into the pathway specific PRS and longitudinal RPE-BM complex thickening
447 reveals that multiple players are at game, of which the complement and lipid pathways
448 seem to be strongest associated. However, in RPE-BM complex thickening, the second
449 major AMD SNP, ARMS2, is also involved. Variants in this region are known to be
450 associated with changes in mitochondrial function and increased inflammation, both
451 contributing to retinal damage.⁵¹⁻⁵³

452 Taken together, the current knowledge on morphologic and functional data indicates the
453 importance of primarily RPE-BM complex thickening and photoreceptor degeneration/loss
454 in the development of AMD. Further, similar to the normalized database of retinal nerve
455 fiber measurements as used in glaucoma clinics, the use of RPE-BM complex
456 measurements could be considered in AMD clinics. These measurements are readily
457 available in the current version of the Heidelberg Spectralis. Longitudinal change of the
458 RPE-BM complex might serve as an indicator of patients with high risk of conversion to
459 advanced AMD.

460 The strengths of the study include a large and well-defined cohort, a detailed standardized
461 ophthalmologic examination and long follow-up time. Although our prospective study

462 provides new insights into the genetic and morphologic correlation in AMD, limitations of
463 this study have to be acknowledged: The OCT protocol performed systematically in the
464 Alienor study employed 19 B-scans with an interscan distance of 236 μm , which implies
465 that macular thickness values are based on a relatively limited number of scans. However,
466 research on features of retinal diseases, in particular drusen volume and exudative retinal
467 changes, has demonstrated that less dense scans (up to an interscan distance of 240 μm)
468 do not significantly affect drusen volume measurements or impair the detection of
469 exudative retinal changes.^{8,54,55}

470 While the Heyex system's inbuilt segmentation algorithm is easily accessible, it may be
471 less accurate than newer automated segmentation algorithms, especially when handling
472 pathological alterations. To minimize the possibility of systematic misclassification of
473 macular layer thicknesses, we manually reviewed all OCT scans with quality below 15dB,
474 scans from patients with diagnosed retinal disease and any outliers (identified using linear
475 regression). To ensure reproducibility, no manual corrections were performed, and all
476 OCT scans with segmentations errors or disrupted retinal structure due to underlying
477 disease were systematically excluded. Segmentation errors included missing retinal
478 layers, overlapping or merging layers, incorrect retinal layer boundary detection,
479 segmentation gaps, erroneous inclusion of pathological features in retinal layer. However,
480 the measurement errors due to the acquisition protocol and the segmentation algorithm
481 are most probably not differential and therefore would tend to attenuate the estimated
482 associations. Future studies should strive to minimize such methodological limitations by
483 using more numerous scans and more precise algorithms.

484 The genetic risk score used in this study is based on the foundational work by Fritsche et
485 al. 2016, which identified key AMD risk SNPs.⁶ However, due to the rapid advancements
486 in genetic research, more recent studies have significantly expanded on this by providing
487 higher SNP resolution, particularly through fine-mapping techniques. These
488 advancements have enabled the identification of more specific, potentially causal variants
489 at known risk loci, especially with the inclusion of multi-ethnic cohorts.^{56,57} While these
490 recent papers offer enhanced resolution, considering the European ethnicity of
491 participants in the Alienor study, the comprehensive coverage of major AMD risk SNPs
492 and the comparable discriminatory power, the use of the current PRS score based on
493 Fritsche et al. 2016 remains justified.

494 Finally, we considered a wide range of potential cofounders but cannot exclude that some
495 residual cofounders or sources of misclassification could explain our observed
496 association between RPE-BM complex thickening and incident intermediate and
497 advanced AMD.

498 **Conclusion**

499 We found a strong association of thicker RPE-BM complex with incident intermediate and
500 advanced AMD in a population-based cohort of elderly individuals. Thickening of the RPE-
501 BM complex was also more pronounced in participants with high genetic susceptibility to
502 AMD, as well as ONL thinning and thinner PSL at baseline. These results highlight the
503 importance of RPE-BM complex thickening in the pathophysiological sequence of AMD.
504 Further longitudinal studies are needed, in particular to determine the value of RPE-BM

505 complex thickening and PSL thinning measured using SD-OCT for the clinical follow-up
506 of AMD patients.

Journal Pre-proof

507 **References**

- 508 1. Guymer RH, Campbell TG. Age-related macular degeneration. *The Lancet*.
509 2023;401(10386):1459-1472. doi:10.1016/S0140-6736(22)02609-5
- 510 2. Fleckenstein M, Keenan TDL, Guymer RH, et al. Age-related macular degeneration. *Nat Rev Dis*
511 *Primer*. 2021;7(1). doi:10.1038/s41572-021-00265-2
- 512 3. Wong WL, Su X, Li X, et al. Global prevalence of age-related macular degeneration and disease
513 burden projection for 2020 and 2040: A systematic review and meta-analysis. *Lancet Glob Health*.
514 2014;2(2):e106-e116. doi:10.1016/S2214-109X(13)70145-1
- 515 4. Ferris FL, Wilkinson CP, Bird A, et al. Clinical classification of age-related macular degeneration.
516 *Ophthalmology*. 2013;120(4):844-851. doi:10.1016/j.ophtha.2012.10.036
- 517 5. Delcourt C, Korobelnik JF, Barberger-Gateau P, et al. Nutrition and age-related eye diseases: The
518 Alienor (Antioxydants, lipides essentiels, nutrition et maladies oculaires) study. *J Nutr Health Aging*.
519 2010;14(10):854-861. doi:10.1007/s12603-010-0131-9
- 520 6. Fritsche LG, Igl W, Bailey JNC, et al. A large genome-wide association study of age-related
521 macular degeneration highlights contributions of rare and common variants. *Nat Genet*. 2016;48(2):134-
522 143. doi:10.1038/ng.3448
- 523 7. Holz FG, Sadda SVR, Staurengi G, et al. Imaging Protocols in Clinical Studies in Advanced Age-
524 Related Macular Degeneration: Recommendations from Classification of Atrophy Consensus Meetings.
525 *Ophthalmology*. 2017;124(4):464-478. doi:10.1016/j.ophtha.2016.12.002
- 526 8. Fang PP, Domdei N, Herrmann P, et al. Minimal optical coherence tomography B-scan density for
527 reliable detection of intra- and subretinal fluid in macular diseases. *Retina*. 2019;39(1):150-156.
528 doi:10.1097/IAE.0000000000001918
- 529 9. Grover S, Murthy RK, Brar VS, Chalam KV. Normative Data for Macular Thickness by High-
530 Definition Spectral-Domain Optical Coherence Tomography (Spectralis). *Am J Ophthalmol*.
531 2009;148(2):266-271. doi:10.1016/j.ajo.2009.03.006
- 532 10. Hopf S, Tüscher O, Schuster AK. Retinal OCT biomarkers and neurodegenerative diseases of the
533 central nervous system beyond Alzheimer's disease. *Ophthalmol*. 2024;121(2):93-104.
534 doi:10.1007/s00347-023-01974-7
- 535 11. Keane PA, Grossi CM, Foster PJ, et al. Optical Coherence Tomography in the UK Biobank Study –
536 Rapid Automated Analysis of Retinal Thickness for Large Population-Based Studies. Sørensen TL, ed. *PLOS*
537 *ONE*. 2016;11(10):e0164095. doi:10.1371/journal.pone.0164095
- 538 12. Arnould L, Guillemin M, Seydou A, et al. Association between the retinal vascular network and
539 retinal nerve fiber layer in the elderly: The Montrachet study. Grulkowski I, ed. *PLOS ONE*.
540 2020;15(10):e0241055. doi:10.1371/journal.pone.0241055
- 541 13. Emamverdi M, Vatanatham C, Fasih-Ahmad S, et al. Probing Deposit-Driven Age-Related Macular
542 Degeneration Via Thicknesses of Outer Retinal Bands and Choroid: ALSTAR2 Baseline. *Invest Ophthalmol*
543 *Vis Sci*. 2024;65(5):17. doi:10.1167/iovs.65.5.17

- 544 14. Bonilha V. Age and disease-related structural changes in the retinal pigment epithelium. *Clin*
545 *Ophthalmol*. Published online June 2008:413. doi:10.2147/OPHTH.S2151
- 546 15. Krohne TU, Stratmann NK, Kopitz J, Holz FG. Effects of lipid peroxidation products on
547 lipofuscinogenesis and autophagy in human retinal pigment epithelial cells. *Exp Eye Res*. 2010;90(3):465-
548 471. doi:10.1016/j.exer.2009.12.011
- 549 16. Sarks S, Cherepanoff S, Killingsworth M, Sarks J. Relationship of Basal Lamina Deposit and
550 Membranous Debris to the Clinical Presentation of Early Age-Related Macular Degeneration. *Investig*
551 *Ophthalmology Vis Sci*. 2007;48(3):968. doi:10.1167/iovs.06-0443
- 552 17. Boulton M, Dayhaw-Barker P. The role of the retinal pigment epithelium: Topographical variation
553 and ageing changes. *Eye*. 2001;15(3):384-389. doi:10.1038/eye.2001.141
- 554 18. Sura AA, Chen L, Messinger JD, et al. Measuring the Contributions of Basal Lamina Deposit and
555 Bruch's Membrane in Age-Related Macular Degeneration. *Investig Ophthalmology Vis Sci*. 2020;61(13):19.
556 doi:10.1167/iovs.61.13.19
- 557 19. Curcio CA, Medeiros NE, Millican CL. Photoreceptor loss in age-related macular degeneration.
558 *Invest Ophthalmol Vis Sci*. 1996;37(7):1236-1249.
- 559 20. Curcio CA. Photoreceptor topography in ageing and age-related maculopathy. *Eye*.
560 2001;15(3):376-383. doi:10.1038/eye.2001.140
- 561 21. Owsley C, Jackson GR, Cideciyan AV, et al. Psychophysical Evidence for Rod Vulnerability in Age-
562 Related Macular Degeneration. 2000;41(1).
- 563 22. Zekavat SM, Sekimitsu S, Ye Y, et al. Photoreceptor Layer Thinning Is an Early Biomarker for Age-
564 Related Macular Degeneration: Epidemiologic and Genetic Evidence from UK Biobank OCT Data.
565 *Ophthalmology*. 2022;129(6):694-707. doi:10.1016/j.ophtha.2022.02.001
- 566 23. Ferrara D, Silver RE, Louzada RN, Novais EA, Collins GK, Seddon JM. Optical Coherence
567 Tomography Features Preceding the Onset of Advanced Age-Related Macular Degeneration. *Investig*
568 *Ophthalmology Vis Sci*. 2017;58(9):3519. doi:10.1167/iovs.17-21696
- 569 24. Brandl C, Brücklmayer C, Günther F, et al. Retinal layer thicknesses in early age-related macular
570 degeneration: Results from the German AugUR study. *Invest Ophthalmol Vis Sci*. 2019;60(5):1581-1594.
571 doi:10.1167/iovs.18-25332
- 572 25. Farinha C, Silva AL, Coimbra R, et al. Retinal layer thicknesses and neurodegeneration in early
573 age-related macular degeneration: insights from the Coimbra Eye Study. *Graefes Arch Clin Exp*
574 *Ophthalmol*. 2021;259(9):2545-2557. doi:10.1007/s00417-021-05140-0
- 575 26. Alperovitch A, Amouyel P, Dartigues JF, et al. Vascular Factors and Risk of Dementia: Design of
576 the Three-City Study and Baseline Characteristics of the Study Population. *Neuroepidemiology*.
577 2003;22(6):316-325. doi:10.1159/000072920
- 578 27. Staurengi G, Sadda S, Chakravarthy U, Spaide RF. Proposed lexicon for anatomic landmarks in
579 normal posterior segment spectral-domain optical coherence tomography: The IN•OCT consensus.
580 *Ophthalmology*. 2014;121(8):1572-1578. doi:10.1016/j.ophtha.2014.02.023

- 581 28. Mauschitz MM, Holz FG, Finger RP, Breteler MMB. Determinants of Macular Layers and Optic
582 Disc Characteristics on SD-OCT: The Rhineland Study. *Transl Vis Sci Technol.* 2019;8(3):34.
583 doi:10.1167/tvst.8.3.34
- 584 29. Bird AC, Bressler NM, Bressler SB, et al. An international classification and grading system for
585 age-related maculopathy and age-related macular degeneration. *Surv Ophthalmol.* 1995;39(5):367-374.
586 doi:10.1016/S0039-6257(05)80092-X
- 587 30. Klein R, Klein BEKK, Knudtson MD, et al. Prevalence of age-related macular degeneration in 4
588 racial/ethnic groups in the multi-ethnic study of atherosclerosis. *Ophthalmology.* 2006;113(3):373-380.
589 doi:10.1016/j.ophtha.2005.12.013
- 590 31. Lambert JC, Heath S, Even G, et al. Genome-wide association study identifies variants at CLU and
591 CR1 associated with Alzheimer's disease. *Nat Genet.* 2009;41(10):1094-1099. doi:10.1038/ng.439
- 592 32. Merle BMJ, Cougnard-Grégoire A, Korobelnik JF, et al. Plasma Lutein, a Nutritional Biomarker for
593 Development of Advanced Age-Related Macular Degeneration: The Alienor Study. *Nutrients.*
594 2021;13(6):2047. doi:10.3390/nu13062047
- 595 33. De Breuk A, Acar IE, Kersten E, et al. Development of a Genotype Assay for Age-Related Macular
596 Degeneration. *Ophthalmology.* 2021;128(11):1604-1617. doi:10.1016/j.ophtha.2020.07.037
- 597 34. Colijn JM, Meester-Smoor M, Verzijden T, et al. Genetic Risk, Lifestyle, and Age-Related Macular
598 Degeneration in Europe. *Ophthalmology.* 2021;128(7):1039-1049. doi:10.1016/j.ophtha.2020.11.024
- 599 35. Pais de Barros JP, Gautier T, Sali W, et al. Quantitative lipopolysaccharide analysis using
600 HPLC/MS/MS and its combination with the limulus amoebocyte lysate assay. *J Lipid Res.* 2015;56(7):1363-
601 1369. doi:10.1194/jlr.D059725
- 602 36. Sofi F, Dinu M, Pagliai G, Marcucci R, Casini A. Validation of a literature-based adherence score to
603 Mediterranean diet: the MEDI-LITE score. *Int J Food Sci Nutr.* 2017;68(6):757-762.
604 doi:10.1080/09637486.2017.1287884
- 605 37. Gayraud L, Mortamais M, Schweitzer C, et al. Association of long-term exposure to ambient air
606 pollution with retinal neurodegeneration: the prospective Alienor study. *Environ Res.*
607 2023;232(May):116364. doi:10.1016/j.envres.2023.116364
- 608 38. Larsen PP, Feart C, Pais De Barros JP, et al. Association of lipopolysaccharide-type endotoxins
609 with retinal neurodegeneration: the Alienor study. *Ophthalmol Sci.* Published online August
610 2024:100610. doi:10.1016/j.xops.2024.100610
- 611 39. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful
612 Approach to Multiple Testing. *J R Stat Soc Ser B Stat Methodol.* 1995;57(1):289-300. doi:10.1111/j.2517-
613 6161.1995.tb02031.x
- 614 40. Johnson LV, Leitner WP, Staples MK, Anderson DH. Complement Activation and Inflammatory
615 Processes in Drusen Formation and Age Related Macular Degeneration. *Exp Eye Res.* 2001;73(6):887-896.
616 doi:10.1006/exer.2001.1094

- 617 41. Schmidt-Erfurth U, Waldstein SM, Klmscha S, et al. Prediction of Individual Disease Conversion in
618 Early AMD Using Artificial Intelligence. *Investig Ophthalmology Vis Sci*. 2018;59(8):3199.
619 doi:10.1167/iovs.18-24106
- 620 42. Koh VT, Tham YC, Cheung CY, et al. Determinants of Ganglion Cell–Inner Plexiform Layer
621 Thickness Measured by High-Definition Optical Coherence Tomography. *Investig Ophthalmology Vis Sci*.
622 2012;53(9):5853. doi:10.1167/iovs.12-10414
- 623 43. Edwards AO, Ritter R, Abel KJ, Manning A, Panhuysen C, Farrer LA. Complement Factor H
624 Polymorphism and Age-Related Macular Degeneration. *Science*. 2005;308(5720):421-424.
625 doi:10.1126/science.1110189
- 626 44. Klein RJ, Zeiss C, Chew EY, et al. Complement Factor H Polymorphism in Age-Related Macular
627 Degeneration. *Science*. 2005;308(5720):385-389. doi:10.1126/science.1109557
- 628 45. Haines JL, Hauser MA, Schmidt S, et al. Complement factor H variant increases the risk of age-
629 related macular degeneration. *Science*. 2005;308(5720):419-421. doi:10.1126/science.1110359
- 630 46. Hageman GS, Anderson DH, Johnson LV, et al. A common haplotype in the complement
631 regulatory gene factor H (HF1/CFH) predisposes individuals to age-related macular degeneration. *Proc*
632 *Natl Acad Sci*. 2005;102(20):7227-7232. doi:10.1073/pnas.0501536102
- 633 47. Gold B, Merriam JE, Zernant J, et al. Variation in factor B (BF) and complement component 2 (C2)
634 genes is associated with age-related macular degeneration. *Nat Genet*. 2006;38(4):458-462.
635 doi:10.1038/ng1750
- 636 48. Kersten E, Paun CC, Schellevis RL, et al. Systemic and ocular fluid compounds as potential
637 biomarkers in age-related macular degeneration. *Surv Ophthalmol*. 2018;63(1):9-39.
638 doi:10.1016/j.survophthal.2017.05.003
- 639 49. Gao XR, Huang H, Kim H. Genome-wide association analyses identify 139 loci associated with
640 macular thickness in the UK Biobank cohort. *Hum Mol Genet*. 2019;28(7):1162-1172.
641 doi:10.1093/hmg/ddy422
- 642 50. Zouache MA, Bennion A, Hageman JL, Pappas C, Richards BT, Hageman GS. Macular retinal
643 thickness differs markedly in age-related macular degeneration driven by risk polymorphisms on
644 chromosomes 1 and 10. *Sci Rep*. 2020;10(1):21093. doi:10.1038/s41598-020-78059-x
- 645 51. Rivera A, Fisher SA, Fritsche LG, et al. Hypothetical LOC387715 is a second major susceptibility
646 gene for age-related macular degeneration, contributing independently of complement factor H to
647 disease risk. *Hum Mol Genet*. 2005;14(21):3227-3236. doi:10.1093/hmg/ddi353
- 648 52. Delcourt C, Delyfer MN, Rougier MB, et al. ARMS2 A69S Polymorphism and the Risk for Age-
649 Related Maculopathy: The ALIENOR Study. *Arch Ophthalmol*. 2012;130(8):1077.
650 doi:10.1001/archophthalmol.2012.420
- 651 53. Jakobsdottir J, Conley YP, Weeks DE, Mah TS, Ferrell RE, Gorin MB. Susceptibility Genes for Age-
652 Related Maculopathy on Chromosome 10q26. *Am J Hum Genet*. 2005;77(3):389-407.
653 doi:10.1086/444437

- 654 54. Garzone D, Terheyden JH, Morelle O, et al. Comparability of automated drusen volume
655 measurements in age-related macular degeneration: a MACUSTAR study report. *Sci Rep.* 2022;12(1):1-
656 10. doi:10.1038/s41598-022-26223-w
- 657 55. Thiele S, Nadal J, Fleckenstein M, et al. Longitudinal Analysis of Drusen Volume in Intermediate
658 Age-Related Macular Degeneration Using Two Spectral-Domain Optical Coherence Tomography Scan
659 Patterns. *Ophthalmologica.* 2018;239(2-3):110-120. doi:10.1159/000485260
- 660 56. He W, Han X, Ong JS, et al. Genome-Wide Meta-analysis Identifies Risk Loci and Improves Disease
661 Prediction of Age-Related Macular Degeneration. *Ophthalmology.* 2024;131(1):16-29.
662 doi:10.1016/j.ophtha.2023.08.023
- 663 57. Han X, Gharahkhani P, Mitchell P, Liew G, Hewitt AW, MacGregor S. Genome-wide meta-analysis
664 identifies novel loci associated with age-related macular degeneration. *J Hum Genet.* 2020;65(8):657-
665 665. doi:10.1038/s10038-020-0750-x
- 666

667 **Figure Legends**

668

669 **Figure 1.** Example of an auto- segmented spectral domain-OCT scan using the inbuilt
 670 automated segmentation of the Heidelberg Eye Explorer.

671 RNFL: retinal nerve fiber layer, GCL: ganglion cell layer, IPL: inner plexiform layer, INL: inner
 672 nuclear layer, OPL: outer plexiform layer, ONL: outer nuclear layer, PSL: photoreceptor segment
 673 layer, RPE-BM: retinal pigment epithelium- Bruch's Membrane complex.

674

675 **Figure 3.** Association between Retinal Pigment Epithelium (RPE) - Bruch's Membrane (BM)
 676 complex thickness within the 1 – 3 mm inner circle of the ETDRS grid and incident advanced
 677 AMD by eye, using time-dependent proportional hazards Cox Model, along with exemplary OCT
 678 photos of participants with an RPE - BM complex thickness of 10 μm , 15 μm and 20 μm at
 679 baseline. The model was adjusted for age, sex, high-density (HDL) cholesterol, low-density
 680 lipoprotein (LDL) cholesterol, triglycerides (TG), diet quality, body mass index (BMI), lipid-
 681 lowering medication, physical activity, 3-hydroxy fatty acids as a proxy of Lipopolysaccharide -
 682 type endotoxins, smoking status, polygenic risk score and AMD stage at baseline. AMD stage at
 683 baseline was categorized in 3 categories (no/early/intermediate). The hazard function is
 684 represented using p-spline with 4 degrees of freedom. Exemplary OCT images of participants
 685 with an RPE-BM complex thickness of 10 μm , 15 μm and 20 μm are shown. Data from the
 686 Alienor Study, 2009–2020.

687 HR: hazard ratio.

688

689 **Figure 5.** The evolution of photoreceptor segment layer (PSL) thinning over the follow-up time for
 690 both the 1 mm central and the 1 – 3 mm inner circle of the Early Treatment Diabetic Retinopathy
 691 Study (ETDRS) grid as predicted by linear mixed models (adjusted for age, sex and follow-up
 692 time) shown for the longitudinal relationship with standardized total (PRS) for AMD. The PRS was
 693 standardized using the formula $Z = (x-\mu)/\sigma$, where Z is the standard score, μ the mean and σ the
 694 standard deviation, respectively. Data from the Alienor Study, 2009–2024.

695

696 **Figure 6.** The evolution of the outer nuclear layer (ONL) and the retinal pigment epithelium (RPE)
 697 - Bruch's Membrane (BM) complex in the 1 – 3 mm inner circle of the Early Treatment Diabetic
 698 Retinopathy Study (ETDRS) grid over the follow-up time as predicted by linear mixed models
 699 (adjusted for age, sex and follow-up time) shown for the longitudinal relationship with standardized
 700 total polygenic risk score (PRS) for AMD. The PRS was standardized using the formula $Z = (x-$
 701 $\mu)/\sigma$, where Z is the standard score, μ the mean and σ the standard deviation, respectively. Data
 702 from the Alienor Study, 2009–2024.

Table 1. Baseline Characteristics of the Study Population (n = 653), Data from the Alienor Study.

Characteristics	Mean \pm SD or n (%)
Age (years)	82.2 \pm 4.2
Sex	
Male	253 (38.7 %)
Female	400 (61.3 %)
Smoking (pack-years)	
Never smoked	424 (64.9 %)
< 20	122 (18.7 %)
\geq 20	107 (16.4 %)
Body mass index (BMI)	
[0,18.5)	3 (0.5 %)
[18.5,25)	260 (39.8 %)
[25,30)	287 (44.0 %)
[30,59]	103 (15.8 %)
Plasma lipids (mmol/L)	
LDL cholesterol	3.6 \pm 0.8
HDL cholesterol	1.6 \pm 0.4
Triglycerides	1.2 \pm 0.6
Diabetes	46 (7.0 %)
Hypertension	473 (72.4 %)
Cardiovascular Diseases	57 (8.7 %)
Mediterranean diet score	10.6 \pm 2.0

LDL: low-density lipoprotein, HDL: high-density lipoprotein;
SD: standard deviation.

Table 3. Associations between the macular layer thicknesses and incidence of intermediate and advanced AMD. Alienor study, 2009–2020.

		Intermediate AMD (n = 103)				Advanced AMD (n = 39)			
		Central circle of the ETDRS grid		Inner circle of the ETDRS grid		Central circle of the ETDRS grid		Inner circle of the ETDRS grid	
		HR (95% CI)	P _{FDR}	HR (95% CI)	P _{FDR}	HR (95% CI)	P _{FDR}	HR (95% CI)	P _{FDR}
RNFL	Model 1	1.03 [0.97; 1.10]	0.62	1.00 [0.95; 1.05]	0.99	1.09 [0.97; 1.21]	0.53	1.06 [0.98; 1.15]	0.53
	Model 2	1.04 [0.97; 1.11]	0.60	1.00 [0.95; 1.06]	0.99	1.05 [0.93; 1.19]	0.69	1.04 [0.96; 1.13]	0.60
GCL	Model 1	1.01 [0.98; 1.04]	0.83	1.02 [0.99; 1.06]	0.53	1.03 [0.97; 1.09]	0.60	1.01 [0.96; 1.06]	0.83
	Model 2	1.02 [0.98; 1.05]	0.67	1.02 [0.99; 1.05]	0.54	1.02 [0.95; 1.08]	0.83	1.02 [0.97; 1.07]	0.73
IPL	Model 1	1.01 [0.96; 1.06]	0.92	1.03 [0.98; 1.08]	0.58	1.04 [0.96; 1.12]	0.60	1.00 [0.93; 1.08]	0.99
	Model 2	1.01 [0.96; 1.07]	0.83	1.03 [0.98; 1.08]	0.60	1.02 [0.95; 1.11]	0.75	1.02 [0.95; 1.10]	0.79
INL	Model 1	1.01 [0.98; 1.03]	0.74	0.99 [0.94; 1.04]	0.89	1.04 [1.01; 1.08]	0.13	1.03 [0.95; 1.12]	0.73
	Model 2	1.01 [0.98; 1.04]	0.60	0.98 [0.93; 1.03]	0.69	1.02 [0.98; 1.07]	0.60	1.01 [0.92; 1.11]	0.95
OPL	Model 1	1.00 [0.97; 1.03]	0.99	0.97 [0.93; 1.01]	0.55	0.99 [0.94; 1.06]	0.95	1.00 [0.93; 1.08]	0.99
	Model 2	1.00 [0.98; 1.03]	0.94	0.97 [0.93; 1.02]	0.55	1.00 [0.94; 1.06]	0.99	0.99 [0.92; 1.06]	0.90
ONL	Model 1	1.01 [1.00; 1.03]	0.54	1.01 [0.99; 1.04]	0.60	0.98 [0.95; 1.01]	0.60	0.95 [0.92; 0.99]	0.06
	Model 2	1.01 [0.99; 1.03]	0.60	1.01 [0.99; 1.03]	0.60	0.98 [0.95; 1.01]	0.53	0.96 [0.93; 0.99]	0.06
PSL	Model 1	0.97 [0.93; 1.03]	0.60	0.99 [0.90; 1.09]	0.94	0.93 [0.85; 1.03]	0.53	1.17 [1.04; 1.31]	0.07
	Model 2	0.97 [0.92; 1.03]	0.62	0.99 [0.89; 1.10]	0.95	0.96 [0.89; 1.04]	0.60	1.20 [1.02; 1.40]	0.14
RPE-BM	Model 1	1.12 [1.06; 1.18]	9.75 x 10⁻⁴	1.09 [0.97; 1.22]	0.53	1.16 [1.10; 1.22]	1.01 x 10⁻⁷	1.42 [1.30; 1.56]	7.49 x 10⁻¹³
	Model 2	1.13 [1.06; 1.20]	8.08 x 10⁻⁴	1.09 [0.97; 1.23]	0.53	1.09 [1.04; 1.14]	0.005	1.28 [1.16; 1.42]	1.61 x 10⁻⁵

*For the inner circle, the thickness was calculated as the average of the nasal/temporal/superior/inferior subfields.

^aModel 1, HR was estimated using Cox proportional model adjusted for age at baseline and sex. HR for 1 µm increase of the respective layer. ^bModel 2, HR was estimated using Cox proportional model adjusted for age, sex, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL) cholesterol, triglycerides (TG), diet quality, body mass index (BMI), lipid-lowering medication, physical activity, 3-hydroxy fatty acids as a proxy of Lipopolysaccharide -type endotoxins, smoking, polygenic risk score and AMD stage at baseline. For intermediate AMD, AMD stage at baseline was in 2 categories (no/early), for advanced AMD, it was in 3 categories (no/early/intermediate). HR for 1 µm increase of the respective layer. ETDRS: Early Treatment Diabetic Retinopathy Study, CI: Confidence interval, FDR: false discovery rate, HR, hazard ratio, RNFL: retinal nerve fiber layer, GCL: ganglion cell layer, IPL: inner plexiform layer, INL: inner nuclear layer, OPL: outer plexiform layer, ONL: outer nuclear layer, PSL: photoreceptor segment layer, RPE-BM: retinal pigment epithelium- Bruch's Membrane complex.

Table 9. Longitudinal change of ONL, PSL and RPE-BM complex according to pathway-specific polygenic risk scores for AMD, Alienor study, 2009–2024. (n = 2780 examinations of 653 participants)

		Central Circle of the ETDRS Grid		Inner Circle of the ETDRS Grid*	
		β [95% CI]	P_{FDR}	β [95% CI]	P_{FDR}
ONL	Complement Pathway PRS**	0.04 [-0.89; 0.97]	0.99	0.11 [-0.62; 0.83]	0.94
	Complement Pathway PRS * Time	-0.09 [-0.17; -0.01]	0.12	-0.16 [-0.21; -0.11]	2.46 x 10⁻⁷
	Extracellular Matrix Pathway PRS***	0.22 [-0.70; 1.14]	0.88	0.11 [-0.61; 0.83]	0.94
	Extracellular Matrix Pathway PRS * Time	0.02 [-0.06; 0.10]	0.84	-0.03 [-0.08; 0.02]	0.53
	Lipid Pathway PRS****	-0.30 [-1.22; 0.62]	0.81	0.06 [-0.66; 0.78]	0.97
	Lipid Pathway PRS * Time	-0.02 [-0.10; 0.06]	0.88	-0.02 [-0.07; 0.03]	0.72
	ARMS2	-0.16 [-1.09; 0.78]	0.94	-0.16 [-0.88; 0.57]	0.91
	ARMS2 * Time	0.12 [0.03; 0.21]	0.04	0.09 [0.03; 0.15]	0.02
PSL	Complement Pathway PRS**	-0.65 [-0.93; -0.38]	4.72 x 10⁻⁵	-0.40 [-0.56; -0.25]	6.66 x 10⁻⁶
	Complement Pathway PRS * Time	0.07 [0.03; 0.11]	3.31 x 10⁻³	0.03 [0.01; 0.05]	0.01
	Extracellular Matrix Pathway PRS***	-0.17 [-0.45; 0.11]	0.50	-0.04 [-0.19; 0.12]	0.88
	Extracellular Matrix Pathway PRS * Time	-0.02 [-0.06; 0.02]	0.53	-0.002 [-0.02; 0.02]	0.94
	Lipid Pathway PRS****	-0.18 [-0.46; 0.09]	0.45	-0.11 [-0.27; 0.04]	0.36
	Lipid Pathway PRS * Time	0.01 [-0.03; 0.05]	0.94	-0.01 [-0.03; 0.01]	0.69
	ARMS2	-0.22 [-0.50; 0.06]	0.34	-0.16 [-0.31; 0.00]	0.16
	ARMS2 * Time	-0.02 [-0.07; 0.02]	0.56	-0.02 [-0.04; 0.01]	0.39
RPE-BM	Complement Pathway PRS**	-0.10 [-0.35; 0.15]	0.69	-0.21 [-0.38; -0.05]	0.05
	Complement Pathway PRS * Time	0.05 [0.004; 0.09]	0.11	0.04 [0.02; 0.06]	3.23 x 10⁻⁵
	Extracellular Matrix Pathway PRS***	0.002 [-0.24; 0.25]	0.99	-0.02 [-0.18; 0.14]	0.94
	Extracellular Matrix Pathway PRS * Time	-0.01 [-0.05; 0.03]	0.93	0.002 [-0.01; 0.02]	0.94
	Lipid Pathway PRS****	0.004 [-0.24; 0.25]	0.99	-0.09 [-0.25; 0.07]	0.56
	Lipid Pathway PRS * Time	-0.001 [-0.04; 0.04]	0.99	0.03 [0.02; 0.05]	3.74 x 10⁻⁴
	ARMS2	0.18 [-0.06; 0.43]	0.34	0.15 [-0.02; 0.31]	0.23
	ARMS2 * Time	0.003 [-0.04; 0.05]	0.99	0.03 [0.02; 0.05]	0.002

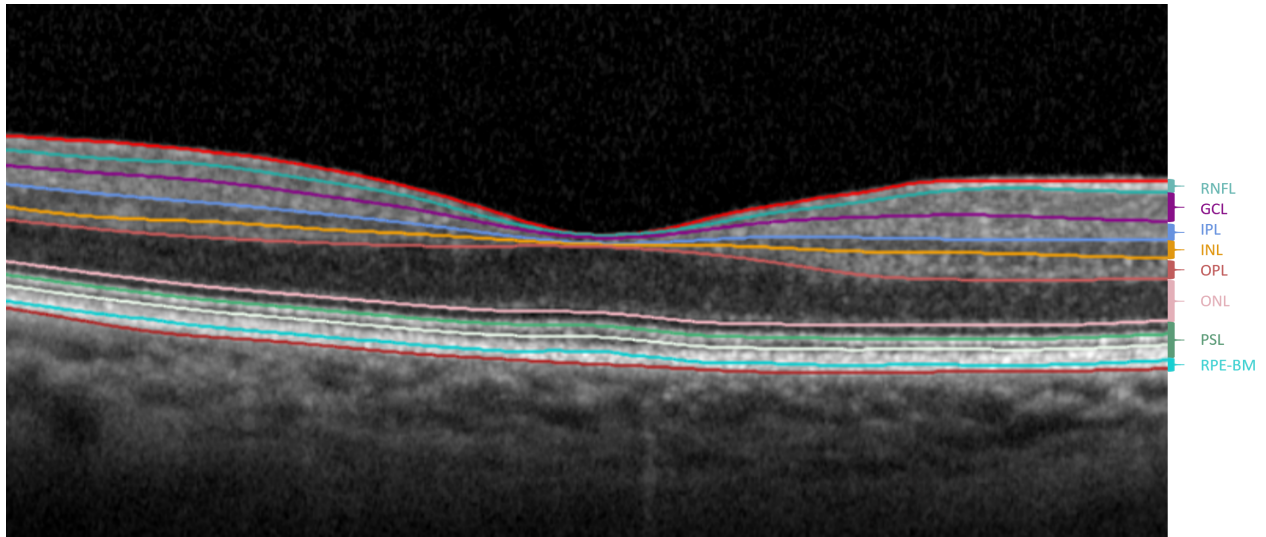
Mixed linear models are adjusted for age, sex and follow-up time. β in μm for 1-SD increase in PRS and in $\mu\text{m} / \text{year}$ for 1-SD increase in PRS for the interaction with follow-up time. ETDRS: Early Treatment Diabetes Retinopathy Study, CI: confidence interval, FDR: false discovery rate, SD: standard deviation, ONL: outer nuclear layer, PSL: photoreceptor segment layer, RPE-BM: retinal pigment epithelium- Bruch's Membrane complex, SD: standard deviation.

*For the inner circle, the thickness was calculated as the average of the nasal/temporal/superior/inferior subfields.

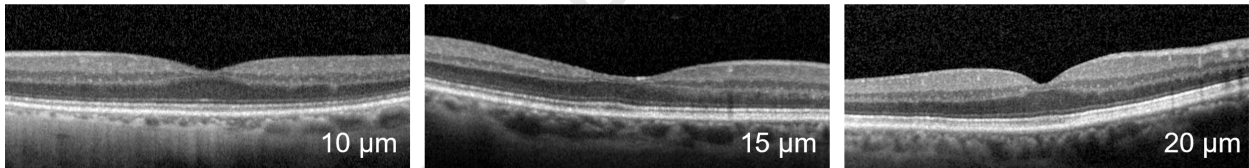
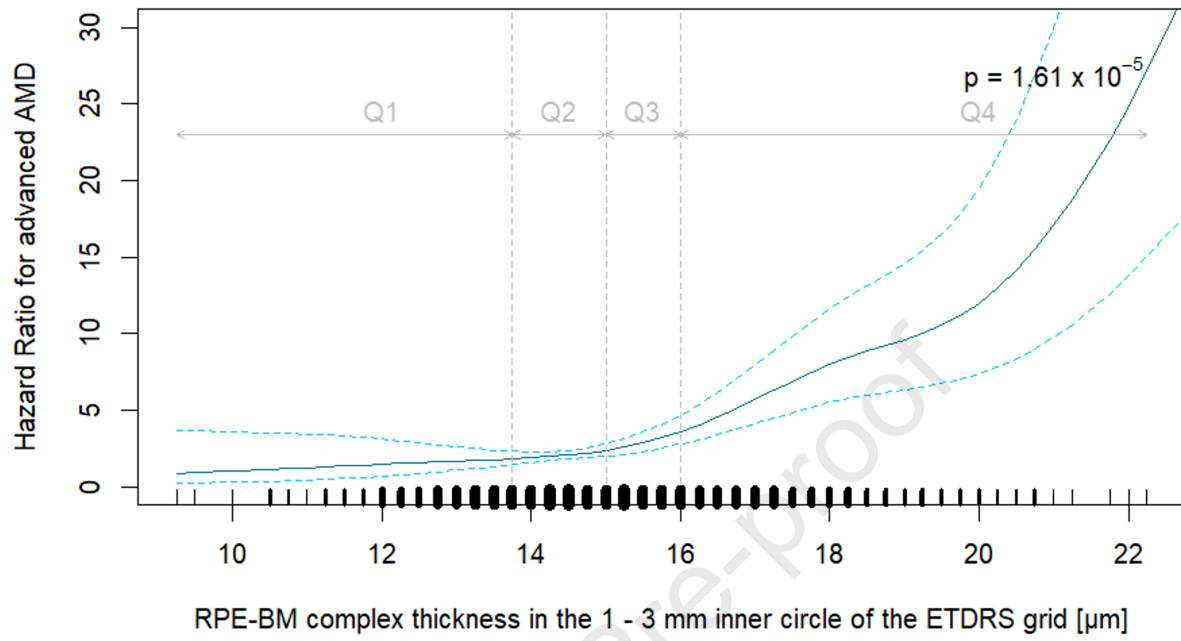
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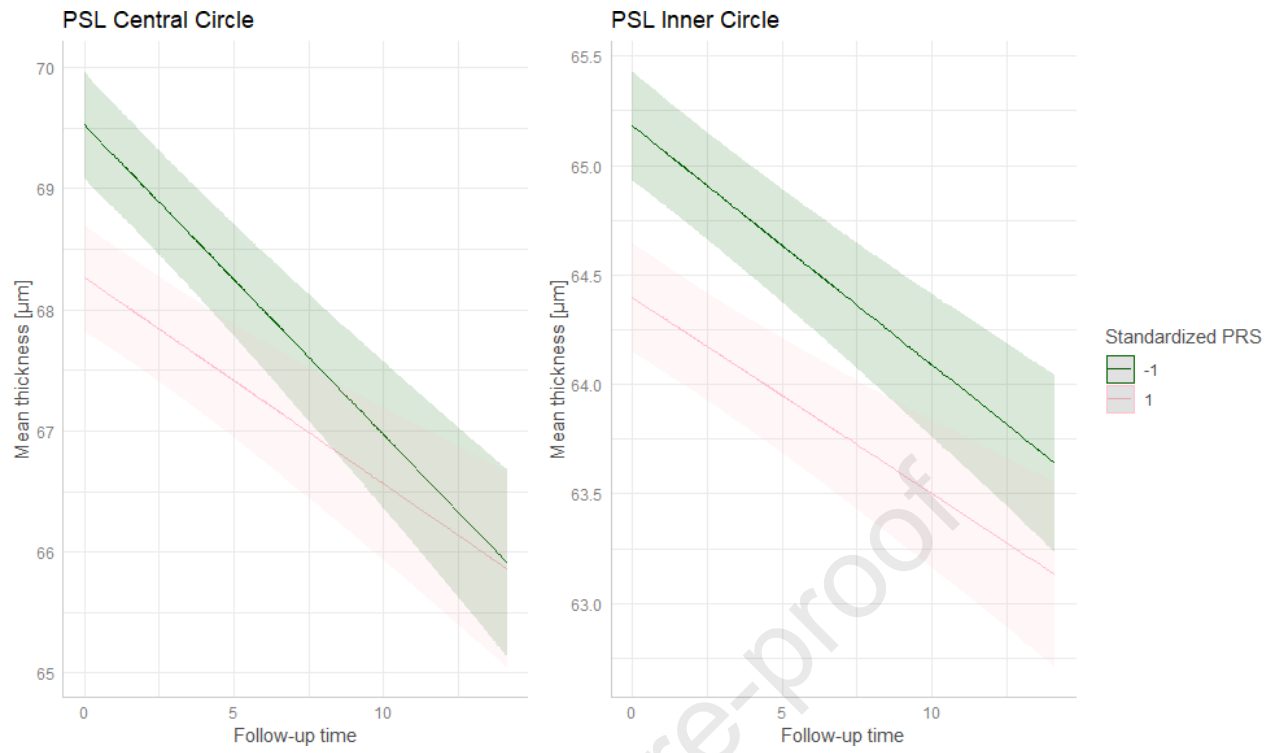
***The Extracellular Matrix Pathway is based on the following genes: ADAMTS9, COL4A3, COL8A1, SYN3, VEGFA.

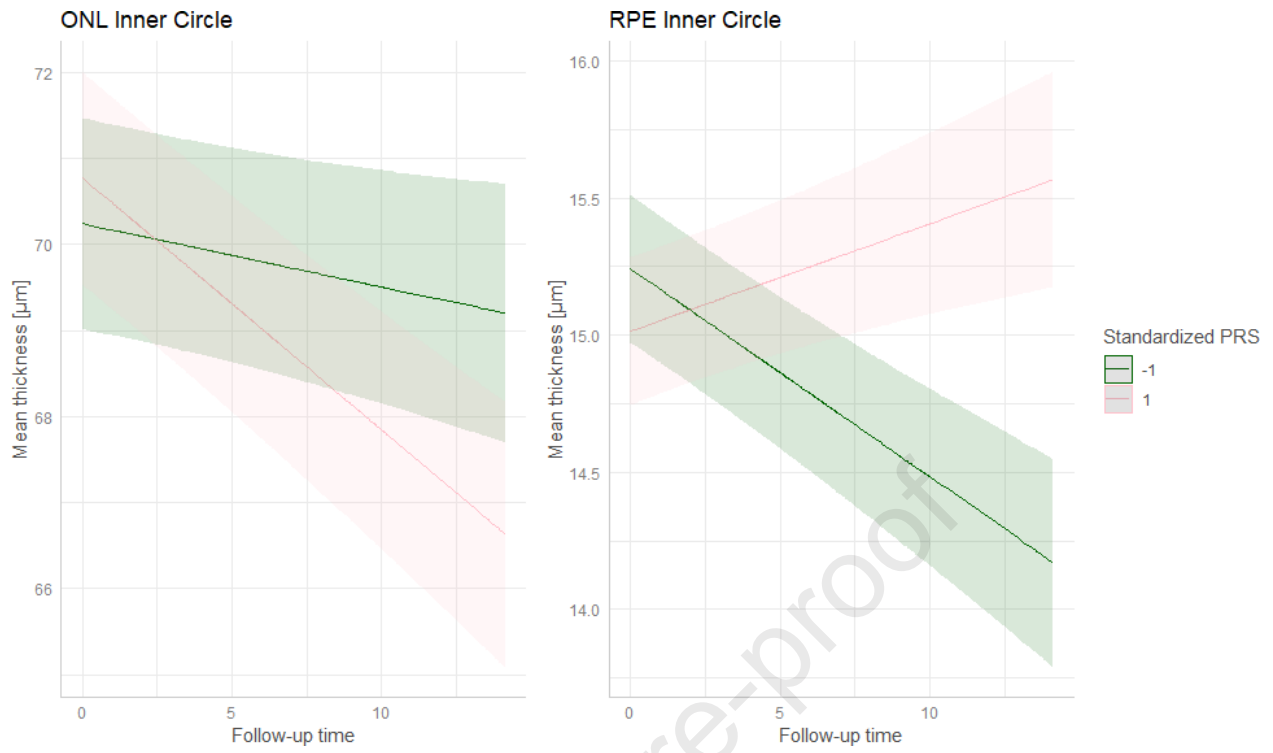
****The Lipid Pathway is based on the following genes: ABCA1, APOE, CETP, LIPC.



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Précis

A strong association of RPE-BM complex thickening with both incident AMD and the genetic risk for AMD was evidenced, highlighting its pivotal role in the clinical course of AMD.

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